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**FED - D0000 - INITIAL COMMENTS**

**Title** INITIAL COMMENTS

**CFR**

**Type** Memo Tag

**Regulation Definition**

**Interpretive Guideline**

**FED - D1000 - CERTIFICATE OF WAIVER TESTS**

**Title** CERTIFICATE OF WAIVER TESTS

**CFR** 493.15(c)

**Type** Standard

**Regulation Definition**

Certificate of waiver tests. A laboratory may qualify for a certificate of waiver under section 353 of the PHS Act if it restricts the tests that it performs to one or more of the following tests or examinations (or additional tests added to this list as provided under paragraph (d) of this section) and no others:

(1) Dipstick or Tablet Reagent Urinalysis (non-automated) for the following:

- (i) Bilirubin;
- (ii) Glucose;
- (iii) Hemoglobin;
- (iv) Ketone;
- (v) Leukocytes;
- (vi) Nitrite;
- (vii) pH;
- (viii) Protein;
- (ix) Specific gravity; and

**Interpretive Guideline**

Cite D1000 on the Form CMS-2567 and solicit a Plan of Correction when a laboratory has failed to obtain a registration, accreditation or compliance certificate before performing and reporting patient results for tests not categorized as waived. To determine which tests are categorized as waived or nonwaived (i.e., moderate or high complexity tests), refer to the following web link for the FDA categorization database (<http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfCLIA/Search.cfm?sAN=0>). Test systems, assays, and examinations not yet classified are considered high complexity. Test systems, assays and examinations that are waived, but are used in a manner that is inconsistent with manufacturer's instructions are also considered high complexity. Significant deficiencies cited under this condition may also indicate deficiencies under personnel responsibilities.

Notify the RO of a possible action by the OIG if the laboratory does not obtain the appropriate certificate or cease nonwaived testing.

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- (x) Urobilinogen.
- (2) Fecal occult blood;
- (3) Ovulation tests-visual color comparison tests for human luteinizing hormone;
- (4) Urine pregnancy tests - visual color comparison tests;
- (5) Erythrocyte sedimentation rate-non-automated;
- (6) Hemoglobin-copper sulfate-non-automated;
- (7) Blood glucose by glucose monitoring devices cleared by the FDA specifically for home use;
- (8) Spun microhematocrit; and
- (9) Hemoglobin by single analyte instruments with self-contained or component features to perform specimen/reagent interaction, providing direct measurement and readout.

**FED - D1001 - CERTIFICATE OF WAIVER TESTS**

**Title** CERTIFICATE OF WAIVER TESTS

**CFR** 493.15(e)

**Type** Standard

**Regulation Definition**

Laboratories eligible for a certificate of waiver must--

- (1) Follow manufacturers' instructions for performing the test; and
- (2) Meet the requirements in subpart B, Certificate of Waiver, of this part.

**Interpretive Guideline**

Tests listed on the waiver list in §493.15(c) are not subject to routine survey. A survey of waived tests may be conducted only when authorized by the RO in the following instances:

- o Determine if a laboratory is testing outside its certificate;
- o Collect information regarding the appropriateness of tests specified as waived tests
- o Investigate a complaint from the public; and/or
- o Determine if the laboratory is operated and if testing is performed in a manner that does not constitute an imminent and serious risk to public health.

Refer to §§493.1773 and 493.1775 for additional guidelines for inspecting laboratories issued a certificate of waiver.

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Laboratories holding a Certificate of Waiver must follow the current manufacturer's instructions for using the waived test systems they are using for patient testing. To meet the waived testing regulatory requirements, these laboratories must comply with the manufacturer's requirements. We encourage laboratories to also comply with the manufacturer's recommendations for testing. These laboratories may only use the specimen types that were approved by the Food and Drug Administration (FDA) with the waived test system they are using, and they must follow the manufacturer's quality control (QC) and test performance requirements. We encourage laboratories to also comply with manufacturer's recommendations for the waived test system. Some manufacturers produce tests that can be run as a waived test or a moderate complexity test. Any laboratory with a Certificate of Waiver that uses the nonwaived test system instructions from a manufacturer should be advised that they must use the manufacturer's instructions for waived testing. If the situation remains uncorrected, the laboratory may be cited for performing tests beyond the scope of the certificate held by the laboratory, as well as failing to follow manufacturer's instructions. See S&C-04-05.

NOTE: It is never acceptable for a laboratory operating under a Certificate of Waiver to modify the manufacturer's instructions for the waived test system. Any such change will result in a test that is no longer waived (i.e., the waived test is uncategorized for CLIA and therefore a high complexity test). For example, if a test specifies urine as the waived specimen type and the laboratory tests a different body fluid, then the laboratory is no longer performing a waived test and the laboratory is then subject to inspections and the CLIA requirements for high complexity testing. Waived laboratory testing personnel must follow the manufacturer's instructions in their entirety and without variation. Great care should be taken to add the proper reagents in the order and amount specified by the manufacturer's instructions to ensure compliance with the CLIA regulations and reliable test results.

**FED - D2000 - ENROLLMENT AND TESTING OF SAMPLES**

**Title** ENROLLMENT AND TESTING OF SAMPLES

**CFR** 493.801

**Type** Condition

**Regulation Definition**

Each laboratory must enroll in a proficiency testing (PT) program that meets the criteria in subpart I of this part and is approved by HHS. The laboratory must enroll in an approved program or programs for each of the specialties and subspecialties for which it seeks certification. The laboratory must test the samples in the same manner as patients' specimens. For laboratories subject to 42 CFR part 493

**Interpretive Guideline**

Each laboratory must determine the extent of patient testing it performs. The laboratory must review the specialty, subspecialties and analytes listed in Subpart I and determine which specialty, subspecialties and analytes they must enroll in to meet this requirement. Enrollment must be in a CMS-approved PT program that offers modules containing at least three (3) testing events annually (excluding mycobacteriology, which only needs to contain two (2) testing events annually) with a minimum of five (5) samples per event (§§493.909 - 493.459). The surveyor should verify that the laboratory is properly enrolled in an approved PT program.

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published on March 14, 1990 (55 FR 9538) prior to September 1, 1992, the rules of this subpart are effective on September 1, 1992. For all other laboratories, the rules of this subpart are effective January 1, 1994.

NOTE: If a laboratory has not enrolled for one or more tests that it performs and the tests are listed in Subpart I, cite ONLY D2000, Enrollment and testing of samples; do not cite D2016, Successful Participation.

PT requirements apply to the nonwaived tests listed in Subpart I, except for PT referral which applies to PT for all testing (waived, nonwaived, tests listed in Subpart I and tests not listed in Subpart I).

PT enrollment and participation is required, as applicable, for each certificate other than a Certificate of Waiver. A facility offering testing at more than one site, with testing included under one certificate, must enroll in an approved PT program(s) for the collective tests covered under that certificate, not for each site.

A general rule is "PT enrollment per certificate."

Facilities that perform laboratory testing at multiple sites and are certified under one CLIA certificate include the following examples:

- o A hospital with satellite laboratories throughout the hospital;
- o Different departments of the laboratory;
- o A hospital that performs point-of-care testing;
- o Limited public health testing performed by non-profit or Federal, State or local government laboratories; or
- o Mobile laboratories or temporary testing sites.

The following examples give instruction and guidance for determining compliance with the PT requirement for enrollment where a specialty, subspecialty or analyte is performed by different methods, specimen types and locations:

- o A laboratory with a single certificate must enroll in an approved PT program for each analyte listed in Subpart I that it performs. When an analyte is performed using different methodologies within the laboratory, only one PT enrollment is required. After the laboratory has determined which analyte to enroll for, it must participate in PT using its primary method for patient testing during the event. Other methods for the same analyte must be evaluated as required in §493.1236. If the laboratory performs unsuccessfully for an analyte and sanctions are imposed, the sanctions are applicable to the analyte, not to the test methodology. For example, if a laboratory uses three different

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methods to perform cholesterol measurements, it must participate in PT using the primary method at the time of the PT event. If the laboratory is unsuccessful in PT performance for cholesterol and the CLIA certificate is limited for cholesterol, the laboratory would be precluded from performing cholesterol by any test method.

- o A multisite laboratory that performs testing at the various sites under a single certificate must participate in PT for each analyte listed in Subpart I that is under that certificate. The performance of PT testing events may be rotated between different sites, provided the primary method at the time of the PT event is used to perform the PT. All samples from the testing event must be evaluated at a single site. Should the facility not perform successfully for an analyte, that analyte may not be tested at any location under that certificate.
- o A laboratory with multiple sites covered by a single certificate that participates in one PT program per analyte, must be aware that a failure in PT could lead to the limitation or revocation of its certificate for all sites for the failed analyte, subspecialty, or specialty, not just the one participating in PT.

When problems occur that cannot be resolved with the instructions in these guidelines, gather all information available and consult with the RO for guidance and resolution.

**FED - D2001 - ENROLLMENT**

**Title** ENROLLMENT

**CFR** 493.801(a)(1)(2)(i)

**Type** Standard

**Regulation Definition**

The laboratory must--  
(1) Notify HHS of the approved program or programs in which it chooses to participate to meet proficiency testing requirements of this subpart.  
(2)(i) Designate the program(s) to be used for each specialty, subspecialty, and analyte or test to determine compliance with this subpart if the laboratory participates in more than one proficiency testing program approved by CMS;

**Interpretive Guideline**

For late enrollment, refer to Laboratory Director Responsibilities (D6015 Moderate Complexity or D6088 High Complexity).

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**FED - D2003 - ENROLLMENT**

**Title** ENROLLMENT

**CFR** 493.801(a)(2)(ii)

**Type** Standard

**Regulation Definition**

For those tests performed by the laboratory that are not included in subpart I of this part, a laboratory must establish and maintain the accuracy of its testing procedures, in accordance with §493.1236(c)(1)

**Interpretive Guideline**

During the on-site survey, verify that the laboratory is enrolled in an approved program or programs for all specialties, subspecialties, analytes, or tests listed in Subpart I for which it performs patient testing.

To meet the requirements of this section, it may be necessary for a laboratory to enroll in more than one program to cover all tests listed in Subpart I for which the laboratory performs testing. The approved program in which a laboratory has enrolled may not offer every analyte that the laboratory performs. The laboratory must then enroll in an additional program(s) to cover the testing not included in the first program.

The laboratory must indicate to the PT program which specialty, subspecialty, analyte, or test it intends the program to grade and score for regulatory purposes. This is particularly necessary when the laboratory subscribes to multiple PT modules that contain the same analyte(s).

**FED - D2004 - ENROLLMENT**

**Title** ENROLLMENT

**CFR** 493.801(a)(3)

**Type** Standard

**Regulation Definition**

For each specialty, subspecialty and analyte or test, participate in one approved proficiency testing program or programs, for one year before designating a different program and must notify CMS before any change in designation;

**Interpretive Guideline**

When a laboratory initially applies for CLIA certification or adds a specialty or subspecialty in the middle of the calendar year, it may change PT programs at the next enrollment period instead of having to wait until a full year has passed. Otherwise, laboratories may not change programs after they have enrolled and participated in a PT program for a given calendar year.

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**FED - D2005 - ENROLLMENT**

**Title** ENROLLMENT

**CFR** 493.801(a)(4)

**Type** Standard

**Regulation Definition**

Authorize the proficiency testing program to release to HHS all data required to--

- (i) Determine the laboratory's compliance with this subpart; and
- (ii) Make PT results available to the public as required in section 353(f)(3)(F) of the Public Health Service Act.

**Interpretive Guideline**

The laboratory director authorizes PT data to be released to regulatory agencies when he/she signs the CLIA application for certification. The laboratory should also provide the PT program with the appropriate accreditation organization or Federal or State Agency address to which PT results must be sent. Laboratories that are accredited by a CMS- approved accreditation organization must meet the PT requirements in subpart H of the CLIA regulations, including, but not limited to, releasing all required PT data to its accreditation organization (§493.551(b)(3)).

All CLIA-exempt laboratories must enroll and participate in a CMS-approved program(s) for all analytes performed that are listed in Subpart I.

**FED - D2006 - TESTING OF PROFICIENCY TESTING SAMPLES**

**Title** TESTING OF PROFICIENCY TESTING SAMPLES

**CFR** 493.801(b)

**Type** Standard

**Regulation Definition**

The laboratory must examine or test, as applicable, the proficiency testing samples it receives from the proficiency testing program in the same manner as it tests patient specimens. This testing must be conducted in conformance with paragraph (b)(4) of this section. If the laboratory's patient specimen testing procedures would normally require reflex, distributive, or confirmatory testing at another laboratory, the laboratory should test the proficiency testing sample as it would a patient specimen up until the point it would refer a patient specimen to a second laboratory for any

**Interpretive Guideline**

Review testing records to determine if special handling was given to PT samples. Consider the unique requirements of many PT samples when evaluating "same manner" of testing. The laboratory should document any necessary reconstitution, longer mixing times, unit conversion of results, etc., as required in §493.801(b)(5).

A laboratory that routinely performs only presumptive testing or screening methods and refers patient samples to another laboratory for definitive or confirmatory testing or comparison of test results must not refer PT samples to another laboratory for confirmatory testing. A laboratory should limit the testing of PT specimens to that which is done in-house. With the exception of specimen preparation such as Immunohistochemistry (IHC) staining, laboratories need to take great care to avoid sending PT specimens or results to any entity other than their PT provider prior to the PT testing event cutoff date.

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form of further testing.

A central laboratory with more than one instrument or methodology for the same test may alternate methods or instruments from one testing event to the next as long as both are routinely used to test patient specimens. All samples for one analyte within a shipment must be tested with the same instrument.

Probes §493.801(b)

- o What procedure or test method was used?
- o Is this a routine test method used in the laboratory?
- o Did routine personnel perform the PT?
- o How often were PT samples tested? Does this conform with the laboratory's written policies for patient specimens?
- o How are deviations from general laboratory practices (if any) justified?
- o Do the PT results documented in the laboratory work records (worksheet) correlate with the results reported to the PT program?
- o Do reports submitted to the PT program provider accurately reflect the procedure (i.e., instrument, method) used in the laboratory?

Check to see if patient samples were reported on the same day that PT samples were tested. (In a small facility, infrequent testing may necessitate the testing of PT samples without patient specimens to ensure that the PT test results are returned on time.) Did the laboratory use the same procedure for both patient specimens and PT samples?

**FED - D2007 - TESTING OF PROFICIENCY TESTING SAMPLES**

**Title** TESTING OF PROFICIENCY TESTING SAMPLES

**CFR** 493.801(b)(1)

**Type** Standard

**Regulation Definition**

The samples must be examined or tested with the laboratory's regular patient workload by personnel who routinely perform the testing in the laboratory, using the laboratory's routine methods

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**FED - D2009 - TESTING OF PROFICIENCY TESTING SAMPLES**

**Title** TESTING OF PROFICIENCY TESTING SAMPLES

**CFR** 493.801(b)(1)

**Type** Standard

**Regulation Definition**

The individual testing or examining the samples and the laboratory director must attest to the routine integration of the samples into the patient workload using the laboratory's routine methods.

**Interpretive Guideline**

This requirement is NOT to be interpreted as prohibiting more than one testing individual from performing PT if the laboratory routinely performs patient testing using more than one "individual". PT samples are to be tested in the same manner as patient specimens. IF patient specimens are tested using procedures that require more than one individual to perform, PT must be performed in the same manner.

Review records to ensure that the analyst or analysts performing the testing and the director or his/her designee have signed a statement attesting that PT samples were tested in the same manner as patient specimens. For moderate complexity testing, in accordance with §493.1407(e)(4)(i), the director may delegate the responsibility for signing the attestation statement to a technical consultant meeting the qualifications of §493.1409. For high complexity testing, in accordance with §493.1445(e)(4)(i), the director may delegate the responsibility for signing the attestation statement to a technical supervisor meeting the qualifications of §493.1447.

**FED - D2010 - TESTING OF PROFICIENCY TESTING SAMPLES**

**Title** TESTING OF PROFICIENCY TESTING SAMPLES

**CFR** 493.801(b)(2)

**Type** Standard

**Regulation Definition**

The laboratory must test samples the same number of times that it routinely tests patient samples.

**Interpretive Guideline**

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**FED - D2011 - TESTING OF PROFICIENCY TESTING SAMPLES**

**Title** TESTING OF PROFICIENCY TESTING SAMPLES

**CFR** 493.801(b)(3)

**Type** Standard

**Regulation Definition**

Laboratories that perform tests on proficiency testing samples must not engage in any inter-laboratory communications pertaining to the results of proficiency testing sample(s) until after the date by which the laboratory must report proficiency testing results to the program for the testing event in which the samples were sent.

Laboratories with multiple testing sites or separate locations must not participate in any communications or discussions across sites/locations concerning proficiency testing sample results until after the date by which the laboratory must report proficiency testing results to the program.

**Interpretive Guideline**

Handle allegations of inter-laboratory communications or referral of proficiency testing specimens as a complaint and investigate using the complaint investigation procedures outlined in §6136 of the SOM. Immediately contact the RO if you find evidence to support these kinds of allegations.

**FED - D2013 - TESTING OF PROFICIENCY TESTING SAMPLES**

**Title** TESTING OF PROFICIENCY TESTING SAMPLES

**CFR** 493.801(b)(4)

**Type** Standard

**Regulation Definition**

The laboratory must not send proficiency testing samples or portions of proficiency testing samples to another laboratory for any analysis for which it is certified to perform in its own laboratory. Any laboratory that CMS determines intentionally referred a proficiency testing sample to another laboratory for analysis may have its certification revoked for at least one

**Interpretive Guideline**

The regulation refers to referral of PT specimens to another laboratory for analysis.

For those tests not listed under Subpart I (not regulated), the laboratory is free to enroll in a PT program to verify the accuracy of their test or procedure. Due to the breadth of the statutory bar on PT sample referrals, however, laboratories should take great measures to avoid sending any such PT samples (or test results) to another laboratory for any reason prior to the PT testing event cutoff date. The PT referral consequences (loss of certificate and bar on

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year. If CMS determines that a proficiency testing sample was referred to another laboratory for analysis, but the requested testing was limited to reflex, distributive, or confirmatory testing that, if the sample were a patient specimen, would have been in full conformance with written, legally accurate and adequate standard operating procedures for the laboratory's testing of patient specimens, and if the proficiency testing referral is not a repeat proficiency testing referral, CMS will consider the referral to be improper and subject to alternative sanctions in accordance with §493.1804(c), but not intentional. Any laboratory that receives a proficiency testing sample from another laboratory for testing must notify CMS of the receipt of that sample regardless of whether the referral was made for reflex or confirmatory testing, or any other reason.

owner operator) apply equally to all PT testing samples and results.

Do not solicit a Plan of Correction from a laboratory when it has been determined that the laboratory intentionally referred its PT samples to another laboratory for analysis. Immediately notify the RO recommending revocation of the certificate (a statutory requirement) and forward to the RO all documentation necessary to support the findings.

**FED - D2015 - TESTING OF PROFICIENCY TESTING SAMPLES**

**Title** TESTING OF PROFICIENCY TESTING SAMPLES

**CFR** 493.801(b)(5)(6)

**Type** Standard

**Regulation Definition**

(5) The laboratory must document the handling, preparation, processing, examination, and each step in the testing and reporting of results for all proficiency testing samples. The laboratory must maintain a copy of all records, including a copy of the proficiency testing program report forms used by the laboratory to record proficiency testing results including the attestation statement provided by the PT program, signed by the analyst and the laboratory director, documenting that proficiency testing samples were tested in the same manner as patient specimens, for a minimum of two years from the date of the proficiency testing event.

(6) PT is required for only the test system, assay, or examination used as the primary method for patient testing

**Interpretive Guideline**

Interpretative Guidelines §493.801(b)(5)

Review records to ensure that the analyst or analysts performing the testing and the director or his/her designee have signed the statement attesting that PT samples were tested in the same manner as patient specimens. For moderate complexity testing, in accordance with §493.1407(e)(4)(i), the director may delegate the responsibility for signing the attestation statement to a technical consultant meeting the qualifications of §493.1409. For high complexity testing, in accordance with §493.1445(e)(4)(i), the director may delegate the responsibility for signing the attestation statement to a technical supervisor meeting the qualifications of §493.1447. The signature of the director or technical consultant/supervisor need not be obtained prior to reporting PT results to the PT provider.

Interpretative Guidelines §493.801(b)(6)

Primary means the test system(s), assay(s) or examination(s) routinely used for patient testing at the time of the PT

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during the PT event.

testing event.

**FED - D2016 - SUCCESSFUL PARTICIPATION**

**Title** SUCCESSFUL PARTICIPATION

**CFR** 493.803(a)(b)(c)

**Type** Condition

**Regulation Definition**

- (a) Each laboratory performing nonwaived testing must successfully participate in a proficiency testing program approved by CMS, if applicable, as described in subpart I of this part for each specialty, subspecialty, and analyte or test in which the laboratory is certified under CLIA.
- (b) Except as specified in paragraph (c) of this section, if a laboratory fails to participate successfully in proficiency testing for a given specialty, subspecialty, analyte or test, as defined in this section, or fails to take remedial action when an individual fails gynecologic cytology, CMS imposes sanctions, as specified in subpart R of this part.
- (c) If a laboratory fails to perform successfully in a CMS-approved proficiency testing program, for the initial unsuccessful performance, CMS may direct the laboratory to undertake training of its personnel or to obtain technical assistance, or both, rather than imposing alternative or principle sanctions except when one or more of the following conditions exists:
- (1) There is immediate jeopardy to patient health and safety.
  - (2) The laboratory fails to provide CMS or a CMS agent with satisfactory evidence that it has taken steps to correct the problem identified by the unsuccessful proficiency testing performance.
  - (3) The laboratory has a poor compliance history.

**Interpretive Guideline**

Only the PT program has the capability to correct scores in the CMS PT monitoring system.

No single PT enforcement protocol is universally applicable for all situations. Unique circumstances may require special considerations or actions that may not conform to the general approach outlined below. The laboratory's compliance history, its willingness to take remedial actions, and the professional judgment of surveyors, RO CLIA laboratory consultants and enforcement personnel may be factors in determining an appropriate PT enforcement plan.

Careful review of PT performance reports and other available information should always be performed to determine whether the PT results truly represent failed PT. The potential of a PT program data input error or other factors beyond the laboratory's control should be considered. If the laboratory has made a transcription error(s), it is considered erroneous PT result(s).

If review and verification of PT performance reports confirm unsuccessful PT, cite as a Condition-level deficiency (use D2016 on the CMS-2567).

NOTE: The CMS PT monitoring system may NOT be used alone to determine unsuccessful participation. Surveyors must verify any unsuccessful participation indicated in the PT monitoring system. This may be done by reviewing PT results supplied by the approved PT program (they will send copies to the surveyor if requested) or from results sent to the laboratory by the PT program.

If the unsuccessful PT participation is the first occurrence for the laboratory, and none of the exceptions listed at §493.803(c)(1-3) exist, notify the laboratory and instruct them to seek training of its personnel, obtain the necessary technical assistance to correct the problem causing the unsuccessful participation, or both. SAs may initiate training and/or technical assistance after first obtaining RO concurrence. No on-site review is required to initiate this action.

The laboratory must submit an acceptable plan of remedial action, listing projected completion dates and other pertinent information for its training and/or technical assistance efforts. Follow-up is necessary to verify that the

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laboratory has carried out its plan. Satisfactory participation in the next PT event would provide verification that the laboratory's remedial action, training and/or technical assistance were successful. The remedial action plan should demonstrate that the laboratory will correct its problems within 3 months, although special circumstances may be considered. If a laboratory refuses to take acceptable training and/or technical assistance actions (including failure to submit an acceptable plan of remedial action, or failure to complete its plan), sanction action may be initiated with concurrence from the RO.

When the unsuccessful PT participation is not the first such occurrence for the laboratory, cite as a condition-level deficiency and take appropriate enforcement action. For immediate jeopardy cases the procedures in Subpart R apply. For non-immediate jeopardy situations, enforcement procedures should be completed within 90 days from the date that the unsuccessful PT was first identified. In immediate jeopardy situations, enforcement procedures should be completed within 23 days from the date unsuccessful participation of PT is first identified.

Example:

A laboratory scores 60% on a testing event in mycobacteriology. On the next testing event, the laboratory fails to participate in mycobacteriology. The citations are D2030 (§493.825), D2037 (§493.825) and D2016 (§493.803). (Note: It is not necessary to cite the standard for unsatisfactory analyte performance. However, it is necessary to cite the standard when the laboratory fails to participate in a testing event so that the laboratory is made aware that such deficient practice results in a score of 0 for the testing event.)

Example:

A laboratory scores 60% on uric acid PT samples. On the next testing event, the laboratory scores 40% on the same analyte. The citations are §§493.841(f), and 493.803. (Note: Cite the standard for unsuccessful performance and the condition for unsuccessful participation. It is not necessary to cite the standard for unsatisfactory analyte performance.)

When recommending to the RO that a laboratory be subject to sanctions, submit copies of the laboratory's testing event or analyte score(s) that were unsatisfactory and the correct responses provided by the PT program. Also, enclose copies of any correspondence sent to or received by the laboratory concerning its PT performance.

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**FED - D2017 - REINSTATEMENT OF NONWAIVED LABORATORIES**

**Title** REINSTATEMENT OF NONWAIVED LABORATORIES

**CFR** 493.807(a)(b)

**Type** Condition

**Regulation Definition**

(a) If a laboratory's certificate is suspended or limited or its Medicare or Medicaid approval is cancelled or its Medicare or Medicaid payments are suspended because it fails to participate successfully in proficiency testing for one or more specialties, subspecialties, analyte or test, or voluntarily withdraws its certification under CLIA for the failed specialty, subspecialty, or analyte, the laboratory must then demonstrate sustained satisfactory performance on two consecutive proficiency testing events, one of which may be on site, before CMS will consider it for reinstatement for certification and Medicare or Medicaid approval in that specialty, subspecialty, analyte or test.

(b) The cancellation period for Medicare and Medicaid approval or period for suspension of Medicare or Medicaid payments or suspension or limitation of certification under CLIA for the failed specialty, subspecialty, or analyte or test is for a period of not less than six months from the date of cancellation, limitation or suspension of the CLIA certificate.

**Interpretive Guideline**

The surveyor may review Report #155 of the PT monitoring system to determine whether the laboratory has performed two consecutive PT events successfully. These data are identified as "non-routine" scores in the system. The PT program supplying the re-instatement samples will grade the events and enter the scores in the system.

**FED - D2020 - BACTERIOLOGY**

**Title** BACTERIOLOGY

**CFR** 493.823(a)

**Type** Standard

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**Regulation Definition**

**Interpretive Guideline**

Failure to attain an overall testing event score of at least 80 percent is unsatisfactory performance.

**FED - D2021 - BACTERIOLOGY**

**Title** BACTERIOLOGY

**CFR** 493.823(b)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to participate in a testing event is unsatisfactory performance and results in a score of 0 for the testing event. Consideration may be given to those laboratories failing to participate in a testing event only if--

- (1) Patient testing was suspended during the time frame allotted for testing and reporting proficiency testing results;
- (2) The laboratory notifies the inspecting agency and the proficiency testing program within the time frame for submitting proficiency testing results of the suspension of patient testing and the circumstances associated with failure to perform tests on proficiency testing samples; and
- (3) The laboratory participated in the previous two proficiency testing events.

**FED - D2025 - BACTERIOLOGY**

**Title** BACTERIOLOGY

**CFR** 493.823(c)

**Type** Standard

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**Regulation Definition**

Failure to return proficiency testing results to the proficiency testing program within the time frame specified by the program is unsatisfactory performance and results in a score of 0 for the testing event.

**Interpretive Guideline**

**FED - D2026 - BACTERIOLOGY**

**Title** BACTERIOLOGY

**CFR** 493.823(d)

**Type** Standard

**Regulation Definition**

(1) For any unsatisfactory testing event for reasons other than a failure to participate, the laboratory must undertake appropriate training and employ the technical assistance necessary to correct problems associated with a proficiency testing failure.

(2) Remedial action must be taken and documented, and the documentation must be maintained by the laboratory for two years from the date of participation in the proficiency testing event.

**Interpretive Guideline**

**FED - D2028 - BACTERIOLOGY**

**Title** BACTERIOLOGY

**CFR** 493.823(e)

**Type** Standard

**Regulation Definition**

Failure to achieve an overall testing event score of satisfactory performance for two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

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**FED - D2029 - MYCOBACTERIOLOGY**

**Title** MYCOBACTERIOLOGY

**CFR** 493.825(a)

**Type** Standard

**Regulation Definition**

Failure to attain an overall testing event score of at least 80 percent is unsatisfactory performance.

**Interpretive Guideline**

**FED - D2030 - MYCOBACTERIOLOGY**

**Title** MYCOBACTERIOLOGY

**CFR** 493.825(b)

**Type** Standard

**Regulation Definition**

Failure to participate in a testing event is unsatisfactory performance and results in a score of 0 for the testing event. Consideration may be given to those laboratories failing to participate in a testing event only if--

- (1) Patient testing was suspended during the time frame allotted for testing and reporting proficiency testing results;
- (2) The laboratory notifies the inspecting agency and the proficiency testing program within the time frame for submitting proficiency testing results of the suspension of patient testing and the circumstances associated with failure to perform tests on proficiency testing samples; and
- (3) The laboratory participated in the previous two proficiency testing events.

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**FED - D2034 - MYCOBACTERIOLOGY**

**Title** MYCOBACTERIOLOGY

**CFR** 483.825(c)

**Type** Standard

**Regulation Definition**

Failure to return proficiency testing results to the proficiency testing program within the time frame specified by the program is unsatisfactory performance and results in a score of 0 for the testing event.

**Interpretive Guideline**

**FED - D2035 - MYCOBACTERIOLOGY**

**Title** MYCOBACTERIOLOGY

**CFR** 493.825(d)

**Type** Standard

**Regulation Definition**

(1) For any unsatisfactory testing event for reasons other than a failure to participate, the laboratory must undertake appropriate training and employ the technical assistance necessary to correct problems associated with a proficiency testing failure.

(2) Remedial action must be taken and documented, and the documentation must be maintained by the laboratory for two years from the date of participation in the proficiency testing event.

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FED - D2037 - MYCOBACTERIOLOGY

**Title** MYCOBACTERIOLOGY

**CFR** 493.825(e)

**Type** Standard

**Regulation Definition**

Failure to achieve an overall testing event score of satisfactory performance for two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

**Interpretive Guideline**

FED - D2038 - MYCOLOGY

**Title** MYCOLOGY

**CFR** 493.827(a)

**Type** Standard

**Regulation Definition**

Failure to attain an overall testing event score of at least 80 percent is unsatisfactory performance.

**Interpretive Guideline**

FED - D2039 - MYCOLOGY

**Title** MYCOLOGY

**CFR** 493.827(b)

**Type** Standard

**Regulation Definition**

Failure to participate in a testing event is unsatisfactory performance and results in a score of 0 for the testing event. Consideration may be given to those laboratories failing to

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participate in a testing event only if--

- (1) Patient testing was suspended during the time frame allotted for testing and reporting proficiency testing results;
- (2) The laboratory notifies the inspecting agency and the proficiency testing program within the time frame for submitting proficiency testing results of the suspension of patient testing and the circumstances associated with failure to perform tests on proficiency testing samples; and
- (3) The laboratory participated in the previous two proficiency testing events.

**FED - D2043 - MYCOLOGY**

**Title** MYCOLOGY

**CFR** 493.827(c)

**Type** Standard

**Regulation Definition**

Failure to return proficiency testing results to the proficiency testing program within the time frame specified by the program is unsatisfactory performance and results in a score of 0 for the testing event.

**Interpretive Guideline**

**FED - D2044 - MYCOLOGY**

**Title** MYCOLOGY

**CFR** 493.827(d)

**Type** Standard

**Regulation Definition**

- (1) For any unsatisfactory testing event for reasons other than a failure to participate, the laboratory must undertake appropriate training and employ the technical assistance necessary to correct problems associated with a proficiency

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testing failure.

(2) Remedial action must be taken and documented, and the documentation must be maintained by the laboratory for two years from the date of participation in the proficiency testing event.

**FED - D2046 - MYCOLOGY**

**Title** MYCOLOGY

**CFR** 493.827(e)

**Type** Standard

**Regulation Definition**

Failure to achieve an overall testing event score of satisfactory performance for two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

**Interpretive Guideline**

**FED - D2047 - PARASITOLOGY**

**Title** PARASITOLOGY

**CFR** 493.829(a)

**Type** Standard

**Regulation Definition**

Failure to attain an overall testing event score of at least 80 percent is unsatisfactory performance.

**Interpretive Guideline**

**FED - D2048 - PARASITOLOGY**

**Title** PARASITOLOGY

**CFR** 493.829(b)

**Type** Standard

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**Regulation Definition**

Failure to participate in a testing event is unsatisfactory performance and results in a score of 0 for the testing event. Consideration may be given to those laboratories failing to participate in a testing event only if--

- (1) Patient testing was suspended during the time frame allotted for testing and reporting proficiency testing results;
- (2) The laboratory notifies the inspecting agency and the proficiency testing program within the time frame for submitting proficiency testing results of the suspension of patient testing and the circumstances associated with failure to perform tests on proficiency testing samples; and
- (3) The laboratory participated in the previous two proficiency testing events.

**Interpretive Guideline**

FED - D2052 - PARASITOLOGY

**Title** PARASITOLOGY

**CFR** 493.829(c)

**Type** Standard

**Regulation Definition**

Failure to return proficiency testing results to the proficiency testing program within the time frame specified by the program is unsatisfactory performance and results in a score of 0 for the testing event.

**Interpretive Guideline**

FED - D2053 - PARASITOLOGY

**Title** PARASITOLOGY

**CFR** 492.829(d)

**Type** Standard

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**Regulation Definition**

**Interpretive Guideline**

(1) For any unsatisfactory testing event for reasons other than a failure to participate, the laboratory must undertake appropriate training and employ the technical assistance necessary to correct problems associated with a proficiency testing failure.

(2) Remedial action must be taken and documented, and the documentation must be maintained by the laboratory for two years from the date of participation in the proficiency testing event.

**FED - D2055 - PARASITOLOGY**

**Title** PARASITOLOGY

**CFR** 493.829(e)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to achieve an overall testing event score of satisfactory performance for two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

**FED - D2056 - VIROLOGY**

**Title** VIROLOGY

**CFR** 493.831(a)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to attain an overall testing event score of at least 80 percent is unsatisfactory performance.

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**FED - D2057 - VIROLOGY**

**Title** VIROLOGY

**CFR** 493.831(b)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to participate in a testing event is unsatisfactory performance and results in a score of 0 for the testing event. Consideration may be given to those laboratories failing to participate in a testing event only if--

- (1) Patient testing was suspended during the time frame allotted for testing and reporting proficiency testing results;
- (2) The laboratory notifies the inspecting agency and the proficiency testing program within the time frame for submitting proficiency testing results of the suspension of patient testing and the circumstances associated with failure to perform tests on proficiency testing samples; and
- (3) The laboratory participated in the previous two proficiency testing events.

**FED - D2061 - VIROLOGY**

**Title** VIROLOGY

**CFR** 493.831(c)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to return proficiency testing results to the proficiency testing program within the time frame specified by the program is unsatisfactory performance and results in a score of 0 for the testing event.

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**FED - D2062 - VIROLOGY**

**Title** VIROLOGY

**CFR** 493.831(d)

**Type** Standard

**Regulation Definition**

- (1) For any unsatisfactory testing event for reasons other than a failure to participate, the laboratory must undertake appropriate training and employ the technical assistance necessary to correct problems associated with a proficiency testing failure.
- (2) For any unsatisfactory testing events, remedial action must be taken and documented, and the documentation must be maintained by the laboratory for two years from the date of participation in the proficiency testing event.

**Interpretive Guideline**

**FED - D2064 - VIROLOGY**

**Title** VIROLOGY

**CFR** 493.831(e)

**Type** Standard

**Regulation Definition**

Failure to achieve an overall testing event score of satisfactory performance for two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

**Interpretive Guideline**

Any laboratory testing patient specimens for the Human Papillomavirus (HPV) must enroll and successfully participate in a CMS-approved proficiency testing program for HPV. Laboratories should refer to Subpart H for further information. For example: A Cytology laboratory that performs HPV testing must have a CLIA certificate that includes the subspecialty of Virology.

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**FED - D2066 - SYPHILIS SEROLOGY**

**Title** SYPHILIS SEROLOGY

**CFR** 493.835(a)

**Type** Standard

**Regulation Definition**

Failure to attain an overall testing event score of at least 80 percent is unsatisfactory performance.

**Interpretive Guideline**

**FED - D2067 - SYPHILIS SEROLOGY**

**Title** SYPHILIS SEROLOGY

**CFR** 493.835(b)

**Type** Standard

**Regulation Definition**

Failure to participate in a testing event is unsatisfactory performance and results in a score of 0 for the testing event. Consideration may be given to those laboratories failing to participate in a testing event only if--

- (1) Patient testing was suspended during the time frame allotted for testing and reporting proficiency testing results;
- (2) The laboratory notifies the inspecting agency and the proficiency testing program within the time frame for submitting proficiency testing results of the suspension of patient testing and the circumstances associated with failure to perform tests on proficiency testing samples; and
- (3) The laboratory participated in the previous two proficiency testing events.

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**FED - D2071 - SYPHILIS SEROLOGY**

**Title** SYPHILIS SEROLOGY

**CFR** 493.835(c)

**Type** Standard

**Regulation Definition**

Failure to return proficiency testing results to the proficiency testing program within the time frame specified by the program is unsatisfactory performance and results in a score of 0 for the testing event.

**Interpretive Guideline**

**FED - D2072 - SYPHILIS SEROLOGY**

**Title** SYPHILIS SEROLOGY

**CFR** 493.835(d)

**Type** Standard

**Regulation Definition**

- (1) For any unsatisfactory testing event for reasons other than a failure to participate, the laboratory must undertake appropriate training and employ the technical assistance necessary to correct problems associated with a proficiency testing failure.
- (2) For any unacceptable testing event score, remedial action must be taken and documented, and the documentation must be maintained by the laboratory for two years from the date of participation in the proficiency testing event.

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**FED - D2074 - SYPHILIS SEROLOGY**

**Title** SYPHILIS SEROLOGY

**CFR** 493.835(e)

**Type** Standard

**Regulation Definition**

Failure to achieve an overall testing event score of satisfactory performance for two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

**Interpretive Guideline**

**FED - D2075 - GENERAL IMMUNOLOGY**

**Title** GENERAL IMMUNOLOGY

**CFR** 493.837(a)

**Type** Standard

**Regulation Definition**

Failure to attain a score of at least 80 percent of acceptable responses for each analyte in each testing event is unsatisfactory analyte performance for the testing event.

**Interpretive Guideline**

Refer to Subpart I for analytes or tests for which laboratory PT performance is to be evaluated.  
Note: If a laboratory performs both a quantitative and a qualitative procedure of a test or analyte, it may choose which to enroll in to fulfill the enrollment requirement. It need not enroll in both quantitative and qualitative PT for the same analyte.

**FED - D2076 - GENERAL IMMUNOLOGY**

**Title** GENERAL IMMUNOLOGY

**CFR** 493.837(b)

**Type** Standard

**Regulation Definition**

Failure to attain an overall testing event score of at least 80

**Interpretive Guideline**

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percent is unsatisfactory performance.

**FED - D2077 - GENERAL IMMUNOLOGY**

**Title** GENERAL IMMUNOLOGY

**CFR** 493.837(c)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to participate in a testing event is unsatisfactory performance and results in a score of 0 for the testing event. Consideration may be given to those laboratories failing to participate in a testing event only if--

- (1) Patient testing was suspended during the time frame allotted for testing and reporting proficiency testing results;
- (2) The laboratory notifies the inspecting agency and the proficiency testing program within the time frame for submitting proficiency testing results of the suspension of patient testing and the circumstances associated with failure to perform tests on proficiency testing samples; and
- (3) The laboratory participated in the previous two proficiency testing events.

**FED - D2081 - GENERAL IMMUNOLOGY**

**Title** GENERAL IMMUNOLOGY

**CFR** 493.837(d)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to return proficiency testing results to the proficiency testing program within the time frame specified by the program is unsatisfactory performance and results in a score of 0 for the testing event.

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**FED - D2082 - GENERAL IMMUNOLOGY**

**Title** GENERAL IMMUNOLOGY

**CFR** 493.837(e)

**Type** Standard

**Regulation Definition**

(1) For any unsatisfactory analyte or test performance or testing event for reasons other than a failure to participate, the laboratory must undertake appropriate training and employ the technical assistance necessary to correct problems associated with a proficiency testing failure.

(2) For any unacceptable analyte or testing event score, remedial action must be taken and documented, and the documentation must be maintained by the laboratory for two years from the date of participation in the proficiency testing event.

**Interpretive Guideline**

**FED - D2084 - GENERAL IMMUNOLOGY**

**Title** GENERAL IMMUNOLOGY

**CFR** 493.837(f)

**Type** Standard

**Regulation Definition**

Failure to achieve satisfactory performance for the same analyte or test in two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

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FED - D2085 - GENERAL IMMUNOLOGY

**Title** GENERAL IMMUNOLOGY

**CFR** 493.837(g)

**Type** Standard

**Regulation Definition**

Failure to achieve an overall testing event score of satisfactory performance for two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

**Interpretive Guideline**

FED - D2087 - ROUTINE CHEMISTRY

**Title** ROUTINE CHEMISTRY

**CFR** 493.841(a)

**Type** Standard

**Regulation Definition**

Failure to attain a score of at least 80 percent of acceptable responses for each analyte in each testing event is unsatisfactory analyte performance for the testing event.

**Interpretive Guideline**

Refer to Subpart I for analytes or tests for which laboratory PT performance is to be evaluated which include serum, plasma or blood samples.

FED - D2088 - ROUTINE CHEMISTRY

**Title** ROUTINE CHEMISTRY

**CFR** 493.841(b)

**Type** Standard

**Regulation Definition**

Failure to attain an overall testing event score of at least 80 percent is unsatisfactory performance.

**Interpretive Guideline**

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**FED - D2089 - ROUTINE CHEMISTRY**

**Title** ROUTINE CHEMISTRY

**CFR** 493.841(c)

**Type** Standard

**Regulation Definition**

Failure to participate in a testing event is unsatisfactory performance and results in a score of 0 for the testing event. Consideration may be given to those laboratories failing to participate in a testing event only if--

- (1) Patient testing was suspended during the time frame allotted for testing and reporting proficiency testing results;
- (2) The laboratory notifies the inspecting agency and the proficiency testing program within the time frame for submitting proficiency testing results of the suspension of patient testing and the circumstances associated with failure to perform tests on proficiency testing samples; and
- (3) The laboratory participated in the previous two proficiency testing events.

**Interpretive Guideline**

**FED - D2093 - ROUTINE CHEMISTRY**

**Title** ROUTINE CHEMISTRY

**CFR** 493.841(d)

**Type** Standard

**Regulation Definition**

Failure to return proficiency testing results to the proficiency testing program within the time frame specified by the program is unsatisfactory performance and results in a score of 0 for the testing event.

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**FED - D2094 - ROUTINE CHEMISTRY**

**Title** ROUTINE CHEMISTRY

**CFR** 493.841(e)

**Type** Standard

**Regulation Definition**

(1) For any unsatisfactory analyte or test performance or testing event for reasons other than a failure to participate, the laboratory must undertake appropriate training and employ the technical assistance necessary to correct problems associated with a proficiency testing failure.

(2) For any unacceptable analyte or testing event score, remedial action must be taken and documented, and the documentation must be maintained by the laboratory for two years from the date of participation in the proficiency testing event.

**Interpretive Guideline**

**FED - D2096 - ROUTINE CHEMISTRY**

**Title** ROUTINE CHEMISTRY

**CFR** 493.841(f)

**Type** Standard

**Regulation Definition**

Failure to achieve satisfactory performance for the same analyte or test in two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

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FED - D2097 - ROUTINE CHEMISTRY

**Title** ROUTINE CHEMISTRY

**CFR** 493.841(g)

**Type** Standard

**Regulation Definition**

Failure to achieve an overall testing event score of satisfactory performance for two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

**Interpretive Guideline**

FED - D2098 - ENDOCRINOLOGY

**Title** ENDOCRINOLOGY

**CFR** 493.843(a)

**Type** Standard

**Regulation Definition**

Failure to attain a score of at least 80 percent of acceptable responses for each analyte in each testing event is unsatisfactory analyte performance for the testing event.

**Interpretive Guideline**

Refer to Subpart I for analytes or tests for which laboratory PT performance is to be evaluated which include serum, plasma, blood, or urine.  
Note: If the laboratory performs the same analyte on different specimen types, it may choose which specimen type to enroll in PT. The laboratory need not enroll for each specimen type of the same analyte.

FED - D2099 - ENDOCRINOLOGY

**Title** ENDOCRINOLOGY

**CFR** 493.843(b)

**Type** Standard

**Regulation Definition**

Failure to attain an overall testing event score of at least 80

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percent is unsatisfactory performance.

**FED - D2100 - ENDOCRINOLOGY**

**Title** ENDOCRINOLOGY

**CFR** 493.843(c)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to participate in a testing event is unsatisfactory performance and results in a score of 0 for the testing event. Consideration may be given to those laboratories failing to participate in a testing event only if--

- (1) Patient testing was suspended during the time frame allotted for testing and reporting proficiency testing results;
- (2) The laboratory notifies the inspecting agency and the proficiency testing program within the time frame for submitting proficiency testing results of the suspension of patient testing and the circumstances associated with failure to perform tests on proficiency testing samples; and
- (3) The laboratory participated in the previous two proficiency testing events.

**FED - D2104 - ENDOCRINOLOGY**

**Title** ENDOCRINOLOGY

**CFR** 493.843(d)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to return proficiency testing results to the proficiency testing program within the time frame specified by the program is unsatisfactory performance and results in a score of 0 for the testing event.

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**FED - D2105 - ENDOCRINOLOGY**

**Title** ENDOCRINOLOGY

**CFR** 493.843(e)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

(1) For any unsatisfactory analyte or test performance or testing event for reasons other than a failure to participate, the laboratory must undertake appropriate training and employ the technical assistance necessary to correct problems associated with a proficiency testing failure.

(2) For any unacceptable analyte or testing event score, remedial action must be taken and documented, and the documentation must be maintained by the laboratory for two years from the date of participation in the proficiency testing event.

**FED - D2107 - ENDOCRINOLOGY**

**Title** ENDOCRINOLOGY

**CFR** 493.843(f)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to achieve satisfactory performance for the same analyte or test in two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

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FED - D2108 - ENDOCRINOLOGY

**Title** ENDOCRINOLOGY

**CFR** 493.843(g)

**Type** Standard

**Regulation Definition**

Failure to achieve an overall testing event score of satisfactory performance for two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

**Interpretive Guideline**

FED - D2109 - TOXICOLOGY

**Title** TOXICOLOGY

**CFR** 493.845(a)

**Type** Standard

**Regulation Definition**

Failure to attain a score of at least 80 percent of acceptable responses for each analyte in each testing event is unsatisfactory analyte performance for the testing event.

**Interpretive Guideline**

Refer to Subpart I for analytes or tests for which laboratory PT performance is to be evaluated which include serum, plasma, or blood.

FED - D2110 - TOXICOLOGY

**Title** TOXICOLOGY

**CFR** 493.845(b)

**Type** Standard

**Regulation Definition**

Failure to attain an overall testing event score of at least 80 percent is unsatisfactory performance.

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**FED - D2111 - TOXICOLOGY**

**Title** TOXICOLOGY

**CFR** 493.845(c)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to participate in a testing event is unsatisfactory performance and results in a score of 0 for the testing event. Consideration may be given to those laboratories failing to participate in a testing event only if--

- (1) Patient testing was suspended during the time frame allotted for testing and reporting proficiency testing results;
- (2) The laboratory notifies the inspecting agency and the proficiency testing program within the time frame for submitting proficiency testing results of the suspension of patient testing and the circumstances associated with failure to perform tests on proficiency testing samples; and
- (3) The laboratory participated in the previous two proficiency testing events.

**FED - D2115 - TOXICOLOGY**

**Title** TOXICOLOGY

**CFR** 493.845(d)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to return proficiency testing results to the proficiency testing program within the time frame specified by the program is unsatisfactory performance and results in a score of 0 for the testing event.

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**FED - D2116 - TOXICOLOGY**

**Title** TOXICOLOGY

**CFR** 493.845(e)

**Type** Standard

**Regulation Definition**

(1) For any unsatisfactory analyte or test performance or testing event for reasons other than a failure to participate, the laboratory must undertake appropriate training and employ the technical assistance necessary to correct problems associated with a proficiency testing failure.

(2) For any unacceptable analyte or testing event score, remedial action must be taken and documented, and the documentation must be maintained by the laboratory for two years from the date of participation in the proficiency testing event.

**Interpretive Guideline**

**FED - D2118 - TOXICOLOGY**

**Title** TOXICOLOGY

**CFR** 493.845(f)

**Type** Standard

**Regulation Definition**

Failure to achieve satisfactory performance for the same analyte or test in two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

**Interpretive Guideline**

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FED - D2119 - TOXICOLOGY

**Title** TOXICOLOGY

**CFR** 493.845(g)

**Type** Standard

**Regulation Definition**

Failure to achieve an overall testing event score of satisfactory performance for two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

**Interpretive Guideline**

FED - D2121 - HEMATOLOGY

**Title** HEMATOLOGY

**CFR** 493.851(a)

**Type** Standard

**Regulation Definition**

Failure to attain a score of at least 80 percent of acceptable responses for each analyte in each testing event is unsatisfactory analyte performance for the testing event.

**Interpretive Guideline**

Refer to Subpart I for analytes or tests for which laboratory PT performance is to be evaluated.

FED - D2122 - HEMATOLOGY

**Title** HEMATOLOGY

**CFR** 493.851(b)

**Type** Standard

**Regulation Definition**

Failure to attain an overall testing event score of at least 80 percent is unsatisfactory performance.

**Interpretive Guideline**

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**FED - D2123 - HEMATOLOGY**

**Title** HEMATOLOGY

**CFR** 493.851(c)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to participate in a testing event is unsatisfactory performance and results in a score of 0 for the testing event. Consideration may be given to those laboratories failing to participate in a testing event only if--

- (1) Patient testing was suspended during the time frame allotted for testing and reporting proficiency testing results;
- (2) The laboratory notifies the inspecting agency and the proficiency testing program within the time frame for submitting proficiency testing results of the suspension of patient testing and the circumstances associated with failure to perform tests on proficiency testing samples; and
- (3) The laboratory participated in the previous two proficiency testing events.

**FED - D2127 - HEMATOLOGY**

**Title** HEMATOLOGY

**CFR** 493.851(d)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to return proficiency testing results to the proficiency testing program within the time frame specified by the program is unsatisfactory performance and results in a score of 0 for the testing event.

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**FED - D2128 - HEMATOLOGY**

**Title** HEMATOLOGY

**CFR** 493.851(e)

**Type** Standard

**Regulation Definition**

(1) For any unsatisfactory analyte or test performance or testing event for reasons other than a failure to participate, the laboratory must undertake appropriate training and employ the technical assistance necessary to correct problems associated with a proficiency testing failure.

(2) For any unacceptable analyte or testing event score, remedial action must be taken and documented, and the documentation must be maintained by the laboratory for two years from the date of participation in the proficiency testing event.

**Interpretive Guideline**

**FED - D2130 - HEMATOLOGY**

**Title** HEMATOLOGY

**CFR** 493.851(f)

**Type** Standard

**Regulation Definition**

Failure to achieve satisfactory performance for the same analyte in two consecutive events or two out of three consecutive testing events is unsuccessful performance.

**Interpretive Guideline**

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**FED - D2131 - HEMATOLOGY**

**Title** HEMATOLOGY

**CFR** 493.851(g)

**Type** Standard

**Regulation Definition**

Failure to achieve an overall testing event score of satisfactory performance for two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

**Interpretive Guideline**

**FED - D2132 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.855

**Type** Standard

**Regulation Definition**

To participate successfully in a cytology proficiency testing program for gynecologic examinations (Pap smears), the laboratory must meet the requirements of paragraphs (a) through (c) of this section.

**Interpretive Guideline**

**FED - D2133 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.855(a)

**Type** Standard

**Regulation Definition**

The laboratory must ensure that each individual engaged in the

**Interpretive Guideline**

Confirm by review of the attestation of enrollment documentation that all the individuals examining gynecologic

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examination of gynecologic preparations is enrolled in a proficiency testing program approved by CMS by January 1, 1995, if available in the State in which he or she is employed.

cytology slides are enrolled in a CMS-approved cytology PT program.

If an individual works at more than one laboratory, the individual will be required to indicate, prior to the first testing event, one laboratory as the primary laboratory where the individual will be tested. Each laboratory is responsible for ensuring that all individuals examining gynecologic preparations in their laboratory indicate a location of testing.

Pathologists who routinely examine gynecologic cytology slide preparations, only after they have been examined and marked by a cytotechnologist, may be tested by one of two methods:

- a. Using a test that has been previously examined or marked by a cytotechnologist in their laboratory accompanied by the cytotechnologist's PT answers or
- b. Using a test set that has not been previously examined.

A pathologist, who examine and interprets slide preparations without pre-screening by a cytotechnologist, must be tested using a test set that has not been previously examined.

Each individual participating in a CMS-approved Cytology PT Program will be assigned a unique national PT registration number (PRT#) that will remain, regardless of the CMS-approved PT program utilized or future places of employment. Identifying information for individuals will be placed in a Privacy Act protected System of Records at CMS and its confidentiality will be maintained in accordance with applicable law.

**Personnel Requirement for Cytology Proficiency Testing (PT)**

**Cytotechnologist-Newly Certified by ASCP**

- o New graduates of schools of cytotechnology who have taken the Certification Examination in Cytotechnology administered by the American Society for Clinical Pathology (ASCP) Board of Registry (BOR) and obtained a passing score have demonstrated an initial competency level in the examination of cervical cytology. These newly certified individuals will not be monitored for PT by CMS throughout the calendar year in which they passed their ASCP BOR Examination.
- o New graduates of schools of cytotechnology who are employed, have taken the Certification Examination in Cytotechnology administered by the ASCP BOR, but have not obtained a passing score are required to participate in a CMS-approved Cytology Proficiency Testing Program.

**Pathologists-Newly Board Certified**

- o Anatomic pathologists who are newly certified by the American Board of Pathology or the American Osteopathic Board of Pathology have demonstrated an initial level of competency interpreting cervical cytology specimens by

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passing the examination. These newly board certified individuals will not be monitored for PT by CMS throughout the calendar year in which they became board certified in Anatomic Pathology.

- o Cytopathologists who receive added qualifications in Cytopathology from the American Board of Pathology or the American Osteopathic Board of Pathology have demonstrated competency interpreting cervical cytology specimens by passing this examination. These newly board certified individuals will not be monitored for PT by CMS throughout the calendar year in which they became board certified in Cytopathology.

Residents and Fellows

- o Anatomic pathology residents are not required to participate in a CMS-approved Cytology PT Program. Pathology residents are under the constant supervision of fully licensed physicians and are not responsible for the final diagnosis of cervical cytology specimens.
- o Anatomic pathology fellows whose responsibilities in the cytology laboratory include the examination and interpretation of gynecologic specimens must enroll and achieve a passing score in a CMS-approved Cytology PT Program each calendar year.

All Other Cytotechnologists and Pathologists

- o All other individuals subject to Cytology PT must enroll and be tested during each calendar year.

**FED - D2134 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.855(a)

**Type** Standard

**Regulation Definition**

The laboratory must ensure that each individual is tested at least once per year and obtains a passing score. To ensure this annual testing of individuals, an announced or unannounced testing event will be conducted on-site in each laboratory at least once each year. Laboratories will be notified of the time of each announced on-site testing event at least 30 days prior to each event. Additional testing events will be conducted as necessary in each State or region for the purpose of testing individuals who miss the on-site testing event and for retesting individuals as described in paragraph (b) of this section.

**Interpretive Guideline**

The regulations require that all laboratory personnel who examine gynecologic cytology slide preparations must be present in the laboratory to take the proficiency test on the date the laboratory is scheduled for the testing. The precise dates of testing and logistical arrangements are the responsibility of the laboratory and the PT provider. Those individuals not present for the test on the scheduled date will need to have an excused absence, verified by the Laboratory Director. Participants who miss the scheduled on-site test without an excused absence will receive a failing score of "0." Laboratories must contact the PT program to determine when and where the make-up examination will take place. Examples of "excused" absences include prior scheduled leave, natural disasters, hospitalization, death in the family, etc. Those individuals working at more than one location must identify the laboratory where they will be tested prior to the first testing event. A passing score is 90%.

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FED - D2136 - CYTOLOGY

**Title** CYTOLOGY

**CFR** 493.855(b)

**Type** Standard

**Regulation Definition**

The laboratory must ensure that each individual participates in an annual testing event that involves the examination of a 10-slide test set as described in §493.945.

**Interpretive Guideline**

FED - D2137 - CYTOLOGY

**Title** CYTOLOGY

**CFR** 493.855(b)

**Type** Standard

**Regulation Definition**

Individuals who fail this testing event are retested with another 10-slide test set as described in paragraphs (b)(1) and (b)(2) of this section.

**Interpretive Guideline**

FED - D2138 - CYTOLOGY

**Title** CYTOLOGY

**CFR** 493.855(b)

**Type** Standard

**Regulation Definition**

Individuals who fail this second test are subsequently retested with a 20-slide test set as described in paragraphs (b)(2) and

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(b)(3) of this section. Individuals are given not more than 2 hours to complete a 10-slide test and not more than 4 hours to complete a 20-slide test.

**FED - D2141 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.855(b)

**Type** Standard

**Regulation Definition**

Unexcused failure to appear by an individual for a retest will result in test failure with resulting remediation and limitations on slide examinations as specified in (b)(1), (b)(2), and (b)(3) of this section.

**Interpretive Guideline**

If a test is missed due to an unexcused absence, the individual receives a test score of "0".

If the test is missed for an excused absence, laboratories must contact the proficiency testing program to determine when and where the make-up examination will take place. Examples of "excused" absences include prior scheduled leave, natural disasters, hospitalization, death in the family, etc.

**FED - D2142 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 483.855(b)(1)

**Type** Standard

**Regulation Definition**

An individual is determined to have failed the annual testing event if he or she scores less than 90 percent on a 10-slide test set.

**Interpretive Guideline**

**FED - D2143 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.855(b)(1)

**Type** Standard

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**Regulation Definition**

For an individual who fails an annual proficiency testing event, the laboratory must schedule a retesting event which must take place not more than 45 days after receipt of the notification of failure.

**Interpretive Guideline**

**FED - D2144 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.855(b)(2)

**Type** Standard

**Regulation Definition**

An individual is determined to have failed the second testing event if he or she scores less than 90 percent on a 10-slide test set.

**Interpretive Guideline**

**FED - D2145 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.855(b)(2)

**Type** Standard

**Regulation Definition**

For an individual who fails a second testing event, the laboratory must provide him or her with documented, remedial training and education in the area of failure, and

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FED - D2146 - CYTOLOGY

**Title** CYTOLOGY

**CFR** 493.855(b)(2)

**Type** Standard

**Regulation Definition**

must ensure that all gynecologic slides evaluated subsequent to the notice of failure are reexamined until the individual is again retested with a 20-slide test set and scores at least 90 percent.

**Interpretive Guideline**

FED - D2147 - CYTOLOGY

**Title** CYTOLOGY

**CFR** 493.855(b)(2)

**Type** Standard

**Regulation Definition**

Reexamination of slides must be documented.

**Interpretive Guideline**

FED - D2148 - CYTOLOGY

**Title** CYTOLOGY

**CFR** 493.855(b)(3)

**Type** Standard

**Regulation Definition**

An individual is determined to have failed the third testing event if he or she scores less than 90 percent on a 20-slide test set.

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**FED - D2149 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.855(b)(3)

**Type** Standard

**Regulation Definition**

An individual who fails the third testing event must cease examining gynecologic slide preparations immediately upon notification of test failure and

**Interpretive Guideline**

**FED - D2150 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.855(b)(3)

**Type** Standard

**Regulation Definition**

may not resume examining gynecologic slides until the laboratory ensures that the individual obtains at least 35 hours of documented, formally structured, continuing education in diagnostic cytopathology that focuses on the examination of gynecologic preparations, and until he or she is retested with a 20-slide test set and scores at least 90 percent.

**Interpretive Guideline**

**FED - D2151 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.855(c)

**Type** Standard

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**Regulation Definition**

If a laboratory fails to ensure that individuals are tested or those who fail a testing event are retested, or fails to take required remedial actions as described in paragraphs (b)(1), (b)(2) or (b)(3) of this section, CMS will initiate intermediate sanctions or limit the laboratory's certificate to exclude gynecologic cytology testing under CLIA, and, if applicable, suspend the laboratory's Medicare and Medicaid payments for gynecologic cytology testing in accordance with subpart R of this part.

**Interpretive Guideline**

Any laboratory testing patient specimens for the Human Papillomavirus (HPV) must enroll and successfully participate in a CMS-approved proficiency testing program for HPV beginning in 2008. Laboratories should refer to Subpart H for further information. The laboratory's CLIA certificate must include the subspecialty of Virology regardless of where the testing is performed.

**FED - D2153 - ABO GROUP AND D(RHO) TYPING**

**Title** ABO GROUP AND D(RHO) TYPING

**CFR** 493.859(a)

**Type** Standard

**Regulation Definition**

Failure to attain a score of at least 100 percent of acceptable responses for each analyte or test in each testing event is unsatisfactory analyte performance for the testing event.

**Interpretive Guideline**

Analytes or tests for which laboratory PT performance is to be evaluated:

- o ABO group (excluding subgroups)
- o D(Rho) typing
- o Unexpected antibody detection
- o Compatibility testing
- o Antibody identification

**FED - D2154 - ABO GROUP AND D(RHO) TYPING**

**Title** ABO GROUP AND D(RHO) TYPING

**CFR** 493.859(b)

**Type** Standard

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**Regulation Definition**

Failure to attain an overall testing event score of at least 100 percent is unsatisfactory performance.

**Interpretive Guideline**

**FED - D2155 - ABO GROUP AND D(RHO) TYPING**

**Title** ABO GROUP AND D(RHO) TYPING

**CFR** 493.859(c)

**Type** Standard

**Regulation Definition**

Failure to participate in a testing event is unsatisfactory performance and results in a score of 0 for the testing event. Consideration may be given to those laboratories failing to participate in a testing event only if--

- (1) Patient testing was suspended during the time frame allotted for testing and reporting proficiency testing results;
- (2) The laboratory notifies the inspecting agency and the proficiency testing program within the time frame for submitting proficiency testing results of the suspension of patient testing and the circumstances associated with failure to perform tests on proficiency testing samples; and
- (3) The laboratory participated in the previous two proficiency testing events.

**Interpretive Guideline**

**FED - D2159 - ABO GROUP AND D(RHO) TYPING**

**Title** ABO GROUP AND D(RHO) TYPING

**CFR** 493.859(d)

**Type** Standard

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**Regulation Definition**

Failure to return proficiency testing results to the proficiency testing program within the time frame specified by the program is unsatisfactory performance and results in a score of 0 for the testing event.

**Interpretive Guideline**

**FED - D2160 - ABO GROUP AND D(RHO) TYPING**

**Title** ABO GROUP AND D(RHO) TYPING

**CFR** 493.859(e)

**Type** Standard

**Regulation Definition**

(1) For any unsatisfactory testing event for reasons other than a failure to participate, the laboratory must undertake appropriate training and employ the technical assistance necessary to correct problems associated with a proficiency testing failure.

(2) For any unacceptable analyte or unsatisfactory testing event score, remedial action must be taken and documented, and the documentation must be maintained by the laboratory for two years from the date of participation in the proficiency testing event.

**Interpretive Guideline**

**FED - D2162 - ABO GROUP AND D(RHO) TYPING**

**Title** ABO GROUP AND D(RHO) TYPING

**CFR** 493.859(f)

**Type** Standard

**Regulation Definition**

Failure to achieve satisfactory performance for the same analyte in two consecutive testing events or two out of three

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consecutive testing events is unsuccessful performance.

**FED - D2163 - ABO GROUP AND D(RHO) TYPING**

**Title** ABO GROUP AND D(RHO) TYPING

**CFR** 493.859(g)

**Type** Standard

**Regulation Definition**

Failure to achieve an overall testing event score of satisfactory for two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

**Interpretive Guideline**

**FED - D2164 - UNEXPECTED ANTIBODY DETECTION**

**Title** UNEXPECTED ANTIBODY DETECTION

**CFR** 493.861(a)

**Type** Standard

**Regulation Definition**

Failure to attain an overall testing event score of at least 80 percent is unsatisfactory performance.

**Interpretive Guideline**

**FED - D2165 - UNEXPECTED ANTIBODY DETECTION**

**Title** UNEXPECTED ANTIBODY DETECTION

**CFR** 493.861(b)

**Type** Standard

**Regulation Definition**

Failure to participate in a testing event is unsatisfactory performance and results in a score of 0 for the testing event.

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Consideration may be given to those laboratories failing to participate in a testing event only if--

- (1) Patient testing was suspended during the time frame allotted for testing and reporting proficiency testing results;
- (2) The laboratory notifies the inspecting agency and the proficiency testing program within the time frame for submitting proficiency testing results of the suspension of patient testing and the circumstances associated with failure to perform tests on proficiency testing samples; and
- (3) The laboratory participated in the previous two proficiency testing events.

**FED - D2169 - UNEXPECTED ANTIBODY DETECTION**

**Title** UNEXPECTED ANTIBODY DETECTION

**CFR** 493.861(c)

**Type** Standard

**Regulation Definition**

Failure to return proficiency testing results to the proficiency testing program within the time frame specified by the program is unsatisfactory performance and results in a score of 0 for the testing event.

**Interpretive Guideline**

**FED - D2170 - UNEXPECTED ANTIBODY DETECTION**

**Title** UNEXPECTED ANTIBODY DETECTION

**CFR** 493.861(d)

**Type** Standard

**Regulation Definition**

- (1) For any unsatisfactory testing event for reasons other than a failure to participate, the laboratory must undertake appropriate training and employ the technical assistance

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necessary to correct problems associated with a proficiency testing failure.

(2) For any unsatisfactory testing event score, remedial action must be taken and documented, and the documentation must be maintained by the laboratory for two years from the date of participation in the proficiency testing event.

**FED - D2172 - UNEXPECTED ANTIBODY DETECTION**

**Title** UNEXPECTED ANTIBODY DETECTION

**CFR** 493.861(e)

**Type** Standard

**Regulation Definition**

Failure to achieve an overall testing event score of satisfactory for two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

**Interpretive Guideline**

**FED - D2173 - COMPATIBILITY TESTING**

**Title** COMPATIBILITY TESTING

**CFR** 493.863(a)

**Type** Standard

**Regulation Definition**

Failure to attain an overall testing event score of at least 100 percent is unsatisfactory performance.

**Interpretive Guideline**

**FED - D2174 - COMPATIBILITY TESTING**

**Title** COMPATIBILITY TESTING

**CFR** 493.863(b)

**Type** Standard

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**Regulation Definition**

**Interpretive Guideline**

Failure to participate in a testing event is unsatisfactory performance and results in a score of 0 for the testing event. Consideration may be given to those laboratories failing to participate in a testing event only if--

- (1) Patient testing was suspended during the time frame allotted for testing and reporting proficiency testing results;
- (2) The laboratory notifies the inspecting agency and the proficiency testing program within the time frame for submitting proficiency testing results of the suspension of patient testing and the circumstances associated with failure to perform tests on proficiency testing samples; and
- (3) The laboratory participated in the previous two proficiency testing events.

FED - D2178 - COMPATIBILITY TESTING

**Title** COMPATIBILITY TESTING

**CFR** 493.863(c)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to return proficiency testing results to the proficiency testing program within the time frame specified by the program is unsatisfactory performance and results in a score of 0 for the testing event.

FED - D2179 - COMPATIBILITY TESTING

**Title** COMPATIBILITY TESTING

**CFR** 493.863(d)

**Type** Standard

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**Regulation Definition**

**Interpretive Guideline**

(1) For any unsatisfactory testing event for reasons other than a failure to participate, the laboratory must undertake appropriate training and employ the technical assistance necessary to correct problems associated with a proficiency testing failure.

(2) For any unsatisfactory testing event score, remedial action must be taken and documented, and the documentation must be maintained by the laboratory for two years from the date of participation in the proficiency testing event.

**FED - D2181 - COMPATIBILITY TESTING**

**Title** COMPATIBILITY TESTING

**CFR** 493.863(e)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to achieve an overall testing event score of satisfactory for two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

**FED - D2182 - ANTIBODY IDENTIFICATION**

**Title** ANTIBODY IDENTIFICATION

**CFR** 493.865(a)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to attain an overall testing event score of at least 80 percent is unsatisfactory performance.

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**FED - D2183 - ANTIBODY IDENTIFICATION**

**Title** ANTIBODY IDENTIFICATION

**CFR** 493.865(b)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to participate in a testing event is unsatisfactory performance and results in a score of 0 for the testing event. Consideration may be given to those laboratories failing to participate in a testing event only if--

- (1) Patient testing was suspended during the time frame allotted for testing and reporting proficiency testing results;
- (2) The laboratory notifies the inspecting agency and the proficiency testing program within the time frame for submitting proficiency testing results of the suspension of patient testing and the circumstances associated with failure to perform tests on proficiency testing samples; and
- (3) The laboratory participated in the previous two proficiency testing events.

**FED - D2187 - ANTIBODY IDENTIFICATION**

**Title** ANTIBODY IDENTIFICATION

**CFR** 493.865(c)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

Failure to return proficiency testing results to the proficiency testing program within the time frame specified by the program is unsatisfactory performance and results in a score of 0 for the testing event.

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**FED - D2188 - ANTIBODY IDENTIFICATION**

**Title** ANTIBODY IDENTIFICATION

**CFR** 493.865(d)

**Type** Standard

**Regulation Definition**

- (1) For any unsatisfactory testing event for reasons other than a failure to participate, the laboratory must undertake appropriate training and employ the technical assistance necessary to correct problems associated with a proficiency testing failure.
- (2) For any unsatisfactory testing event score, remedial action must be taken and documented, and the documentation must be maintained by the laboratory for two years from the date of participation in the proficiency testing event.

**Interpretive Guideline**

**FED - D2190 - ANTIBODY IDENTIFICATION**

**Title** ANTIBODY IDENTIFICATION

**CFR** 493.865(e)

**Type** Standard

**Regulation Definition**

Failure to identify the same antibody in two consecutive or two out of three consecutive testing events is unsuccessful performance.

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**FED - D2191 - ANTIBODY IDENTIFICATION**

**Title** ANTIBODY IDENTIFICATION

**CFR** 493.865(f)

**Type** Standard

**Regulation Definition**

Failure to achieve an overall testing event score of satisfactory for two consecutive testing events or two out of three consecutive testing events is unsuccessful performance.

**Interpretive Guideline**

**FED - D3000 - FACILITY ADMINISTRATION**

**Title** FACILITY ADMINISTRATION

**CFR** 493.1100

**Type** Condition

**Regulation Definition**

Each laboratory that performs nonwaived testing must meet the applicable requirements under §§493.1101 through 493.1105, unless HHS approves a procedure that provides equivalent quality testing as specified in Appendix C of the State Operations Manual (CMS Pub. 7).

**Interpretive Guideline**

To determine which tests are categorized as waived or nonwaived (i.e. moderate or high complexity tests), refer to the following web link for the FDA categorization database (<<http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfCLIA/Search.cfm?sAN=0>> Test systems, assays, and examinations not yet classified are considered high complexity.

Significant deficiencies cited under this condition may also indicate deficiencies under personnel responsibilities.

**FED - D3001 - FACILITIES**

**Title** FACILITIES

**CFR** 493.1101(a)(1)

**Type** Standard

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**Regulation Definition**

The laboratory must be constructed, arranged, and maintained to ensure the space, ventilation, and utilities necessary for conducting all phases of the testing process.

**Interpretive Guideline**

Work areas should be arranged to minimize problems in specimen handling, examination and testing, and the reporting of test results.

Workbench space should be sufficient for test performance, well lit, and have water, gas, suction, and, electrical outlets as necessary. Instruments, equipment, and computer systems should be placed in locations where their operation is not affected adversely by physical or chemical factors, such as heat, direct sunlight, vibrations, power fluctuations or fumes from acid or alkaline solutions. Equipment tops should not be used as workbench space.

Determination of proper lighting is subjective since the regulations do not specify the foot-candles or other measures of light intensity required. Ensure that lighting or background is appropriate for visual interpretation of test results (e.g. macroscopic evaluation of hemagglutination reactions or strep screen; dark background with reflected light for reading K-B disk diffusion AST). When citing deficiencies, document the circumstances in which lighting adversely or may adversely affect test performance or personnel safety.

Determine that the laboratory has a system to ensure its ventilation system properly removes vapors, fumes, and excessive heat, when appropriate, for the type of testing done in the laboratory.

Ensure that an adequate, stable electrical source is maintained at each location (e.g. outlets, not extension cords) and meets the power requirements for each piece of equipment.

**FED - D3003 - FACILITIES**

**Title** FACILITIES

**CFR** 493.1101(a)(2)

**Type** Standard

**Regulation Definition**

The laboratory must be constructed, arranged, and maintained to ensure contamination of patient specimens, equipment, instruments, reagents, materials, and supplies is minimized.

**Interpretive Guideline**

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**FED - D3005 - FACILITIES**

**Title** FACILITIES

**CFR** 493.1101(a)(3)

**Type** Standard

**Regulation Definition**

Molecular amplification procedures that are not contained in closed systems have a uni-directional workflow. This must include separate areas for specimen preparation, amplification and product detection, and, as applicable, reagent preparation.

**Interpretive Guideline**

The laboratory should establish contamination prevention procedures to minimize contamination of patient specimens, equipment, instruments, reagents, materials, and supplies.

Determine if the laboratory performs wipe tests of areas where radioactive material or amplification procedures are used in order to monitor and prevent contamination.

Laboratories performing molecular amplification procedures should have a mechanism to detect cross-contamination of patient specimens. This may be accomplished by including a "blank" control with each run of patient specimen testing (use D5425).

The "blank" control refers to a no-template control (N.T.C) or a control sample containing all reagents except the target template.

An example of a "closed system" would be an FDA-cleared or FDA-approved test system that contains amplification and detection steps in sealed tubes that are never opened or re-opened during or after the testing process and that is used as directed or suggested by the manufacturer (i.e., without any modifications).

Unidirectional workflow refers to the manner in which testing personnel and patient specimens move through the molecular testing process to prevent cross-contamination, and consists of separate areas for the following:

- o Reagent preparation (as applicable);
- o Pre-amplification area for specimen preparation and amplification reaction set up; and
- o Post-amplification area for specimen amplification, product detection, and storage or disposal of amplified products.

Reagents must be prepared in an area that is separate (as applicable) from where specimens are processed, prepared, "amplified" and detected to prevent contamination. Once a specimen enters the amplification and product detection

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area it should not be brought back to the reagent or specimen preparation areas. The laboratory should store amplified specimens separately from test reagents and patient specimens. All equipment (e.g. reagents, supplies, pens, pipettes and tips, laboratory coats) should remain in designated areas.

Sources of potential cross-contamination in molecular testing include:

- o Patient specimen (i.e. genomic contamination);
- o Amplified patient specimen (i.e. amplicon contamination); and
- o Testing personnel.

**FED - D3007 - FACILITIES**

**Title** FACILITIES

**CFR** 493.1101(b)

**Type** Standard

**Regulation Definition**

The laboratory must have appropriate and sufficient equipment, instruments, reagents, materials, and supplies for the type and volume of testing it performs.

**Interpretive Guideline**

Base deficiencies related to inappropriate or insufficient equipment on a determination that patient results are or may be adversely affected. Ensure that the laboratory has the appropriate equipment to prepare reagents, stains, solutions, controls, and calibration materials (e.g. pipettes, hydrometers, graduated cylinders, autoclaves, balances, centrifuges, distilled/deionized water). If the equipment or instrumentation is found to be inappropriate or insufficient, document the reasons for this finding.

Ensure that the laboratory has test systems, equipment and/or instruments capable of producing results within the laboratory's stated test performance specifications.

Ensure that the laboratory has test systems, equipment and/or instruments necessary to perform the laboratory's volume of testing (preanalytic, analytic, postanalytic) within established turnaround times.

Data capacity in the laboratory's information system should be sufficient for current data entry. If capacity is maintained by deletion of data, it should be scheduled and documented.

For Cytology, laboratories should use coverslips that cover the entire surface of the specimen.

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**FED - D3009 - FACILITIES**

**Title** FACILITIES

**CFR** 493.1101(c)

**Type** Standard

**Regulation Definition**

The laboratory must be in compliance with applicable Federal, State, and local laboratory requirements.

**Interpretive Guideline**

The laboratory must possess a current license issued by the State or local government, if such licensing exists. If a State or local government removes a laboratory's license and the right to operate within the State or locality, Centers for Medicare and Medicaid Services (CMS) may take an action to revoke the Clinical Laboratory Improvement Amendments (CLIA) certificate.

**FED - D3011 - FACILITIES**

**Title** FACILITIES

**CFR** 493.1101(d)

**Type** Standard

**Regulation Definition**

Safety procedures must be established, accessible, and observed to ensure protection from physical, chemical, biochemical, and electrical hazards, and biohazardous materials.

**Interpretive Guideline**

If you observe or obtain information regarding potential safety violations not applicable under CLIA, notify the appropriate State or local authority. Consult with the Regional Office (RO) for notification to other Federal agencies such as the Occupational Safety and Health Administration (OSHA) [www.osha.gov](http://www.osha.gov) <<http://www.osha.gov>>, Environmental Protection Agency (EPA) [www.epa.gov](http://www.epa.gov) <<http://www.epa.gov>>, or Nuclear Regulatory Commission (NRC). The appropriate Federal, State or local authority, if warranted, will investigate and, if necessary, conduct an on-site visit.

Probes §493.1101(d)

What safety protocols are observed and practiced in the laboratory?

How does the laboratory, including temporary testing sites or mobile units:

- o Dispose of radiological, chemical, and biological wastes (including blood drawing equipment);

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- o Clean up spills (chemical, biological, and radiological); and
  
- o Determine the amount of waste that can safely be contained and the precautions necessary to ensure that liquid waste does not spill or splash while in travel status?

What chemical, radiological, or biological precautions are taken, if any, during the preparation or handling of reagents?

**FED - D3013 - FACILITIES**

**Title** FACILITIES

**CFR** 493.1101(e)

**Type** Standard

**Regulation Definition**

Records and, as applicable, slides, blocks, and tissues must be maintained and stored under conditions that ensure proper preservation.

**Interpretive Guideline**

Interpretative Guidelines §493.1101(e)

The laboratory must arrange a secure area for storage of records and, as applicable, slides, blocks, and tissues that will provide conditions that ensure proper preservation of specimens and records.

Paraffin blocks must be stored in a cool dry environment. Exposure to excessive heat may cause blocks to melt.

Probes §493.1101(e)

For Cytology and Histology, how does the laboratory ensure that the slides have completely dried prior to being stored?

**FED - D3015 - REQUIREMENTS FOR TRANSFUSION SERVICES**

**Title** REQUIREMENTS FOR TRANSFUSION SERVICES

**CFR** 493.1103

**Type** Standard

**Regulation Definition**

A facility that provides transfusion services must meet all of

**Interpretive Guideline**

Interpretative Guidelines §493.1103

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the requirements of this section and document all transfusion-related activities.

A "facility that provides transfusion services" is any entity that may store and/or administer blood and blood products to patients.

**FED - D3017 - REQUIREMENTS FOR TRANSFUSION SERVICES**

**Title** REQUIREMENTS FOR TRANSFUSION SERVICES

**CFR** 493.1103(a)

**Type** Standard

**Regulation Definition**

Arrangement for services. The facility must have a transfusion service agreement reviewed and approved by the responsible party(ies) that govern the procurement, transfer, and availability of blood and blood products.

**Interpretive Guideline**

Interpretative Guidelines §493.1103(a)

Determine which services are provided directly by the facility and which are provided through agreement and ensure that the agreement is being met.

**FED - D3019 - REQUIREMENTS FOR TRANSFUSION SERVICES**

**Title** REQUIREMENTS FOR TRANSFUSION SERVICES

**CFR** 493.1103(b)

**Type** Standard

**Regulation Definition**

Provision of testing. The facility must provide prompt ABO grouping, D (Rho) typing, unexpected antibody detection, compatibility testing, and laboratory investigation of transfusion reactions on a continuous basis through a CLIA-certified laboratory or a laboratory meeting equivalent requirements as determined by CMS.

**Interpretive Guideline**

Interpretative Guidelines §493.1103(b)

Review the agreement and determine if the outside laboratory is CLIA-certified or equivalent, as determined by CMS. An equivalent laboratory is a Veterans Health Administration (VHA) laboratory, a CLIA-exempt laboratory or a laboratory under the auspices of the Department of Defense (DoD).

Probes §493.1103

For laboratories performing ABO grouping, D typing, unexpected antibody detection or compatibility testing using automated methods, is there a back-up system in place to ensure availability of service on a continuous basis when the automated system is malfunctioning?

Is staff trained and competent in the back-up system?

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**FED - D3021 - REQUIREMENTS FOR TRANSFUSION SERVICES**

**Title** REQUIREMENTS FOR TRANSFUSION SERVICES

**CFR** 493.1103(c)(1)

**Type** Standard

**Regulation Definition**

Blood and blood products storage and distribution. If a facility stores or maintains blood or blood products for transfusion outside of a monitored refrigerator, the facility must ensure the storage conditions, including temperature, are appropriate to prevent deterioration of the blood or blood product.

**Interpretive Guideline**

Interpretative Guidelines §493.1103(c)(1)

Determine where blood and blood products are stored. There may be various unconventional blood storage areas such as operating rooms, nursing stations, long-term care facilities, and dialysis units. Determine that the facility ensures the appropriate temperature is maintained and documented for each storage area during the time blood and blood products are stored.

Acceptable temperature ranges must be established and actual readings of temperature-controlled storage areas must be recorded during the time that blood or blood products for transfusion are stored. Whole Blood, Red Blood cells, and Thawed Plasma should be stored between 1 and 6°C; Platelets and Thawed Cryoprecipitated AHF should be stored between 20 and 24°C; Fresh Frozen Plasma, Plasma Frozen within 24 hours after Phlebotomy, and Cryoprecipitated AHF should be stored at -18°C or colder.

Facilities that provide transfusion services (not certified for the specialty of Immunohematology) and perform nonwaived testing are held to the requirements for the storage and distribution of blood and blood products. The laboratory providing the blood or blood products may supply these facilities with the following:

- o Policies for the proper storage and transportation of blood or blood products;
- o Procedures to alert the laboratory of blood storage problems;
- o Policies to ensure the positive identification of a blood or blood product recipient (use D3023);
- o Procedures to identify a possible transfusion reaction (use D3025); and
- o Procedures to notify the laboratory of a possible transfusion reaction (use D3025).

Determine how the appropriate temperatures of blood storage areas are maintained during a power failure.

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Blood shall be stored in a clean and orderly environment in a manner to prevent mix-ups. No expired blood should be in the routine inventory. Unacceptable units should be segregated from routine inventory.

Probes §493.1103(c)(1)

If frozen blood products are stored, how does the facility ensure products are maintained at appropriate temperatures to prevent thawing and re-freezing of the products?

**FED - D3023 - REQUIREMENTS FOR TRANSFUSION SERVICES**

**Title** REQUIREMENTS FOR TRANSFUSION SERVICES

**CFR** 493.1103(c)(2)

**Type** Standard

**Regulation Definition**

The facility must establish and follow policies to ensure positive identification of a blood or blood product recipient.

**Interpretive Guideline**

Interpretative Guidelines §493.1103(c)(2)

Review the facility's policies for ensuring positive identification of blood or blood products and the intended recipient.

When possible, observe the actual practice, including issuing the blood and blood products to the intended recipient. This includes proper verification of patient identification prior to initiation of the transfusion.

**FED - D3025 - REQUIREMENTS FOR TRANSFUSION SERVICES**

**Title** REQUIREMENTS FOR TRANSFUSION SERVICES

**CFR** 493.1103(d)

**Type** Standard

**Regulation Definition**

Investigation of transfusion reactions. The facility must have procedures for preventing transfusion reactions and when necessary, promptly identify, investigate, and report blood and blood product transfusion reactions to the laboratory and, as

**Interpretive Guideline**

Interpretative Guidelines §493.1103(d)

Review the procedures for preventing, identifying, and investigating transfusion reactions. Examine records of transfusion reaction investigations for completeness, promptness, and accuracy. Verify that investigations of

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appropriate, to Federal and State authorities.

transfusion reactions are conducted in accordance with the facility's established protocols. Also, verify that incidents such as incomplete compatibility testing or issuing the wrong unit to a specific patient are reported to the appropriate authorities. Records should include each step in the investigation and identify the reviewer.

For facilities that provide transfusion services, confirm that all transfusion reactions identified have been investigated and the Food and Drug Administration (FDA) has been notified of all transfusion related fatalities. If the FDA has not been notified, notify the FDA at:

Food and Drug Administration  
Center for Biologics Evaluation and Research (CBER)  
Director, Office of Compliance and Biologics Quality  
Attn: Fatality Program Manager (HFM-650)  
1401 Rockville Pike  
Rockville, MD 20852-1448

Voicemail: 301-827-6220

E-mail: [fatalities2@cber.fda.gov](mailto:fatalities2@cber.fda.gov)

Fax: 301-827-6748

NOTE: Send the RO reports of all the fatal transfusion reactions identified. These reports are used to ensure that the facilities have properly notified the FDA of fatal transfusion reactions and that both CMS and the FDA have conducted all necessary follow-ups.

Probes §493.1103(d)

Are problems detected during the course of the transfusion reaction investigation documented, and are procedures instituted to prevent a recurrence?

**FED - D3027 - RETENTION REQUIREMENTS**

**Title** RETENTION REQUIREMENTS

**CFR** 493.1105(a)(1)

**Type** Standard

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**Regulation Definition**

Test requisitions and authorizations. Retain records of test requisitions and test authorizations, including the patient's chart or medical record if used as the test requisition or authorization, for at least 2 years.

**Interpretive Guideline**

The regulation applies to manual as well as automated record systems, i.e. laboratory information system (LIS). However, the regulation does not specify the mechanism or frequency for which a laboratory should evaluate its record storage and retrieval system(s). The laboratory should establish its own policies and procedures for evaluating its system(s) and maintain documentation of the evaluation.

**FED - D3029 - RETENTION REQUIREMENTS**

**Title** RETENTION REQUIREMENTS

**CFR** 493.1105(a)(2)

**Type** Standard

**Regulation Definition**

Test procedures. Retain a copy of each test procedure for at least 2 years after a procedure has been discontinued. Each test procedure must include the dates of initial use and discontinuance.

**Interpretive Guideline**

The regulation applies to manual as well as automated record systems, i.e. laboratory information system (LIS). However, the regulation does not specify the mechanism or frequency for which a laboratory should evaluate its record storage and retrieval system(s). The laboratory should establish its own policies and procedures for evaluating its system(s) and maintain documentation of the evaluation.

**FED - D3031 - RETENTION REQUIREMENTS**

**Title** RETENTION REQUIREMENTS

**CFR** 493.1105(a)(3)

**Type** Standard

**Regulation Definition**

Analytic systems records. Retain quality control and patient test records (including instrument printouts, if applicable) and records documenting all analytic systems activities specified in §§493.1252 through 493.1289 for at least 2 years.

**Interpretive Guideline**

Interpretative Guidelines §493.1105(a)(3)

The records must include instrument charts, graphs, printouts, transcribed data, and manufacturers' assay information sheets for control and calibration materials. If data are transcribed, ensure that the original and the transcribed copy are retained for 2 years.

Printouts from an instrument that is not directly interfaced with the laboratory information system must be retained for 2 years.

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NOTE: Thermal paper or pressure sensitive paper may fade over time. Where necessary, the laboratory is expected to make an electronic or hard copy of applicable result printouts to ensure that they are retrievable and legible for at least two years.

The laboratory is responsible for retaining records of interpretative slide results of each gynecologic and nongynecologic cytology case that each cytotechnologist examined or reviewed for at least five years.

The regulation applies to manual as well as automated record systems, i.e. laboratory information system (LIS). However, the regulation does not specify the mechanism or frequency for which a laboratory should evaluate its record storage and retrieval system(s). The laboratory should establish its own policies and procedures for evaluating its system(s) and maintain documentation of the evaluation.

**FED - D3033 - RETENTION REQUIREMENTS**

**Title** RETENTION REQUIREMENTS

**CFR** 493.1105(a)(3)(i)

**Type** Standard

**Regulation Definition**

In addition, the laboratory must retain records of test system performance specifications that the laboratory establishes or verifies under §493.1253 for the period of time the laboratory uses the test system but no less than 2 years.

**Interpretive Guideline**

Interpretative Guidelines §493.1105(a)

The regulation applies to manual as well as automated record systems, i.e. laboratory information system (LIS). However, the regulation does not specify the mechanism or frequency for which a laboratory should evaluate its record storage and retrieval system(s). The laboratory should establish its own policies and procedures for evaluating its system(s) and maintain documentation of the evaluation.

**FED - D3035 - RETENTION REQUIREMENTS**

**Title** RETENTION REQUIREMENTS

**CFR** 493.1105(a)(3)(ii)

**Type** Standard

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**Regulation Definition**

In addition, the laboratory must retain immunohematology records, blood and blood product records, and transfusion records as specified in 21 CFR 606.160(b)(3)(ii), (b)(3)(iv), (b)(3)(v), and (d).

Interpretive Guidelines §493.1105(a)

The regulation applies to manual as well as automated record systems, i.e. laboratory information system (LIS). However, the regulation does not specify the mechanism or frequency for which a laboratory should evaluate its record storage and retrieval system(s). The laboratory should establish its own policies and procedures for evaluating its system(s) and maintain documentation of the evaluation.

**Interpretive Guideline**

Interpretative Guidelines §493.1105(a)(3)(ii)

Refer to the current version of 21 CFR Part 606.160

<<http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=606.160>> for the specified section.

Non-transfusion related immunohematology patient testing and quality control (QC) records, such as instrument function checks, maintenance, and temperature records, must be retained for at least 2 years.

Other immunohematology patient and QC records related to transfusion testing, including but not limited to, donor processing, compatibility testing, and transfusion reaction investigations, must be retained for the time frame stated at 21 CFR §606.160(d). This also includes the visual inspection of whole blood and red blood cells during storage and immediately before distribution [21CFR §606.160(b)(3)(ii)

<<http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=606.160>>], record of reissue, including records of proper temperature maintenance [21CFR §606.160(b)(3)(iv)

<<http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=606.160>>], and emergency release of blood, including signature of requesting physician obtained before or after release [21 CFR §606.160(b)(3)(v)

<<http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=606.160>>].

Interpretative Guidelines §493.1105(a)

The regulation applies to manual as well as automated record systems, i.e. laboratory information system (LIS). However, the regulation does not specify the mechanism or frequency for which a laboratory should evaluate its record storage and retrieval system(s). The laboratory should establish its own policies and procedures for evaluating its system(s) and maintain documentation of the evaluation.

**FED - D3037 - RETENTION REQUIREMENTS**

**Title** RETENTION REQUIREMENTS

**CFR** 493.1105(a)(4)

**Type** Standard

**Regulation Definition**

Proficiency testing records. Retain all proficiency testing

**Interpretive Guideline**

Interpretative Guidelines §493.1105(a)(4)

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records for at least 2 years.

Proficiency testing (PT) records include all information regarding the PT event including testing records, signed attestation statements, PT results and scores from the provider, documentation of review and records of any corrective actions.

Interpretative Guidelines §493.1105(a)

The regulation applies to manual as well as automated record systems, i.e. laboratory information system (LIS). However, the regulation does not specify the mechanism or frequency for which a laboratory should evaluate its record storage and retrieval system(s). The laboratory should establish its own policies and procedures for evaluating its system(s) and maintain documentation of the evaluation.

**FED - D3039 - RETENTION REQUIREMENTS**

**Title** RETENTION REQUIREMENTS

**CFR** 493.1105(a)(5)

**Type** Standard

**Regulation Definition**

Quality system assessment records. Retain all laboratory quality system assessment records for at least 2 years.

**Interpretive Guideline**

Interpretative Guidelines §493.1105(a)(5)

Quality assessment (QA) records do not need to be maintained and stored in one location. The records may be stored in the specific area or department appropriate to the monitoring and evaluation of the laboratory activities (preanalytic, analytic, and postanalytic).

Interpretative Guidelines §493.1105(a)

The regulation applies to manual as well as automated record systems, i.e. laboratory information system (LIS). However, the regulation does not specify the mechanism or frequency for which a laboratory should evaluate its record storage and retrieval system(s). The laboratory should establish its own policies and procedures for evaluating its system(s) and maintain documentation of the evaluation.

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**FED - D3041 - RETENTION REQUIREMENTS**

**Title** RETENTION REQUIREMENTS

**CFR** 493.1105(a)(6)

**Type** Standard

**Regulation Definition**

Test reports. Retain or be able to retrieve a copy of the original report (including final, preliminary, and corrected reports) at least 2 years after the date of reporting.

(i) In addition, retain immunohematology reports as specified in 21 CFR 606.160(d)

(ii) and pathology test reports for at least 10 years after the date of reporting.

**Interpretive Guideline**

Interpretative Guidelines §493.1105(a)(6)

A copy, either paper or electronic, of the original report includes all information sent to recipients, and includes the name and address of the laboratory performing the test. The copy need not be paper, but may be retrieved from a computer system, microfilm or microfiche record, as long as it contains the exact information as sent to the individual ordering the test or utilizing the test results. The laboratory copy of the report should contain information that provides an accurate, complete, display of previously reported data retained or retrieved from the laboratory's record system.

For test reports from histopathology, oral pathology, or cytology that require personnel identifiers or an authorized signature (which may be electronic), the copy must include evidence of the identifiers or signature(s).

A "preliminary report" means a test result that has been reported directly to the authorized person or laboratory that initially requested the test, directly or through an electronic health record provider or health information exchange prior to the issuance of the final test result(s). Frequently, a preliminary report will contain significant, but not definitive information (e.g. a urine culture preliminary report of >100,000 Gram-negative bacilli after 24 hours incubation or a beta subunit preliminary report of >200 miu/ml). It should be noted on the report when the result is a preliminary result and that a final report will follow.

A "partial report" means multiple tests are ordered on the same specimen or patient. If partial reports are issued for only those tests that have been completed, then the report date will be the date when all tests have been completed. However, the laboratory should be able to identify the date that each new test is appended to the report.

The laboratory must have a system for retaining copies of all reports including original, preliminary, corrected, and final reports. This includes computer-generated reports.

Probes §493.1105(a)(6)

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How has the laboratory verified that its record retrieval system functions appropriately?

Interpretative Guidelines §493.1105(a)(6)(i)

Refer to the current version of 21 CFR Part 600.160

<<http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=606.160>> for the specified section.

Transfusion-related Immunohematology test reports, including but not limited to, donor processing [§493.1271(b)], compatibility testing, and transfusion reaction investigations, must be retained for the time frame stated at 21 CFR §606.160(d).

All Immunohematology test reports not subject to 21 CFR §606.130(d)

<<http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=606.160>> must be retained for at least 2 years

**FED - D3043 - RETENTION REQUIREMENTS**

**Title** RETENTION REQUIREMENTS

**CFR** 493.1105(a)(7)

**Type** Standard

**Regulation Definition**

The laboratory must retain cytology slide preparations for at least 5 years from the date of examination (see §493.1274(f) for proficiency testing exception). The laboratory must retain histopathology slides for at least 10 years from the date of examination. The laboratory must retain pathology specimen blocks for at least 2 years from the date of examination. The laboratory must preserve remnants of tissue for pathology examination until a diagnosis is made on the specimen.

**Interpretive Guideline**

Interpretative Guidelines §493.1105(a)(7)(i)(A)

For storage and maintenance requirements use D3013.

NOTE: Cytology specimens include fine needle aspirates.

Retention of cytology slides:

Example:

A laboratory refers all cytology specimens to a reference laboratory for examination. The reference laboratory examines all slide preparations and reports results only on normal, negative, and unsatisfactory cases. At the request of the referring laboratory, the reference laboratory returns those cases that have reactive, reparative atypia (including

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repair), LSIL, HSIL, all invasive squamous carcinomas, adenocarcinoma, all other malignant neoplasms, and 10% of the normal or negatives cases (including reactive and reparative cases) for quality control review. The referring laboratory must maintain the slides of the cases that it examines and for which it provides diagnosis (i.e., slides exhibiting atypical including repair, LSIL, HSIL, all invasive squamous carcinomas, adenocarcinoma, all other malignant neoplasms, and slides chosen for the 10% rescreen).

The laboratory must maintain documentation to acknowledge the donation of each slide submitted to a proficiency testing program or loaned for other purposes.

Probes §493.1105(a)(7)(i)(A)

What protocol has been established to ensure prompt return of slides, when necessary?

**FED - D3045 - RETENTION REQUIREMENTS**

**Title** RETENTION REQUIREMENTS

**CFR** 493.1105(b)

**Type** Standard

**Regulation Definition**

If the laboratory ceases operation, the laboratory must make provisions to ensure that all records and, as applicable, slides, blocks, and tissue are retained and available for the time frames specified in this section.

**Interpretive Guideline**

**FED - D5002 - BACTERIOLOGY**

**Title** BACTERIOLOGY

**CFR** 493.1201

**Type** Condition

**Regulation Definition**

If the laboratory provides services in the subspecialty of Bacteriology, the laboratory must meet the requirements

**Interpretive Guideline**

Interpretative Guidelines §493.1201

Tests or procedures to detect an antigen are categorized in the subspecialty where the antigen is detected or identified.

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specified in §§493.1230 through 493.1256, §493.1261, and §§493.1281 through 493.1299.

For example, tests or procedures for identifying Group A Streptococcus are categorized in Bacteriology.

**FED - D5004 - MYCOBACTERIOLOGY**

**Title** MYCOBACTERIOLOGY

**CFR** 493.1202

**Type** Condition

**Regulation Definition**

If the laboratory provides services in the subspecialty of Mycobacteriology, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, §493.1262, and §§493.1281 through 493.1299.

**Interpretive Guideline**

Interpretative Guidelines §493.1202

Tests or procedures to detect an antigen are categorized in the subspecialty where the antigen is detected or identified. For example, the procedures to identify Mycobacteria are categorized in Mycobacteriology.

**FED - D5006 - MYCOLOGY**

**Title** MYCOLOGY

**CFR** 493.1203

**Type** Condition

**Regulation Definition**

If the laboratory provides services in the subspecialty of Mycology, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, §493.1263, and §§493.1281 through 493.1299.

**Interpretive Guideline**

Interpretative Guidelines §493.1203

Tests or procedures to detect an antigen are categorized in the subspecialty where the antigen is detected or identified. For example, tests for the identification of fungi are categorized in Mycology.

**FED - D5008 - PARASITOLOGY**

**Title** PARASITOLOGY

**CFR** 493.1204

**Type** Condition

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**Regulation Definition**

If the laboratory provides services in the subspecialty of Parasitology, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, §493.1264, and §§493.1281 through 493.1299.

**Interpretive Guideline**

Interpretative Guidelines §493.1204

Tests or procedures to identify an antigen are categorized in the subspecialty where the antigen is detected or identified. For example, procedures to identify a parasite are categorized in the subspecialty of Parasitology; however, procedures to detect or identify an antibody to the parasite are categorized in the subspecialty of General Immunology.

**FED - D5010 - VIROLOGY**

**Title** VIROLOGY

**CFR** 493.1205

**Type** Condition

**Regulation Definition**

If the laboratory provides services in the subspecialty of Virology, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, §493.1265, and §§493.1281 through 493.1299.

**Interpretive Guideline**

Interpretative Guidelines §493.1205

Tests or procedures to identify the virus (antigen) are categorized in the subspecialty where the antigen is detected or identified. For example, tests or procedures to detect Herpes are categorized in the subspecialty of Virology. Tests or procedures to detect antibodies to Herpes are categorized in the subspecialty of General Immunology.

**FED - D5012 - SYPHILIS SEROLOGY**

**Title** SYPHILIS SEROLOGY

**CFR** 493.1207

**Type** Condition

**Regulation Definition**

If the laboratory provides services in the subspecialty of Syphilis serology, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, and §§493.1281 through 493.1299.

**Interpretive Guideline**

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FED - D5014 - GENERAL IMMUNOLOGY

**Title** GENERAL IMMUNOLOGY

**CFR** 493.1208

**Type** Condition

**Regulation Definition**

If the laboratory provides services in the subspecialty of General immunology, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, and §§493.1281 through 493.1299.

**Interpretive Guideline**

Interpretative Guidelines §493.1208  
Tests or procedures to detect or identify antibodies to a bacteria, virus, parasite, etc., are categorized under the subspecialty of General Immunology.

FED - D5016 - ROUTINE CHEMISTRY

**Title** ROUTINE CHEMISTRY

**CFR** 493.1210

**Type** Condition

**Regulation Definition**

If the laboratory provides services in the subspecialty of Routine Chemistry, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, §493.1267, and §§493.1281 through 493.1299.

**Interpretive Guideline**

FED - D5018 - URINALYSIS

**Title** URINALYSIS

**CFR** 493.1211

**Type** Condition

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**Regulation Definition**

If the laboratory provides services in the subspecialty of Urinalysis, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, and §§493.1281 through 493.1299.

**Interpretive Guideline**

**FED - D5020 - ENDOCRINOLOGY**

**Title** ENDOCRINOLOGY

**CFR** 493.1212

**Type** Condition

**Regulation Definition**

If the laboratory provides services in the subspecialty of Endocrinology, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, and §§493.1281 through 493.1299.

**Interpretive Guideline**

**FED - D5022 - TOXICOLOGY**

**Title** TOXICOLOGY

**CFR** 493.1213

**Type** Condition

**Regulation Definition**

If the laboratory provides services in the subspecialty of Toxicology, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, and §§493.1281 through 493.1299.

**Interpretive Guideline**

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**FED - D5024 - HEMATOLOGY**

**Title** HEMATOLOGY

**CFR** 493.1215

**Type** Condition

**Regulation Definition**

If the laboratory provides services in the specialty of Hematology, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, §493.1269, and §§493.1281 through 493.1299.

**Interpretive Guideline**

**FED - D5026 - IMMUNOHEMATOLOGY**

**Title** IMMUNOHEMATOLOGY

**CFR** 493.1217

**Type** Condition

**Regulation Definition**

If the laboratory provides services in the specialty of Immunohematology, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, §493.1271, and §§493.1281 through 493.1299.

**Interpretive Guideline**

**FED - D5028 - HISTOPATHOLOGY**

**Title** HISTOPATHOLOGY

**CFR** 493.1219

**Type** Condition

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**Regulation Definition**

If the laboratory provides services in the subspecialty of Histopathology, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, §493.1273, and §§493.1281 through 493.1299.

**Interpretive Guideline**

**FED - D5030 - ORAL PATHOLOGY**

**Title** ORAL PATHOLOGY

**CFR** 493.1220

**Type** Condition

**Regulation Definition**

If the laboratory provides services in the subspecialty of Oral Pathology, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, and §§493.1281 through 493.1299.

**Interpretive Guideline**

**FED - D5032 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1221

**Type** Condition

**Regulation Definition**

If the laboratory provides services in the subspecialty of Cytology, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, §493.1274, and §§493.1281 through 493.1299.

**Interpretive Guideline**

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FED - D5034 - CLINICAL CYTOGENETICS

**Title** CLINICAL CYTOGENETICS

**CFR** 493.1225

**Type** Condition

**Regulation Definition**

If the laboratory provides services in the specialty of Clinical cytogenetics, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, §493.1276, and §§493.1281 through 493.1299.

**Interpretive Guideline**

FED - D5040 - RADIOBIOASSAY

**Title** RADIOBIOASSAY

**CFR** 493.1226

**Type** Condition

**Regulation Definition**

If the laboratory provides services in the specialty of Radiobioassay, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, and §§493.1281 through 493.1299.

**Interpretive Guideline**

FED - D5042 - HISTOCOMPATIBILITY

**Title** HISTOCOMPATIBILITY

**CFR** 493.1227

**Type** Condition

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**Regulation Definition**

If the laboratory provides services in the specialty of Histocompatibility, the laboratory must meet the requirements specified in §§493.1230 through 493.1256, §493.1278, and §§493.1281 through 493.1299.

**Interpretive Guideline**

**FED - D5200 - GENERAL LABORATORY SYSTEMS**

**Title** GENERAL LABORATORY SYSTEMS

**CFR** 493.1230

**Type** Condition

**Regulation Definition**

Each laboratory that performs nonwaived testing must meet the applicable general laboratory systems requirements in §§493.1231 through 493.1236, unless HHS approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub. 7), that provides equivalent quality testing. The laboratory must monitor and evaluate the overall quality of the general laboratory systems and correct identified problems specified in §493.1239 for each specialty and subspecialty of testing performed.

**Interpretive Guideline**

Interpretative Guidelines §493.1230

Significant deficiencies cited under this condition may indicate deficiencies under personnel responsibilities. Use D5200 when significant deficiencies are identified that have the potential to adversely affect patient testing, are systemic and pervasive throughout the laboratory, and are not limited to any one specialty or subspecialty.

The requirements in this section address those general operational functions that are not specific to any one specialty or subspecialty.

**FED - D5201 - CONFIDENTIALITY OF PATIENT INFORMATION**

**Title** CONFIDENTIALITY OF PATIENT INFORMATION

**CFR** 493.1231

**Type** Standard

**Regulation Definition**

The laboratory must ensure confidentiality of patient information throughout all phases of the total testing process that are under the laboratory's control.

**Interpretive Guideline**

Probes §493.1231

How does the laboratory "control" visitor access to the laboratory areas where patient information may be easily viewed (e.g., computer terminals, facsimile machines, worksheets)?

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Are there safeguards in place to ensure confidentiality of patient information and test reports? For example, are unauthorized users prohibited from gaining entry?

How does the laboratory ensure its record storage system(s) is secure?

**FED - D5203 - SPECIMEN IDENTIFICATION AND INTEGRITY**

**Title** SPECIMEN IDENTIFICATION AND INTEGRITY

**CFR** 493.1232

**Type** Standard

**Regulation Definition**

The laboratory must establish and follow written policies and procedures that ensure positive identification and optimum integrity of a patient's specimen from the time of collection or receipt of the specimen through completion of testing and reporting of results.

**Interpretive Guideline**

Interpretative Guidelines §493.1232

The regulation provides laboratories the flexibility to establish a system that ensures positive patient identification through specimen collection, labeling, accessioning, processing, (e.g., aliquotting), storage, testing, and reporting of results. Review the laboratory's system (policy and practices) for ensuring positive patient identification from specimen collection through reporting of results.

Optimum integrity of a patient's specimen should be determined according to the test methodology utilized by the laboratory. Review manufacturer's instructions for performance of each test method to ensure the specimen is appropriate for the test system, is stored and preserved properly (e.g., maintained at room temperature, kept on ice, separated and frozen or refrigerated), and analyzed within the limitations of the test methodology. For specimen integrity problems in the preanalytic system, see also D5311.

The laboratory must have a procedure to ensure that special handling of specimens is maintained throughout the testing process when necessary (e.g., GC cultures and GC/Chlamydia probes, blood gas specimens, and DNA probes).

Probes §493.1232

How does the laboratory ensure positive identification of patient specimens through all phases of testing, especially when similar patient identification information (e.g., address, sex, names, timed specimens, and birth dates) exists?

How does the laboratory ensure that special handling of specimens (as specified in the laboratory's procedure manual) is maintained throughout the testing process?

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Does the laboratory process patient specimens using separate (distinct) or unique identifiers in order to avoid mislabeling, specimen mix-ups, incorrect test request entry, and incorrect reporting of results?

**FED - D5205 - COMPLAINT INVESTIGATIONS**

**Title** COMPLAINT INVESTIGATIONS

**CFR** 493.1233

**Type** Standard

**Regulation Definition**

The laboratory must have a system in place to ensure that it documents all complaints and problems reported to the laboratory. The laboratory must conduct investigations of complaints, when appropriate.

**Interpretive Guideline**

Interpretative Guidelines §493.1233

Verify that the laboratory documents all complaints and problems reported to the laboratory, and that it has a mechanism to determine which complaints require investigation.

Probes §493.1233

What mechanism does the laboratory have that allows individuals to report complaints or problems to the laboratory?

How does the laboratory inform laboratory personnel of mechanisms to anonymously report complaints about laboratory quality to outside agencies, e.g. State survey agencies?

Does the laboratory have a mechanism to refer complaints or problems to its reference laboratory(s), or other offices or agencies, when appropriate? Does the laboratory document this activity?

**FED - D5207 - COMMUNICATIONS**

**Title** COMMUNICATIONS

**CFR** 493.1234

**Type** Standard

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**Regulation Definition**

The laboratory must have a system in place to identify and document problems that occur as a result of a breakdown in communication between the laboratory and an authorized person who orders or receives test results.

**Interpretive Guideline**

Interpretative Guidelines §493.1234

Such communication could entail problems with the descriptions they have provided to authorized individuals about proper specimen collection or shipment. For example, the laboratory's system for identifying and documenting communication problems should be able to capture instances in which there is a need to request additional information concerning patient specimens. If the laboratory does not receive the appropriate specimen or patient information needed to perform the tests, the laboratory should assess whether the information that is currently being made available to authorized individuals concerning patient preparation and specimen handling requirements is adequate.

The laboratory's system for identifying and documenting communications problems should be able to capture instances where testing was affected. These instances could be due to the lack of necessary patient information from the authorized person, improper specimen collection, improper handling and transport of the specimens to the laboratory, etc. If the appropriate specimen(s) and/or patient information needed to perform the requested tests is not being received by the laboratory, an assessment should be made to determine whether the information that is currently made available to authorized persons concerning patient preparation, specimen collection and handling requirements, is adequate.

**FED - D5209 - PERSONNEL COMPETENCY ASSESSMENT POLICIES**

**Title** PERSONNEL COMPETENCY ASSESSMENT POLICIES

**CFR** 493.1235

**Type** Standard

**Regulation Definition**

As specified in the personnel requirements in subpart M, the laboratory must establish and follow written policies and procedures to assess employee and, if applicable, consultant competency.

**Interpretive Guideline**

Interpretative Guidelines §493.1235

Refer to §§493.1413(b)(8) and 493.1451(b)(8) for specific testing personnel competency requirements and refer to §493.1407(e)(12) and §493.1445(e)(13) for establishing policies to monitor each individual's competency and identify remedial training or continuing education needs. Cite deficiencies at this location when the laboratory has developed but is not following personnel competency policies and procedures for technical and clinical consultants, technical supervisors and other laboratory staff.

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Note: If the laboratory director is the only individual testing and reporting test results, they must establish and document a minimal level of efficiency in order to ensure that they maintain the required competency for accurate and reliable testing and reporting.

Competency Assessment Guidelines

Technical consultant, clinical consultant, technical supervisor, general supervisor

Documented competency assessment is required for the following named positions on the Form 209: technical consultant, clinical consultant, technical supervisor, general supervisor. The laboratory must have policies and procedures to assess competency based on the position responsibilities listed in Subpart M and these assessments must be performed at a frequency determined by the laboratory. Cite D5209 (§493.1235). If these people perform testing on patient specimens, they are required to have the six required procedures in their competency assessment in addition to a competency assessment based on their federal regulatory responsibilities.

Testing personnel in laboratories with a PPMP certificate

Testing personnel in PPMP laboratories, including mid-level practitioners, are required to undergo competency assessment that includes the six procedure found in §493.1413(b)(8). Use D5209 or appropriate Technical Consultant D-tag (D6046 through D6052) relating to competency assessment.

preanalyticpostanalytic  
ensuring

KEY POINT: In situations in which more than one citation may be used, choose the one that is most specific to the situation. This will best allow the laboratory to understand the problem and correct it.

Probes §493.1235

How does the laboratory evaluate the competency of its employees?

If the laboratory uses non-testing personnel to perform preanalytic functions how does it ensure their competency?

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If a laboratory utilizes a consultant, how does the laboratory determine if the consultant is competent? Does the laboratory have a policy/procedure to determine consultant competency? Use D6030 or D6103.

How does the laboratory evaluate personnel for consistency in slide review (e.g., ANA, manual differential, urine sediment)?

**FED - D5211 - EVALUATION OF PROFICIENCY TESTING PERFORMANCE**

**Title** EVALUATION OF PROFICIENCY TESTING PERFORMANCE

**CFR** 493.1236(a)

**Type** Standard

**Regulation Definition**

The laboratory must review and evaluate the results obtained on proficiency testing performed as specified in subpart H of this part.

**Interpretive Guideline**

Probes §493.1236(a)

Is there evidence of review and evaluation of the laboratory's proficiency testing (PT) results?

**FED - D5213 - EVALUATION OF PROFICIENCY TESTING PERFORMANCE**

**Title** EVALUATION OF PROFICIENCY TESTING PERFORMANCE

**CFR** 493.1236(b)(1)

**Type** Standard

**Regulation Definition**

The laboratory must verify the accuracy of any analyte or subspecialty without analytes listed in subpart I of this part that is not evaluated or scored by a CMS-approved proficiency testing program.

**Interpretive Guideline**

Interpretative Guideline §493.1236(b)(1)

NOTE: An analyte submitted to a PT program for evaluation may not be evaluated or scored by the PT program if there are less than 10 participants in a particular peer group (§§493.909 - 493.959).

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**FED - D5215 - EVALUATION OF PROFICIENCY TESTING PERFORMANCE**

**Title** EVALUATION OF PROFICIENCY TESTING PERFORMANCE

**CFR** 493.1236(b)(2)

**Type** Standard

**Regulation Definition**

The laboratory must verify the accuracy of any analyte, specialty or subspecialty assigned a proficiency testing score that does not reflect laboratory test performance (that is, when the proficiency testing program does not obtain the agreement required for scoring as specified in subpart I of this part, or the laboratory receives a zero score for nonparticipation, or late return or results).

**Interpretive Guideline**

Interpretative Guidelines §493.1236(b)(2)

The laboratory must have a mechanism for routine review of its proficiency testing results that are evaluated by its PT providers. This includes a review of its actual PT results against the PT provider's participant summary results for the particular PT event and when any of the following occur:

- o The PT program assigned an artificial score of 100% (e.g., results not evaluated or scored);
- o A zero score for nonparticipation; if the laboratory did not test the specimen, it must document what other means were used to assess the accuracy of the test for the PT event that was missed; or
- o The PT provider notifies the laboratory that its results were not evaluated (given a score of "0") due to missing the return deadline.

Probes §493.1236(b)(2)

Has the laboratory reviewed its test menu to determine if it tests any analyte(s) that are not listed in subpart I?

**FED - D5217 - EVALUATION OF PROFICIENCY TESTING PERFORMANCE**

**Title** EVALUATION OF PROFICIENCY TESTING PERFORMANCE

**CFR** 493.1236(c)(1)

**Type** Standard

**Regulation Definition**

At least twice annually, the laboratory must verify the

**Interpretive Guideline**

Interpretative Guidelines §493.1236(c)(1)

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accuracy of any test or procedure it performs that is not included in subpart I of this part.

Refer to subpart I, Proficiency Testing Programs for Nonwaived Testing. Subpart I includes those specialties, subspecialties, analytes and tests that are considered regulated tests. For those tests not listed in subpart I (not regulated), the laboratory must verify the accuracy of the test or procedure twice annually, including the accuracy of calculated results, if applicable.

For those tests not listed under Subpart I, the laboratory may enroll in a PT program to verify the accuracy of their test or procedure. However, under no circumstances may these PT samples be referred (or results communicated) to another laboratory for any reason prior to the PT testing event cut-off date. The PT referral consequences (loss of certificate and bar on owner operator) apply equally to all PT testing samples and results. (See D2013).

**FED - D5219 - EVALUATION OF PROFICIENCY TESTING PERFORMANCE**

**Title** EVALUATION OF PROFICIENCY TESTING PERFORMANCE

**CFR** 493.1236(c)(2)

**Type** Standard

**Regulation Definition**

At least twice annually, the laboratory must verify the accuracy of any test or procedure listed in subpart I of this part for which compatible proficiency testing samples are not offered by a CMS-approved proficiency testing program.

**Interpretive Guideline**

Interpretative Guidelines §493.1236(c)(2)

Laboratory tests or procedures that are not compatible may include new or emerging technologies for which PT is not yet available.

Probes §493.1236(c)(2)

How does the laboratory verify accuracy of tests not included under subpart I or tests for which compatible PT samples are not available (e.g., blind testing of materials with known values, other external assessment programs, split samples with another laboratory instrument or method, comparison with Kodachrome slides from a reference source)?

**FED - D5221 - EVALUATION OF PROFICIENCY TESTING PERFORMANCE**

**Title** EVALUATION OF PROFICIENCY TESTING PERFORMANCE

**CFR** 493.1236(d)

**Type** Standard

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**Regulation Definition**

All proficiency testing evaluation and verification activities must be documented.

**Interpretive Guideline**

Interpretative Guidelines §493.1236(d)  
Documentation must include review of all unsatisfactory scores and the corrective action taken.

**FED - D5291 - GENERAL LABORATORY SYSTEMS QUALITY ASSESSMENT**

**Title** GENERAL LABORATORY SYSTEMS QUALITY ASSESSMENT

**CFR** 493.1239(a)

**Type** Standard

**Regulation Definition**

The laboratory must establish and follow written policies and procedures for an ongoing mechanism to monitor, assess, and, when indicated, correct problems identified in the general laboratory systems requirements specified at §§493.1231 through 493.1236.

**Interpretive Guideline**

Interpretative Guidelines §493.1239(a)-(c)

Quality Assessment (QA) is an ongoing review process that encompasses all facets of the laboratory's technical and non-technical functions and all locations/sites where testing is performed. QA also extends to the laboratory's interactions with and responsibilities to patients, physicians, and other laboratories ordering tests, and other non-laboratory areas or departments of the facility of which it is a part.

When the laboratory discovers an error or identifies a potential problem, actions must be taken to correct the situation. This correction process involves identification and resolution of the problem, and development of policies that will prevent recurrence. Policies for preventing problems that have been identified must be written as well as communicated to the laboratory personnel and other staff, clients, etc., as appropriate. Over time, the laboratory must monitor the corrective action(s) to ensure the action(s) taken have prevented recurrence of the original problem.

All pertinent laboratory staff must be involved in the assessment process through discussions or active participation.

QA of the General Laboratory System includes assessing practices/issues related to:

- o Patient confidentiality;
- o Specimen identification and integrity;
- o Complaint investigations;

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- o Communications;
- o Personnel competency; and
- o Proficiency testing performance.

An example would be monitoring the type and number of complaints received by the laboratory such as a particular client continuously complaining about the laboratory's failure to promptly respond to STAT test requests. The laboratory must have documentation that the complaint was investigated and appropriate action taken to correct the problem.

Verify that the laboratory has a system in place for monitoring and evaluating confidentiality of patient information.

Probes §493.1239(a)

How does the laboratory ensure that an individual who had problems in performance is competent after appropriate retraining and technical assistance is completed?

How does the laboratory determine which complaints require investigation and which do not?

**FED - D5293 - GENERAL LABORATORY SYSTEMS QUALITY ASSESSMENT**

**Title** GENERAL LABORATORY SYSTEMS QUALITY ASSESSMENT

**CFR** 493.1239(b)(c)

**Type** Standard

**Regulation Definition**

(b) The general laboratory systems quality assessment must include a review of the effectiveness of corrective actions taken to resolve problems, revision of policies and procedures necessary to prevent recurrence of problems, and discussion of general laboratory systems quality assessment reviews with appropriate staff.

(c) The laboratory must document all general laboratory systems quality assessment activities.

**Interpretive Guideline**

Interpretative Guidelines §493.1239(b)

Review assessment policies, procedures and reports to verify that the laboratory has a system in place to ensure continuous improvement. Corrective action reports are one indication that the laboratory is monitoring and evaluating laboratory performance and the quality of services.

Probes §493.1239(b)

When problems are identified in personnel competency, what corrective actions are instituted to improve employee

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performance?

When the laboratory identifies a problem, are corrective actions taken? Are these actions documented and monitored for effectiveness?

How does the laboratory prevent reoccurrences of problems?

How does the laboratory identify and document potential communication problems and any corrective actions that are taken (e.g., with staff, referral laboratories)?

Have the corrective actions that were taken as a result of failures in proficiency testing (PT) and/or verification of accuracy testing (as required under subpart H) improved performance?

Interpretative Guidelines §493.1239(c)

Laboratories must document the steps taken to identify and correct problems, and any efforts to prevent recurrences. This includes laboratory policies amended due to QA activities.

**FED - D5300 - PREANALYTIC SYSTEMS**

**Title** PREANALYTIC SYSTEMS

**CFR** 493.1240

**Type** Condition

**Regulation Definition**

Each laboratory that performs nonwaived testing must meet the applicable preanalytic system(s) requirements in §§493.1241 and 493.1242, unless HHS approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub. 7), that provides equivalent quality testing. The laboratory must monitor and evaluate the overall quality of the preanalytic systems and correct identified problems as specified in §493.1249 for each specialty and subspecialty of testing performed.

**Interpretive Guideline**

Interpretative Guidelines §493.1240

Preanalytic refers to all steps taken prior to the actual testing of a patient specimen from the test request to the actual testing of the specimen. The preanalytic systems requirements include three distinct sections: test requests; specimen submission, handling, and referral; and preanalytic systems quality assessment.

Significant deficiencies cited under this condition may indicate deficiencies under personnel responsibilities. Use D5300 when deficiencies are identified that have the potential to, or adversely affect patient testing, are systemic and pervasive throughout the laboratory, and are not limited to any one specialty or subspecialty.

To determine which tests are categorized as waived or nonwaived testing (i.e., moderate and high complexity tests),

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refer to the "Specific List For Categorization of Laboratory Test Systems, Assays, and Examinations by Complexity" [[www.fda.gov/cdrh/cliaindex.html](http://www.fda.gov/cdrh/cliaindex.html)]. Test systems, assays and examinations not included in this listing (i.e., not yet categorized) are considered high complexity.

**FED - D5301 - TEST REQUEST**

**Title** TEST REQUEST

**CFR** 493.1241(a)

**Type** Standard

**Regulation Definition**

The laboratory must have a written or electronic request for patient testing from an authorized person.

**Interpretive Guideline**

Interpretative Guidelines §493.1241(a)

An "authorized person" means an individual authorized under State law to order tests or receive test results, or both.

See D5305 for specific guidance on the CLIA requirements for the test requisition process.

To ensure that an authorized person is ordering the test, a laboratory using electronic test requests may issue passwords.

Written policies should cover the use of standing orders. Such policies should clearly define which tests may be covered by standing orders and at what interval standing orders should be reconfirmed.

**FED - D5303 - TEST REQUEST**

**Title** TEST REQUEST

**CFR** 493.1241(b)

**Type** Standard

**Regulation Definition**

The laboratory may accept oral requests for laboratory tests if it solicits a written or electronic authorization within 30 days of the oral request and maintains the authorization or documentation of its efforts to obtain the authorization.

**Interpretive Guideline**

Interpretative Guidelines §493.1241(b)

Review the laboratory's policy for requesting written orders within 30 days of the oral requests. If no written order was received, verify the laboratory has documentation showing the attempt.

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**FED - D5305 - TEST REQUEST**

**Title** TEST REQUEST

**CFR** 493.1241(c)

**Type** Standard

**Regulation Definition**

The laboratory must ensure the test requisition solicits the following information:

- (1) The name and address or other suitable identifiers of the authorized person requesting the test and, if appropriate, the individual responsible for using the test results, or the name and address of the laboratory submitting the specimen, including, as applicable, a contact person to enable the reporting of imminently life threatening laboratory results or panic or alert values.
- (2) The patient's name or unique patient identifier.
- (3) The sex and age or date of birth of the patient.
- (4) The test(s) to be performed.
- (5) The source of the specimen, when appropriate.
- (6) The date and, if appropriate, time of specimen collection.
- (7) For Pap smears, the patient's last menstrual period, and indication of whether the patient had a previous abnormal report, treatment, or biopsy.
- (8) Any additional information relevant and necessary for a specific test to ensure accurate and timely testing and reporting of results, including interpretation, if applicable.

**Interpretive Guideline**

Interpretative Guidelines §493.1241(c)(1)-(c)(8)

The test requisition must provide the information necessary to identify and send test results to the individual who ordered the test (the authorized person), or, where applicable, to the authorized person's representative. An authorized person may also use the test requisition to designate additional individuals/entities that will be responsible for using the test results to provide care to the subject individual.

The address(es) to which test results should be sent may include a postal address (street, city or town, state and zip code), a fax number, and/or the information necessary for electronic transmission. When appropriate, a telephone number or other mechanism to contact the individual responsible for using the test results should be provided to the laboratory on the requisition.

Verify that test requisitions solicit all information necessary for the proper interpretation of results. This may include patient's age, sex, date, fasting status, time of collection, specimen type (e.g., plasma, urine, spinal fluid), diagnosis, and date of last menstrual period (LMP) for Papanicolaou (PAP) smears. Verify that the instructions to clients are clear and specify the items that must be completed.

Laboratories must have policies that guide staff on what to do if/when they receive a requisition or patient medical chart or record that is missing required information.. Laboratories must either obtain the missing information or report results and indicate on the test report, medical record or chart any limitations of test results due to the omission of patient information. If the missing information is essential (such as the family history for certain genetic tests) for accurate test results, it must be obtained prior to reporting patient test results.

Interpretative Guidelines §493.1241(c)(8)

This may include such items as preventative or therapeutic medications, or family history.

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Probes §493.1241(c)(1)-(c)(8)

How does the laboratory uniquely identify patient specimens that share the same or similar name, birth date, address or sex?

How does the requisition provide for inclusion of additional information when necessary (e.g., specimen type or source)?

**FED - D5307 - TEST REQUEST**

**Title** TEST REQUEST

**CFR** 493.1241(d)

**Type** Standard

**Regulation Definition**

The patient's chart or medical record may be used as the test requisition or authorization but must be available to the laboratory at the time of testing and available to CMS or a CMS agent upon request.

**Interpretive Guideline**

Probes §493.1241(d)

When the patient's chart or medical record is used as the test requisition, does it provide all the information necessary to ensure accurate testing and reporting of results?

**FED - D5309 - TEST REQUEST**

**Title** TEST REQUEST

**CFR** 493.1241(e)

**Type** Standard

**Regulation Definition**

If the laboratory transcribes or enters test requisition or authorization information into a record system or a laboratory information system, the laboratory must ensure the information is transcribed or entered accurately.

**Interpretive Guideline**

Interpretative Guidelines §493.1241(e)

The laboratory must have an ongoing mechanism to ensure the accuracy of manual entries by personnel into an LIS.

How does the laboratory ensure that all individuals who enter data, including clerical staff, correctly match patient information?

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**FED - D5311 - SPECIMEN SUBMISSION, HANDLING, AND REFERRAL**

**Title** SPECIMEN SUBMISSION, HANDLING, AND REFERRAL

**CFR** 493.1242(a)

**Type** Standard

**Regulation Definition**

The laboratory must establish and follow written policies and procedures for each of the following, if applicable:

- (1) Patient preparation.
- (2) Specimen collection.
- (3) Specimen labeling, including patient name or unique patient identifier and, when appropriate, specimen source.
- (4) Specimen storage and preservation.
- (5) Conditions for specimen transportation.
- (6) Specimen processing.
- (7) Specimen acceptability and rejection.
- (8) Specimen referral.

**Interpretive Guideline**

Probes §493.1242(a)(1)

How does the laboratory ensure that all staff, including phlebotomists, gives appropriate instructions for patient preparation when needed?

Does the laboratory provide instructions directly to patients or to the client when proper patient preparation is required for optimal specimen collection? For example:

- o Proper preservation (temperature) and transportation time of semen specimens;
- o Fasting instructions for lipid profile testing;
- o Dietary restrictions prior to occult blood testing;
- o Twenty-four hour urine collection for specific tests; and
- o Fasting and two hour post-prandial glucose collections.

If a patient has special communication needs (hearing impaired, not fluent in English etc.), are resources available to the client or to the patient, as appropriate, to ensure that instructions for specimen collection, preservation, and transportation to the laboratory, are properly understood?

Has the laboratory provided to its staff and/or individuals external to the laboratory who collect specimens, written procedures to ensure that patient preparation requirements have been followed?

Interpretative Guidelines §493.1242(a)(2)

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Verify that procedures are available to the appropriate staff responsible for collecting the correct specimen, that personnel are using the appropriate collection technique (order and site of draw) and proper containers (e.g., acceptable anti-coagulant, sterile containers for culture specimens, dacron swabs vs. cotton swabs).

Interpretative Guidelines §493.1242(a)(3)

If the laboratory receives two specimens simultaneously with the same first and last name or birth date, the laboratory must have a system in place to process these specimens using distinct identifying indicators in order to distinguish between the specimens. This also pertains to personnel collecting and labeling specimens. This may include a system that involves labeling the specimen container and request slip (or the patient's medical record or chart) with a unique patient identification number, but does not preclude the use of other mechanisms to assist in patient identification and tracking of specimens throughout the collection, accessioning, testing, and reporting processes.

Interpretative Guidelines §493.1242(a)(4)

Review manufacturer's instructions for performance of each test method to ensure that specimens are properly stored (e.g., maintained at room temperature, kept refrigerated after separation, separated and frozen).

Probes §493.1242(a)(4)

What instructions are provided for specimen preservation and transportation, when applicable? For example:

- o Sputum for Cytology;
- o Specimens for parathyroid hormone;
- o Specimens for blood gas analysis;
- o Specimens for urine culture and colony count; and
- o Specimens for 24 hour urine collections requiring preservatives.

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Probes §493.1242(a)(5)

Does the laboratory follow the manufacturer's or the referral laboratory's instructions, as appropriate, for transport of specimens?

Interpretative Guidelines §493.1242(a)(6)

Specimen processing may include receiving the specimen, accessioning the specimen, preparing the specimen for in-house analysis, preparation to send to a reference laboratory, preparing slides, and inoculating primary culture media, etc. Specimen processing also includes: Parasitology: the fixation and concentration of specimens; Virology: the pretreatment of specimens with antibiotics, the manipulation of cell culture tubes and inoculation of the cell cultures prior to incubation; Mycobacteriology: performing digestion-decontamination and concentration procedures on clinical specimens; and Histopathology: specimen accession with or without fixation, embedding the paraffin block, cutting the paraffin block, mounting the embedded cut tissue to a slide, preparing the slide for staining, staining and cover slipping the slide, or any other slide preparation procedures that do not involve examination resulting in diagnostic interpretation.

Note: for histopathology specimens, specimen processing does not constitute a CLIA test. Only gross examinations (including weighing, measuring, describing color, specific orientation for diagnostic interpretation, and other characteristics of the tissue, or performing other mechanical procedures including dissection, inking, and marking) require a CLIA certificate. Microscopic examinations of tissue with diagnostic interpretation and reporting is a Histopathology test and requires CLIA certification.

Probes §493.1242(a)(6)

What policies or systems does the laboratory have in place to differentiate specimens that have similar names or identification information?

How does the laboratory recognize and process timed patient specimens (e.g., peaks and troughs)?

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Interpretative Guidelines §493.1242(a)(7)

Criteria for specimen acceptability and rejection must include the disposition of the rejected specimen(s). Use D5805. The laboratory should promptly notify the authorized person when a specimen meets its rejection criteria and is unsuitable for testing.

Interpretative Guidelines §493.1242(a)(8)

Ensure that the laboratory has a current service manual available for each reference laboratory that it uses that contains the reference laboratory's specimen requirements for the test to be performed.

Probes §493.1242(a)(8)

Are laboratory personnel familiar with procedures to prepare and/or submit specimens to the appropriate reference laboratory?

How does the laboratory ensure the security and preservation of specimens submitted to their reference laboratory (e.g., if the office closes before the arrival of the reference laboratory's courier)? How does the laboratory ensure a timely pick-up of specimens to be performed at the referral laboratory?

**FED - D5313 - SPECIMEN SUBMISSION, HANDLING, AND REFERRAL**

**Title** SPECIMEN SUBMISSION, HANDLING, AND REFERRAL

**CFR** 493.1242(b)

**Type** Standard

**Regulation Definition**

The laboratory must document the date and time it receives a specimen.

**Interpretive Guideline**

Interpretative Guidelines §493.1242(b)

When a sample is collected and a test is performed during the course of a patient's visit, the date and time recorded in the patient "sign-in" log may be used as the date and time of receipt into the laboratory.

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**FED - D5315 - SPECIMEN SUBMISSION, HANDLING, AND REFERRAL**

**Title** SPECIMEN SUBMISSION, HANDLING, AND REFERRAL

**CFR** 493.1242(c)

**Type** Standard

**Regulation Definition**

The laboratory must refer a specimen for testing only to a CLIA-certified laboratory or a laboratory meeting equivalent requirements as determined by CMS.

**Interpretive Guideline**

Interpretative Guidelines §493.1242(c)

Some examples of laboratories meeting equivalent requirements are those of the Veterans Administration (VHA), the Department of Defense (DOD) facilities, and CLIA-exempt laboratories.

Probes §493.1242(c)

How does the laboratory ensure that the reference laboratory has and maintains a current CLIA certificate?

**FED - D5317 - SPECIMEN SUBMISSION, HANDLING, AND REFERRAL**

**Title** SPECIMEN SUBMISSION, HANDLING, AND REFERRAL

**CFR** 493.1242(d)

**Type** Standard

**Regulation Definition**

If the laboratory accepts a referral specimen, written instructions must be available to the laboratory's clients and must include, as appropriate, the information specified in paragraphs (a)(1) through (a)(7) of this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1242(d)

Ensure the laboratory has provided written instructions to each client that sends specimens/test requests. The instructions may contain information on specimen handling (e.g., collection, preservation, storage, transport, testing schedule times and how to obtain additional assistance for unusual circumstances).

**FED - D5391 - PREANALYTIC SYSTEMS QUALITY ASSESSMENT**

**Title** PREANALYTIC SYSTEMS QUALITY ASSESSMENT

**CFR** 493.1249(a)

**Type** Standard

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**Regulation Definition**

The laboratory must establish and follow written policies and procedures for an ongoing mechanism to monitor, assess, and when indicated, correct problems identified in the preanalytic systems specified at §§493.1241 through 493.1242.

**Interpretive Guideline**

Interpretative Guidelines §493.1249(a)-(c)

Quality Assessment (QA) is an ongoing review process that encompasses all facets of the laboratory's technical and non-technical functions and all locations/sites where testing is performed. QA also extends to the laboratory's interactions with and responsibilities to patients, physicians, and other laboratories ordering tests, and the other non-laboratory areas or departments of the facility of which it is a part.

When the laboratory discovers an error or identifies a potential problem, actions must be taken to correct the situation. This correction process involves identification and resolution of the problem, and development of policies that will prevent recurrence. Policies for preventing problems that have been identified must be written as well as communicated to the laboratory personnel and other staff, clients, etc., as appropriate. Over time, the laboratory must monitor the corrective action(s) to ensure the action(s) taken have prevented recurrence of the original problem. All pertinent laboratory staff must be involved in the assessment process through discussions or active participation.

QA of the Preanalytic System includes assessing practices/issues related to test requests, specimen submission, handling and referral.

Some examples include: monitoring the frequency of specimen handling problems (such as the use of an improper blood collection tube, inadequate mixing of blood specimens with anticoagulant after collection), and delays in specimen transport; identifying clients who repeatedly refer unacceptable specimens or improperly complete requisition forms and documentation of its efforts to reduce the recurrence of these problems.

Review assessment policies, procedures and reports to verify that the laboratory has a system in place to ensure continuous improvement. Corrective action reports are one indication that the laboratory is monitoring and evaluating laboratory performance and the quality of services.

Probes §493.1249(a)-(c)

When a laboratory uses off-site drawing facilities, what policies or procedures does the laboratory use to ensure proper accountability or tracking of patient specimens from time of collection to receipt by the laboratory performing the tests?

Does the laboratory perform periodic or spot checks for accurate transfer of information (e.g., manual entries by personnel from test orders to test requisition or into an LIS)? For referral specimens, how does the laboratory check for transcription errors when patient test information is transcribed from the laboratory's original requisition form to the reference laboratory's requisition?

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What actions does the laboratory take if test requisitions from one or more clients are consistently incomplete, illegible or contain incorrect information?

What actions does the laboratory take if specimens received from one client are consistently unsatisfactory for testing (e.g., specimens for Cytology)? Has the laboratory's efforts to reduce the recurrence of these problems been documented and effective?

**FED - D5393 - PREANALYTIC SYSTEMS QUALITY ASSESSMENT**

**Title** PREANALYTIC SYSTEMS QUALITY ASSESSMENT

**CFR** 493.1249(b)(c)

**Type** Standard

**Regulation Definition**

The preanalytic systems assessment must include a review of the effectiveness of corrective actions taken to resolve problems, revision of policies and procedures necessary to prevent recurrence of problems, and discussion of preanalytic systems quality assessment reviews with appropriate staff. The laboratory must document all preanalytic systems quality assessment activities.

**Interpretive Guideline**

Interpretative Guidelines §493.1249(c)

The steps taken by the laboratory to identify and correct problems and prevent their recurrence must be documented. All laboratory policies amended due to its QA activities must also be noted.

**FED - D5400 - ANALYTIC SYSTEMS**

**Title** ANALYTIC SYSTEMS

**CFR** 493.1250

**Type** Condition

**Regulation Definition**

Each laboratory that performs nonwaived testing must meet the applicable analytic systems requirements in §§493.1251 through 493.1283, unless HHS approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub.7), that provides equivalent quality testing. The

**Interpretive Guideline**

Interpretative Guidelines §493.1250

Significant deficiencies cited under this condition may indicate deficiencies under personnel. Use D5400 when deficiencies are identified that are significant and have the potential to, or adversely affect patient testing, are systemic and pervasive throughout the laboratory, and are not limited to any one specialty or

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laboratory must monitor and evaluate the overall quality of the analytic systems and correct identified problems as specified in §493.1289 for each specialty and subspecialty of testing performed.

subspecialty.

Refer to §§493.1261 - 493.1278 for additional requirements for Bacteriology, Mycobacteriology, Mycology, Parasitology, Virology, Routine Chemistry, Hematology, Immunohematology, Histopathology, Cytology, Clinical Cytogenetics, and Histocompatibility.

**FED - D5401 - PROCEDURE MANUAL**

**Title** PROCEDURE MANUAL

**CFR** 493.1251(a)

**Type** Standard

**Regulation Definition**

A written procedures manual for all tests, assays, and examinations performed by the laboratory must be available to, and followed by, laboratory personnel. Textbooks may supplement but not replace the laboratory's written procedures for testing or examining specimens.

**Interpretive Guideline**

Interpretative Guidelines §493.1251(a)

Procedures may be organized in the form of paper-based manuals, or a manual that is stored in and accessed through computers and/or card files. Use D5403, if the procedure manual lacks any of the applicable information as specified in §493.1251(b)(1)-(14). If the laboratory has procedures that are not used for test performance, but are used for reference purposes, they may be placed in a reference section. You need not review reference procedures unless problems are identified with patient test results.

Centers for Disease Control and Prevention (CDC) and Armed Forces Institute of Pathology (AFIP) manuals, manufacturer's operating instructions, and package inserts, are acceptable provided the policies and procedures are available, and the methods in use are clearly indicated. If the laboratory modifies any procedure, the modification must be documented and verified/established as specified in §493.1253.

Probes §493.1251(a)

How does the laboratory ensure that personnel follow the procedures in the procedure manual? How are changes in procedures communicated to laboratory personnel? For competency issues, use D6030 or D6103 as applicable.

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**FED - D5403 - PROCEDURE MANUAL**

**Title** PROCEDURE MANUAL

**CFR** 493.1251(b)

**Type** Standard

**Regulation Definition**

The procedure manual must include the following when applicable to the test procedure:

- (1) Requirements for patient preparation; specimen collection, labeling, storage, preservation, transportation, processing, and referral; and criteria for specimen acceptability and rejection as described in §493.1242.
- (2) Microscopic examination, including the detection of inadequately prepared slides.
- (3) Step-by-step performance of the procedure, including test calculations and interpretation of results.
- (4) Preparation of slides, solutions, calibrators, controls, reagents, stains, and other materials used in testing.
- (5) Calibration and calibration verification procedures.
- (6) The reportable range for test results for the test system as established or verified in §493.1253.
- (7) Control procedures.
- (8) Corrective action to take when calibration or control results fail to meet the laboratory's criteria for acceptability.
- (9) Limitations in the test methodology, including interfering substances.
- (10) Reference intervals (normal values).
- (11) Imminently life-threatening test results, or panic or alert values.
- (12) Pertinent literature references.
- (13) The laboratory's system for entering results in the patient record and reporting patient results including, when appropriate, the protocol for reporting imminently life threatening results, or panic, or alert values.

**Interpretive Guideline**

Interpretative Guidelines §493.1251(b)(1)

If testing is delayed or not performed daily, specimens must be preserved or stored in accordance with the laboratory's procedures to ensure specimen integrity.

Determine if the laboratory has a procedure for handling and identifying aliquotted specimens; e.g., sputum sent for Mycobacteriology and Cytology examinations; stool specimens for occult blood, routine culture, parasitology and C. difficile toxin assay; and cerebrospinal fluids for cell count, culture, glucose and protein.

Interpretative Guidelines §493.1251(b)(5)

Calibration and calibration verification procedures must be established in accordance with §493.1255.

Interpretative Guidelines §493.1251(b)(7)

Determine if the laboratory's quality control procedures include the following:

- o Type of control (e.g., manufacturer or in-house, electronic);
- o Identity (e.g., normal, abnormal, level I, II, patient or a control);
- o Number and frequency of testing controls;
- o Control limits established in accordance with §§493.1253 and 493.1256; and

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(14) Description of the course of action to take if a test system becomes inoperable.

o Criteria to determine acceptable control results.

Interpretative Guidelines §493.1251(b)(8)

Ensure that corrective action procedures are established in accordance with §493.1282(b)(2).

Interpretative Guidelines §493.1251(b)(13)

Ensure the procedure manual provides instructions for reporting the patient's test results in the appropriate units or terminology. Use D5805.

Probes §493.1251(b)(13)

Do laboratory procedures address the process for reporting (oral and written) results on patients with multiple laboratory encounters to ensure that the exact name, date, time and identification of specimen is conveyed to the authorized person?

Interpretative Guidelines §493.1251(b)(14)

Laboratory information systems (LIS) procedures must be available to operators. Instructions should identify the individual(s), either by name or position, to notify if the LIS goes down or if a system error occurs.

Probes §493.1251(b)(14)

When the primary testing system is inoperable, what procedure does the laboratory use to bring the backup system on line?

**FED - D5405 - PROCEDURE MANUAL**

**Title** PROCEDURE MANUAL

**CFR** 493.1251(c)

**Type** Standard

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**Regulation Definition**

Manufacturer's test system instructions or operator manuals may be used, when applicable, to meet the requirements of paragraphs (b)(1) through (b)(12) of this section. Any of the items under paragraphs (b)(1) through (b)(12) of this section not provided by the manufacturer must be provided by the laboratory.

**Interpretive Guideline**

**FED - D5407 - PROCEDURE MANUAL**

**Title** PROCEDURE MANUAL

**CFR** 493.1251(d)

**Type** Standard

**Regulation Definition**

Procedures and changes in procedures must be approved, signed, and dated by the current laboratory director before use.

**Interpretive Guideline**

Interpretative Guidelines §493.1251(d)

Verify that the methods in the procedure manual are current for tests offered by the laboratory (e.g., reagent test kits and instruments used in the laboratory correlate with methods in the procedure manual).

All laboratory procedures including CDC and AFIP manuals, manufacturer's operator manuals, and package inserts must reflect the director's review and approval including any modifications in the procedure.

Approval of procedures and changes to procedures is the responsibility of the laboratory director. This responsibility cannot be delegated. A coversheet may be used for the director to approve the manual. Annual review of procedures is not required.

**FED - D5409 - PROCEDURE MANUAL**

**Title** PROCEDURE MANUAL

**CFR** 493.1251(e)

**Type** Standard

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**Regulation Definition**

The laboratory must maintain a copy of each procedure with the dates of initial use and discontinuance as described in §493.1105(a)(2).

**Interpretive Guideline**

**FED - D5411 - TEST SYSTEMS, EQUIPMENT, INSTRUMENTS, REAGENT**

**Title** TEST SYSTEMS, EQUIPMENT, INSTRUMENTS, REAGENT

**CFR** 493.1252(a)

**Type** Standard

**Regulation Definition**

Test systems must be selected by the laboratory. The testing must be performed following the manufacturer's instructions and in a manner that provides test results within the laboratory's stated performance specifications for each test system as determined under §493.1253.

**Interpretive Guideline**

Interpretative Guidelines §493.1252(a)

The laboratory must meet any and all regulatory requirements to comply with manufacturers' recommendations and requirements for testing.

These include, but are not limited to:

- o Handling reagents, materials, and supplies;
- o Adhering to conditions for storage and testing; and
- o Performing equipment maintenance and function checks

For International Normalized Ratio (INR) calculation, ensure the laboratory:

- o Verifies that the normal patient Prothrombin mean study has been performed according to the manufacturer's instructions;
- o Periodically verifies, for each thromboplastin lot number in use, the correct normal patient Prothrombin time mean and the International Sensitivity Index (ISI) value are being used for calculating the INR value; and
- o Periodically verifies the accuracy of the INR calculation (manual, instrument or LIS).

To verify Prothrombin time testing with INR calculations:

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- o Check the accuracy of normal Prothrombin time mean calculation (manual, instrument or LIS).
- o Verify the ISI used in the calculation correlates with the ISI specified in the reagent package insert. Select an abnormal low or abnormal high Prothrombin time result and verify the calculation.

For Immunology tests such as Syphilis Serology, check for the following parameters:

- o Antigen volume;
- o Incubation time and temperature;
- o Light source;
- o Rotator speed and circumference; and
- o Conjugate titer.

Probes §493.1252(a):

Are instruments with adjustable settings appropriately set for each substance or cell to be analyzed (e.g., gamma counters, flow cytometry)?

**FED - D5413 - TEST SYSTEMS, EQUIPMENT, INSTRUMENTS, REAGENT**

**Title** TEST SYSTEMS, EQUIPMENT, INSTRUMENTS, REAGENT

**CFR** 493.1252(b)

**Type** Standard

**Regulation Definition**

The laboratory must define criteria for those conditions that are essential for proper storage of reagents and specimens, accurate and reliable test system operation, and test result reporting. The criteria must be consistent with the manufacturer's instructions, if provided. These conditions must be monitored and documented and, if applicable, include

**Interpretive Guideline**

Interpretative Guidelines §493.1252(b)

Water quality is classified by several different organizations into different reagent grades dependent on microbial content, resistivity, silicate content, and particulate matter. Each laboratory is expected to use the appropriate water quality as required for each instrument, kit, or test system. Laboratories producing water should consider parameters such as pH, silicate content, particulate matter, and bacterial and organic content in assessing water quality. These

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the following:

- (1) Water quality.
- (2) Temperature.
- (3) Humidity.
- (4) Protection of equipment and instruments from fluctuations and interruptions in electrical current that adversely affect patient test results and test reports.

parameters vary by test system and should be assessed by the laboratory for appropriateness and monitoring. Laboratories purchasing water that has already been classified are not expected to evaluate the above parameters unless specified by the manufacturer or by the laboratory in its procedure manual.

Temperature-controlled spaces, equipment, and instruments must be monitored and results documented for acceptable temperature ranges. Corrective action is needed when acceptable temperature ranges are exceeded. Use D5781 when corrective action not documented.

Continuous monitoring of temperatures by a recording thermograph is acceptable provided the data and time of use are annotated. The charts must be retained to document that temperatures were within the limits established by the laboratory.

In lieu of manual temperature recording, it is acceptable for temperatures to be maintained and monitored internally by the instrument, provided either test results are flagged or not generated when the temperature range for test performance is exceeded.

Probes §493.1252(b)(1)-(b)(4)

How does the laboratory provide special conditions when required for specimen or reagent storage?

How is room temperature and humidity monitored when necessary for test performance, proper operation of reagents, instruments, equipment, or laboratory computer systems?

When temperatures and/or humidity are outside acceptable limits, how does the laboratory rectify the problem?

How does the laboratory that moves from testing site to testing site demonstrate that the conditions necessary for quality testing are maintained?

When mobile laboratory or temporary testing site equipment is not in use (weekends, overnight) how are instruments, reagents, stains, and other solutions protected from extreme temperature fluctuations?

**FED - D5415 - TEST SYSTEMS, EQUIPMENT, INSTRUMENTS, REAGENT**

**Title** TEST SYSTEMS, EQUIPMENT, INSTRUMENTS, REAGENT

**CFR** 493.1252(c)

**Type** Standard

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**Regulation Definition**

Reagents, solutions, culture media, control materials, calibration materials, and other supplies, as appropriate, must be labeled to indicate the following:

- (1) Identity and when significant, titer, strength or concentration.
- (2) Storage requirements.
- (3) Preparation and expiration dates.
- (4) Other pertinent information required for proper use.

**Interpretive Guideline**

Interpretative Guidelines §493.1252(c)(3)

Expiration dates for test kits and/or reagents may differ due to date opened or storage conditions (e.g., refrigerator, room temperature). Verify that laboratory personnel are aware of these differences and document the appropriate expiration date.

**FED - D5417 - TEST SYSTEMS, EQUIPMENT, INSTRUMENTS, REAGENT**

**Title** TEST SYSTEMS, EQUIPMENT, INSTRUMENTS, REAGENT

**CFR** 493.1252(d)

**Type** Standard

**Regulation Definition**

Reagents, solutions, culture media, control materials, calibration materials, and other supplies must not be used when they have exceeded their expiration date, have deteriorated, or are of substandard quality.

**Interpretive Guideline**

Interpretative Guidelines §493.1252(d)

In citing deficiencies, for outdated or deteriorated materials, indicate whether these materials have been used for patient testing. Also, look for contamination, drying or other signs of deterioration. This is as important as checking expiration dates.

**FED - D5419 - TEST SYSTEMS, EQUIPMENT, INSTRUMENTS, REAGENT**

**Title** TEST SYSTEMS, EQUIPMENT, INSTRUMENTS, REAGENT

**CFR** 493.1252(e)

**Type** Standard

**Regulation Definition**

Components of reagent kits of different lot numbers must not be interchanged unless otherwise specified by the manufacturer.

**Interpretive Guideline**

Interpretative Guidelines §493.1252(e)

"Kit" means all components of a test that are packaged together.

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**FED - D5421 - ESTABLISHMENT AND VERIFICATION OF PERFORMANCE**

**Title** ESTABLISHMENT AND VERIFICATION OF PERFORMANCE

**CFR** 493.1253(b)(1)

**Type** Standard

**Regulation Definition**

Each laboratory that introduces an unmodified, FDA-cleared or approved test system must do the following before reporting patient test results:

- (1)(i) Demonstrate that it can obtain performance specifications comparable to those established by the manufacturer for the following performance characteristics:
  - (1)(i)(A) Accuracy.
  - (1)(i)(B) Precision.
  - (1)(i)(C) Reportable range of test results for the test system.
- (1)(ii) Verify that the manufacturer's reference intervals (normal values) are appropriate for the laboratory's patient population.

**Interpretive Guideline**

Interpretative Guidelines §493.1253(b)(1)

The laboratory is responsible for verifying the performance specifications of each nonwaived unmodified FDA-cleared or approved test system that it introduces, prior to reporting patient test results. The verification of method performance should provide evidence that the accuracy, precision, and reportable range of the procedure are adequate to meet the clients' needs, as determined by the laboratory director and clinical consultant. A laboratory may use the manufacturer's performance specifications as a guideline, but is responsible for verifying the manufacturer's analytical claims before initiating patient testing.

If a method was verified by someone other than the laboratory staff (e.g., manufacturer representative), the laboratory must demonstrate that this verification correlates with its in-house test performance. This may be accomplished by the laboratory testing "known" samples.

For some qualitative tests, the laboratory may verify the manufacturer's specifications by testing known positive and negative samples to ensure that the expected results are obtained. (Specimens of known quantitative value may be used to verify the accuracy of a qualitative test.)

Prior to introducing a test for routine patient testing, the laboratory must review and evaluate the verification data.

Each laboratory is responsible for determining that its performance specifications for each test system are not affected by the relocation of the laboratory or test system. (See manufacturer's package insert regarding critical requirements such as set-up, limitations, environmental conditions, etc.) When a temporary replacement (loaner) instrument is received which is identical (i.e., same make and model, and method for the same analyte) to the instrument which is being replaced, the laboratory must verify performance specifications.

If calibration material is used to verify method performance specifications, the laboratory must demonstrate that there is a minimal matrix effect and the calibration material is appropriate for verifying test system performance specifications.

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If the LIS performs any calculations to determine a laboratory result, the calculations must be verified immediately after the LIS is programmed and prior to initial calculation of patient results.

"Less than" is used for reporting test results that are below the laboratory's detection limits for an analyte. (Detection limits must be established through method verification.) "Equivalent designation" is used to report test results for those methods that yield results below a clinically significant level (e.g., for a quantitative immunology test, patient results may be clinically negative at a 1:8 titer and test results may be reported as "1:8 negative"). (The normal value is 1:8 or less.) "Greater than" is used for reporting test results that are above the laboratory's detection limits for an analyte. If patient test results exceed the laboratory's reportable range, the laboratory must report the result as greater than the highest detection limit, re-assay a diluted patient specimen and report the calculated result, or send the specimen to a reference laboratory.

Probes §493.1253(b)(1)

How does the laboratory determine if a new or revised LIS program (whether purchased or developed in-house) performs acceptably before it is integrated into routine operation?

Interpretative Guidelines §493.1253(b)(1)(i)

Laboratories may simultaneously verify multiple performance specifications by choosing appropriate samples; e.g., repeatedly test (precision) samples with known (accuracy) high and low values (reportable range). This testing should be performed among all operators on different days. In addition, for test systems of the same make and model, consider verifying performance specifications of these devices at the same time.

Interpretative Guidelines §493.1253(b)(1)(i)(A)

Accuracy- The laboratory is responsible for verifying that the method produces correct results. Verification of accuracy may be accomplished by:

- o Testing reference materials;

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- o Comparing results of tests performed by the laboratory against the results of a reference method; or
- o Comparing split sample results with results obtained from another method, which has already been shown to provide accurate results.

For qualitative methods, the laboratory must verify that a method will identify the presence/absence of the analyte.

Interpretative Guidelines §493.1253(b)(1)(i)(B)

Precision (Reproducibility) - The laboratory is responsible for verifying the precision of each test system by assessing day-to-day, run-to-run, and within-run variation, as well as operator variance. This may be accomplished by:

- o Repeat testing of known patient samples over time;
- o Testing QC material in duplicate and over time; or
- o Repeat testing of calibration materials over time.

EXCEPTION: For fully automated systems that are not user dependent, operator variance does not need to be evaluated.

Interpretative Guidelines §493.1253(b)(1)(i)(C)

Reportable Range- The laboratory is responsible for verifying the reportable range of patient test results for each test system. Verification of reportable range may be accomplished by:

- o Assaying low and high calibration materials or control materials; or
- o Evaluating known samples of abnormal high and abnormal low values.

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Hematology whole blood high range calibration materials are not generally available. Therefore, laboratories may use patient specimens with verified elevated cell counts to verify the upper limit of the reportable range.

Probes §493.1253(b)(1)(i)(C)

If a dilution procedure is used when patient results exceed the test system's reportable range, how does the laboratory ensure the appropriate diluent is used for each type of specimen?

How does the laboratory verify and document the accuracy of the results for diluted specimens?

Interpretative Guidelines §493.1253(b)(1)(ii)

Reference Range (Normal Values) - The laboratory may use the manufacturer's reference range provided it is appropriate for the laboratory's patient population (i.e., a normal range that reflects the type of specimen and demographic variables such as age and sex, as applicable). If the manufacturer has not provided reference ranges appropriate for the laboratory's patient population, the laboratory may use published reference range(s). The laboratory must evaluate an appropriate number of specimens to verify the manufacturer's claims for normal values or, as applicable, the published reference ranges.

**FED - D5423 - ESTABLISHMENT AND VERIFICATION OF PERFORMANCE**

**Title** ESTABLISHMENT AND VERIFICATION OF PERFORMANCE

**CFR** 493.1253(b)(2)

**Type** Standard

**Regulation Definition**

Each laboratory that modifies an FDA-cleared or approved test system, or introduces a test system not subject to FDA clearance or approval (including methods developed in-house and standardized methods such as text book procedures), or uses a test system in which performance specifications are not provided by the manufacturer must, before reporting patient test results, establish for each test system the performance

**Interpretive Guideline**

Interpretative Guidelines §493.1253(b)(2)

Prior to reporting patient test results, the laboratory is responsible for establishing the performance specifications for each modified FDA-cleared or approved test system, each test system not subject to FDA clearance or approval, and each test system for which the manufacturer does not provide performance specifications. The establishment of method performance specifications should provide evidence that the accuracy, precision, analytical sensitivity, and analytical specificity of the procedure is adequate to meet the clients' needs as determined by the laboratory director

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specifications for the following performance characteristics, as applicable:

- (2)(i) Accuracy.
- (2)(ii) Precision.
- (2)(iii) Analytical sensitivity.
- (2)(iv) Analytical specificity to include interfering substances.
- (2)(v) Reportable range of test results for the test system.
- (2)(vi) Reference intervals (normal values).
- (2)(vii) Any other performance characteristic required for test performance.

and clinical consultant.

"Modified by the laboratory" means any change to the assay that could affect its performance specifications for sensitivity, specificity, accuracy, or precision, etc. Laboratory modification of the manufacturer's instructions that could affect performance specifications include but are not limited to:

- o Change in specimen handling instructions;
- o Change in incubation times or temperatures;
- o Change in dilution of specimen or reagent;
- o Using a different calibration material or reference material, or changing the manufacturer's set-points;
- o Introducing a different antibody (source, monoclonal-vs.-polyclonal);
- o Change or elimination of a procedural step;
- o Change or addition of detector (conjugate) or substrate;
- o Change in the solid phase;
- o Change in the cutoff or method of calculating the cutoff for semi-quantitative assays;
- o Change in the endpoint or calculation of the endpoint;
- o Addition of adsorbent; and
- o Change in the strain of antigen in serologic assays.

A modified moderate complexity test (including modifications in its intended use) is considered uncategorized for CLIA and therefore becomes a high complexity test.

EXCEPTIONS: Use of a manufacturer's reagents that are exempt from the premarket notification procedures in 21 CFR §807 for an instrument produced by another manufacturer is not considered a method modification. If the FDA

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has cleared a manufacturer's reagents and/or calibration materials for use with an instrument produced by another manufacturer, the use of these reagents/materials is not considered a method modification and does not require establishment of performance specifications. However, the laboratory must verify performance specifications as required under §493.1253(b)(1). Reverification of performance specifications is required if reagents are changed to those of another manufacturer.

"Modified by the laboratory" also means any change in intended use that could affect test system performance specifications for sensitivity, specificity, accuracy, and precision, etc., and the clinical utility of the test system. Changes in intended use are considered "off-label" use of a commercial test system. CAUTION: "Off-label" use is not supported by the manufacturer's clinical data.

Examples of changes in intended use are:

- o Using a different sample matrix (plasma vs. urine);
- o Using or promoting the test for another purpose (screening vs. diagnostic); and
- o Changing the type of analysis (qualitative results reported as quantitative).

NOTE: The laboratory is responsible for establishing performance specifications for test systems using analyte specific reagents (ASR).

For automated or semi-automated analyzers, the use of reprocessed (reconditioned) rotors/cuvettes which have passed quality control inspection criteria of the reprocessing company, are not considered a method modification if/when they are returned to the same laboratory that sent them for cleaning and re-use.

Specimens of known quantitative value may be used to determine the laboratory's performance specifications for a qualitative test.

Each laboratory is responsible for determining that its performance specifications for each test method are not affected by the relocation of the laboratory or test system. (See manufacturer's package insert regarding critical requirements such as set-up, limitations, environmental conditions, etc.)

If calibration material is used to establish method performance specifications, the laboratory must demonstrate that there is a minimal matrix effect and the calibration material is appropriate for establishing test system performance

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specifications.

If the LIS performs any calculations to determine a laboratory result, the calculations must be verified immediately after the LIS is programmed and prior to initial calculation of patient results.

NOTE: Public health testing performed on environmental (non-human) samples is not subject to CLIA.

Probes §493.1253(b)(2)

How does the laboratory determine if a new or revised LIS program (whether purchased or developed in-house) performs acceptably before it is integrated into routine operation?

Interpretative Guidelines §493.1253(b)(2)(i)

Accuracy

The laboratory is responsible for establishing that the method produces correct results.

Establishment of accuracy may be accomplished by:

- o Testing reference materials or comparing results of tests performed using an established reference method; or
- o Comparing split sample results with results obtained from another method, which has already been shown to provide accurate results.

For qualitative methods, the laboratory is responsible for establishing that a method will identify the presence/absence of the analyte.

In establishing a test system for a new analyte, research results may be used to document the accuracy of the test by correlation with the clinical presentation. In addition, the laboratory needs to determine the test system's precision and have mechanisms for determining analytical specificity, analytical sensitivity, and interfering substances.

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§493.1253 Standard: Establishment and verification of performance specifications

Interpretative Guidelines §493.1253(b)(2)(ii)

Precision (Reproducibility) - The laboratory is responsible for establishing the precision of each test system by assessing day-to-day, run-to-run, and within-run variation, as well as operator variance.

This may be accomplished by:

- o Repeat testing of known patient samples over time;
- o Testing QC material in duplicate and over time; or
- o Repeat testing of calibration materials over time.

EXCEPTION: For fully automated systems that are not user dependent, operator variance does not need to be evaluated.

§493.1253 Standard: Establishment and verification of performance specifications

Interpretative Guidelines §493.1253(b)(2)(iii)

Analytical Sensitivity - The laboratory is responsible for determining the lowest concentration or amount of the analyte or substance that can be measured or distinguished from a blank, i.e., minimum detection limits or how much of the analyte must be present to be measured.

For modified test systems, the laboratory may use the lower limit of the manufacturer's reportable range if it has demonstrated that the modification has not affected the lower limit.

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Interpretative Guidelines §493.1253(b)(2)(iv)

Analytical Specificity - The laboratory must determine the extent to which the method measures the analyte for which it is reporting results.

Interfering Substances - The laboratory must document information regarding interfering substances from product information, literature, or its own testing. These may include: specimen hemolysis, anticoagulant, lipemia, and turbidity; patients' clinical conditions, disease states, and medications.

§493.1253 Standard: Establishment and verification of performance specifications

Interpretative Guidelines §493.1253(b)(2)(v)

Reportable Range- The laboratory is responsible for establishing the upper and lower limits of the test system.

§493.1253 Standard: Establishment and verification of performance specifications

Interpretative Guidelines §493.1253(b)(2)(vi)

Reference Range (Normal Values) - The laboratory must establish a reference range that is appropriate for the laboratory's patient population (i.e., a normal range that reflects the type of specimen and demographic variables such as age and sex, as applicable).

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(b)(2)(vii) Any other performance characteristic required for test performance.

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- o Frequency of quality control failures; and
- o Training, experience, and competency of technical personnel.

For additional criteria in determining calibration and quality control frequency refer to §§493.1255 and 493.1256.

**FED - D5425 - ESTABLISHMENT AND VERIFICATION OF PERFORMANCE**

**Title** ESTABLISHMENT AND VERIFICATION OF PERFORMANCE

**CFR** 493.1253(b)(3)

**Type** Standard

**Regulation Definition**

The laboratory must determine the test system's calibration procedures and control procedures based upon the performance specifications verified or established under paragraph (b)(1) or (b)(2) of this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1253(b)(3)

Through the verification/establishment process, the laboratory defines the frequency for calibration and control performance as well as the type, number, and concentration of calibration and control materials used to monitor, detect error, and evaluate method performance. The frequency for calibration and control performance must not be less than the frequency specified in the manufacturer's instructions.

In establishing the calibration and quality control frequency, the laboratory must consider:

- o Test system instrument/reagent stability, including relocation;
- o Frequency with which the test is performed;
- o Technique dependence of the method;
- o Frequency of quality control failures; and
- o Training, experience, and competency of technical personnel.

For additional criteria in determining calibration and quality control frequency refer to

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§§493.1255 and 493.1256.

**FED - D5427 - ESTABLISHMENT AND VERIFICATION OF PERFORMANCE**

**Title** ESTABLISHMENT AND VERIFICATION OF PERFORMANCE

**CFR** 493.1253(c)

**Type** Standard

**Regulation Definition**

(c) Documentation. The laboratory must document all activities specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1253(c)

The actual measurement(s) taken, reactions and/or observations must be recorded.

Acceptable formats for documentation may vary.

**FED - D5429 - MAINTENANCE AND FUNCTION CHECKS**

**Title** MAINTENANCE AND FUNCTION CHECKS

**CFR** 493.1254(a)(1)

**Type** Standard

**Regulation Definition**

For unmodified manufacturer's equipment, instruments, or test systems, the laboratory must perform and document maintenance as defined by the manufacturer and with at least the frequency specified by the manufacturer.

**Interpretive Guideline**

Interpretative Guideline §493.1254(a)

When a laboratory introduces a new test system, the laboratory may determine, depending on the outcome of the performance specifications, that additional measures are necessary in order to ensure accurate and reliable test results.

Interpretative Guidelines §493.1254(a)(1)

"As defined by the manufacturer" means that the laboratory complies with the maintenance recommended or required in package inserts and/or instrument operator manuals for each piece of equipment/instrument it uses, including those that are peripherally involved in patient testing (e.g., incubators, centrifuges, safety cabinets, autoclaves and microscopes).

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A laboratory's maintenance program is usually divided into two parts:

- . Unscheduled repairs when needed; and
- . Scheduled preventive maintenance (PM), which is performed to prevent breakdowns or malfunctions, to prolong the life of an instrument and to maintain optimum operating characteristics.

A service contract for PM from an outside source is acceptable provided that for each instrument or piece of equipment, there is a description of the service to be performed and frequency of service.

A service contract does not negate the laboratory's responsibility for performing other routine maintenance not included in the maintenance contract. Acceptable performance parameters (if applicable) must be documented.

The laboratory must perform and document maintenance as specified by the manufacturer for the LIS computer and devices such as monitors, printers and modems. All devices must be maintained to ensure accurate, clear, and interference-free transmission.

Probes §493.1254(a)(1)

Are LIS system components (e.g., server, hard drives, disk packs) maintained according to the manufacturer's instructions?

When downtime is required to perform maintenance on LIS equipment, how are LIS users notified?

How does the laboratory's maintenance program ensure that instruments and equipment maintain optimum operating characteristics and minimize breakdowns?

**FED - D5431 - MAINTENANCE AND FUNCTION CHECKS**

**Title** MAINTENANCE AND FUNCTION CHECKS

**CFR** 493.1254(a)(2)

**Type** Standard

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**Regulation Definition**

For unmodified manufacturer's equipment, instruments, or test systems, the laboratory must perform and document function checks as defined by the manufacturer and with at least the frequency specified by the manufacturer. Function checks must be within the manufacturer's established limits before patient testing is conducted.

**Interpretive Guideline**

Interpretative Guidelines §493.1254(a)(2)

Function checks refer to those activities performed to evaluate critical operating characteristics (e.g., stray light, zeroing, electrical levels, optical alignment, background counts, counting efficiency) according to the accepted method of operation for each type of device or instrument. Daily quality control activities and function checks are performed prior to patient testing to ensure that an instrument is functioning correctly and is properly calibrated (Checking electrical, mechanical, and operational functions may be independent of the procedure). The performance of daily quality control activities may serve as an additional instrument function check, since analysis of external control samples check the operating characteristics of a test system, including instrument stability and calibration.

The laboratory must follow and document the required functions checks as stated by the laboratory information system (LIS) manufacturer for the LIS computer and devices such as monitors, printers and modems.

For instruments that automatically perform function checks and flag problems, the laboratory is required to document the corrective actions in response to the flagged problems. Use D5793 for deficiencies related to documenting corrective actions in response to the flagged problems.

Flow Cytometry:

A fluorescence standard(s) for each fluorochrome should be used each day of patient testing to ensure:

- o Proper alignment of the optical system;
- o Standardization of the fluorescence detectors;
- o Resolution of dimly-stained particles; and
- o Appropriate compensation for spectral overlap of the fluorochromes.

Fluorescence standards should have the same fluorochromes as are used for the test, and with the exception of alignment standards, should have similar fluorescence intensities as found in the test specimens. The laboratory must have an acceptable range of performance for all procedures.

Probes §493.1254(a)(2)

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For those methods in which the centrifugation is a critical portion of the test, does the laboratory check the RPM's and timing periodically (e.g., urine sediments)?

Do the records of a laboratory that moves from testing site to testing site demonstrate the performance of function checks as necessary?

In immunofluorescent test procedures, how does the laboratory ensure that the bulb is emitting ultraviolet light at the correct wave length?

How does the laboratory ensure that the fluorescent light source has not exceeded the manufacturer's established optimal timeframe?

For procedures or test systems that require pipetting or dilution of patient specimens separately from controls or calibrators, how are autodiluters, microdiluters, and/or pipettors checked for adequate and consistent delivery?

For those systems that perform simultaneous fluid delivery to multi-well plates or tubes, how does the laboratory ensure uniform delivery of reagents or washing solutions to all wells or tubes?

**FED - D5433 - MAINTENANCE AND FUNCTION CHECKS**

**Title** MAINTENANCE AND FUNCTION CHECKS

**CFR** 493.1254(b)(1)

**Type** Standard

**Regulation Definition**

For equipment, instruments, or test systems developed in-house, commercially available and modified by the laboratory, or maintenance and function check protocols are not provided by the manufacturer, the laboratory must establish a maintenance protocol that ensures equipment, instrument, and test system performance that is necessary for accurate and reliable test results and test result reporting. The laboratory must perform and document the maintenance activities specified in paragraph (b)(1)(i) of this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1254(b)

The laboratory must establish and follow procedures for performing maintenance and function checks on each piece of equipment/instrument it uses, including those that are peripherally involved in patient testing (e.g., incubators, centrifuges, safety cabinets, autoclaves and microscopes).

A manufacturer's instructions may not require maintenance and function checks. However, if the laboratory determines that a maintenance and/or function check protocol is necessary in order to ensure accurate and reliable test results, the laboratory must establish a maintenance and/or function check protocol and perform and document the described activities as they are carried out over time.

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Interpretative Guidelines §493.1254(b)(1)

A laboratory's maintenance program is usually divided into two parts:

- o Unscheduled repairs when needed; and
- o Scheduled preventive maintenance (PM) which is performed to prevent breakdowns or malfunctions, to prolong the life of an instrument and to maintain optimum operating characteristics.

Probes §493.1254(b)(1)

How does the laboratory's maintenance program ensure that instruments and equipment maintain optimum operating characteristics and minimize breakdowns?

Has the laboratory evaluated whether any modifications it has made to a manufacturer's instrument or piece of equipment has resulted in the need for additional maintenance or function checks, and, if so, have the additional procedures been established and implemented?

**FED - D5435 - MAINTENANCE AND FUNCTION CHECKS**

**Title** MAINTENANCE AND FUNCTION CHECKS

**CFR** 493.1254(b)(2)

**Type** Standard

**Regulation Definition**

For equipment, instruments, or test systems developed in-house, commercially available and modified by the laboratory, or maintenance and function check protocols are not provided by the manufacturer, the laboratory must:

- (i) Define a function check protocol that ensures equipment, instrument, and test system performance that is necessary for accurate and reliable test results and test result reporting.
- (ii) Perform and document the function checks, including background or baseline checks, specified in paragraph (b)(2)

**Interpretive Guideline**

Interpretative Guidelines §493.1254(b)(2)(i)-(b)(2)(ii)

The laboratory must establish and follow procedures for performing function checks on each piece of equipment/instrument it uses, including those that are peripherally involved in patient testing (e.g., incubators, centrifuges, safety cabinets, autoclaves).

Function checks refer to those activities performed to evaluate critical operating characteristics (e.g., stray light, zeroing, electrical levels, optical alignment, background counts, counting efficiency) according to the accepted method of operation for each type of device or instrument. Daily quality control activities and function checks are

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(i) of this section. Function checks must be within the laboratory's established limits before patient testing is conducted.

performed prior to patient testing to ensure that an instrument is functioning correctly and is properly calibrated. Checking electrical, mechanical, and operational functions may be independent of the procedure. The performance of daily quality control activities serves as an additional instrument function check. Analysis of external control samples check the operating characteristics of a test system, including instrument stability and calibration.

When function checks are critical to test performance, the laboratory must have a mechanism in place to monitor such items as:

- o Rotator speed and circumference;
- o Timers;
- o Anaerobic chambers;
- o Cell washers;
- o Radioactive particle counters;
- o Blood cell counters; and
- o Nucleic acid amplification equipment.

Flow Cytometry:

A fluorescence standard(s) for each fluorochrome must be used each day of patient testing to ensure:

- o Proper alignment of the optical system;
- o Standardization of the fluorescence detectors;
- o Resolution of dimly-stained particles; and
- o Appropriate compensation for spectral overlap of the fluorochromes.

Fluorescence standards must have the same fluorochromes incorporated into them as are used for the test, and with

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the exception of alignment standards, must have similar fluorescence intensities as found in the test specimens. The laboratory must have an acceptable range of performance for all procedures.

For flow cytometers with air-cooled lasers, the laser should be tested each day patients are tested by peaking the laser signal and monitoring the current input (amps) to laser light output (milliwatts) to determine whether the brewster windows are in need of cleaning.

Probes §493.1254(b)(2)

For those methods in which the centrifugation is a critical portion of the test, how has the laboratory checked the established RPM's and timing as necessary?

In immunofluorescent test procedures, how does the laboratory ensure that the bulb is emitting ultraviolet light at the correct wavelength?

If function checks are not required or recommended by the manufacturer, how does the laboratory establish the performance criteria of its equipment and instruments?

For RIA testing, are backgrounds or baselines measured for each setting? For example, if the laboratory uses more than one type of isotope, at what window setting are background counts performed and recorded?

When performing flow cytometry analysis using two or more fluorochromes simultaneously, how does the laboratory identify and adjust for "spill over" into the other fluorescence detectors?

**FED - D5437 - CALIBRATION AND CALIBRATION VERIFICATION**

**Title** CALIBRATION AND CALIBRATION VERIFICATION

**CFR** 493.1255(a)

**Type** Standard

**Regulation Definition**

Unless otherwise specified in this subpart, for each applicable test system the laboratory must perform and document calibration procedures--

(1) Following the manufacturer's test system instructions, using calibration materials provided or specified, and with at

**Interpretive Guideline**

Interpretative Guidelines §493.1255

For definitions of calibration and calibration verification, refer to §493.2.

For calibration and calibration verification of blood gas analysis, see §493.1267(a) through (d).

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least the frequency recommended by the manufacturer;  
(2) Using the criteria verified or established by the laboratory as specified in §493.1253(b)(3)--  
(2)(i) Using calibration materials appropriate for the test system and, if possible, traceable to a reference method or reference material of known value; and  
(2)(ii) Including the number, type, and concentration of calibration materials, as well as acceptable limits for and the frequency of calibration; and  
(3) Whenever calibration verification fails to meet the laboratory's acceptable limits for calibration verification.

In many instances, the performance of method calibration serves to satisfy the requirement for instrument calibration. Calibration procedures are not to be confused with instrument/equipment function checks at §493.1254.

Interpretative Guidelines §493.1255(a)

Laboratories must meet or exceed the manufacturer's frequency recommendations for calibration and they must follow the manufacturer's instructions on carrying out the calibration.

The calibration requirement does not apply to a variety of procedures, which include, but are not limited to:

- o Manual procedures not involving an instrument (e.g., microbiology cultures, Kirby-Bauer disk susceptibility tests, tilt-tube prothrombin time test systems, ABO group and D (Rho) typing);
- o Microscopic procedures (e.g., KOH preparations, pinworm preparations, urine sediment analysis, all manual differential procedures, manual cytology screening procedures); and
- o Test systems which include instruments that cannot be adjusted or calibrated because they are factory or manufacturer calibrated (e.g. unit use devices). This would include prothrombin time procedures on a fibrometer, or instruments that utilize a whole blood specimen and single unit use cartridge (PT/INR, Activated Clotting Time).

The term "calibration material" has generally replaced "standard" since many instruments now use serum-based reference materials. "Calibration material" means a solution that has a known amount of analyte weighed in or has a value determined by repetitive testing using a reference/definitive test method or is traceable to a National Institute for Standards and Technology (NIST) Standard, if possible.

Test method calibration procedures must follow the manufacturer's recommendations and requirements. However, if a calibration system proves less stable than expected by the manufacturer, additional calibration materials and/or more frequent calibration may be required, as established or verified by the laboratory under §493.1253(b)(3).

The actual measurement(s) taken, reactions and/or observations must be recorded.

Probes §493.1255(a)

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If the laboratory calculates values for one or more calibration materials, are the calculations correct, and do the records reflect that the measured values are within the laboratory's established limits for the calibration materials?

**FED - D5439 - CALIBRATION AND CALIBRATION VERIFICATION**

**Title** CALIBRATION AND CALIBRATION VERIFICATION

**CFR** 493.1255(b)

**Type** Standard

**Regulation Definition**

Unless otherwise specified in this subpart, for each applicable test system the laboratory must do the following:

Perform and document calibration verification procedure -

(b)(1) Following the manufacturer's calibration verification instructions;

(b)(2) Using the criteria verified or established by the laboratory under §493.1253(b)(3)--

(b)(2)(i) Including the number, type, and concentration of the materials, as well as acceptable limits for calibration verification; and

(b)(2)(ii) Including at least a minimal (or zero) value, a mid-point value, and a maximum value near the upper limit of the range to verify the laboratory's reportable range of test results for the test system; and

(b)(3) At least once every 6 months and whenever any of the following occur:

(b)(3)(i) A complete change of reagents for a procedure is introduced, unless the laboratory can demonstrate that changing reagent lot numbers does not affect the range used to report patient test results, and control values are not adversely affected by reagent lot number changes.

(b)(3)(ii) There is major preventive maintenance or replacement of critical parts that may influence test performance.

(b)(3)(iii) Control materials reflect an unusual trend or shift,

**Interpretive Guideline**

Interpretative Guidelines §493.1255(b)

The calibration verification requirements may be met by verifying the procedure using a high level material such as a control, calibration material, or patient specimen and diluting it to cover the reportable range if allowed by the manufacturer.

Control activities routinely used to satisfy the requirement for §493.1256 do not satisfy the calibration verification requirements.

**EXCEPTIONS:**

1. Laboratories must perform and document calibration procedures following the manufacturer's test system instructions, using calibration materials provided or specified, and at a frequency that meets or exceeds that recommended by the manufacturer. Where the manufacturer does not provide such instruction, the laboratory may calibrate using 3 or more levels of calibration materials that include a low, mid, and high value at least every 6 months.
2. For automated cell counters, the calibration verification requirements are considered met if the laboratory follows the manufacturer's instructions for instrument operation and tests 2 levels of control materials each day of testing provided the control results meet the laboratory's criteria for acceptability. This exception does not apply to centrifugal hematology test systems.
3. For automated chemistry analyzers, the calibration verification requirements are considered met if the laboratory follows the manufacturer's instructions for instrument operation and routinely tests three levels of control materials (lowest level available, mid-level, and highest level available) more than once each day of testing, the control material

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or are outside of the laboratory's acceptable limits, and other means of assessing and correcting unacceptable control values fail to identify and correct the problem.

(b)(3)(iv) The laboratory's established schedule for verifying the reportable range for patient test results requires more frequent calibration verification.

results meet the laboratory's criteria for acceptability and the control materials are traceable to National Institute of Standards and Technology (NIST) reference materials.

Calibration materials, proficiency testing samples with known results, or control materials with known values may be used to perform calibration verification. For these materials, the laboratory must define acceptable limits for the difference between the measured value obtained, versus the actual concentration of the materials.

NOTE: PT samples can only be used after the event cut-off date.

"Calibration material" means a solution that has a known amount of analyte weighed in, has a value determined by repetitive testing using a reference/definitive test method or is traceable to National Institute of Standards and Technology (NIST) reference material, if possible.

If a manufacturer provides reagents for a test where all of the reagents for a test are packaged together, calibration verification is not required for each additional reagent package with the same lot number that is received in the same shipment. For example, if the laboratory receives 12 packs of reagents and the laboratory has verified calibration for at least one of the 12 packs of reagents, then the laboratory does not have to verify calibration for the remaining 11 packs of reagents provided that all 12 packs of reagents have the same lot number and were received on the same shipment to the laboratory. However, this exception does not override the requirement to perform calibration verification as specified at 493.1255(b)(3).

4. Calibration verification is not required on:

- o Instruments that are factory or manufacturer calibrated and/or
- o Tests that are considered non-quantitative (e.g. Prothrombin time and Activated Clotting Time, which are measured in units of time)

When reviewing the laboratory's maintenance and function check records as required in §493.1254, determine whether the laboratory performed calibration verification when major maintenance occurred or critical parts were replaced.

The actual measurement(s) taken, reactions and/or observations must be recorded.

Probes §493.1255(b)

If a laboratory does not perform calibration verification after a complete change of reagents, what data does the

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laboratory have to document that changing reagent lot numbers does not affect the reportable range of patient test results, and does not adversely affect control results?

If reagents are obtained from a manufacturer and all of the reagents for a test are packaged together, the laboratory is not required to perform calibration verification for each package of reagents, provided the packages of reagents are received in the same shipment and contain the same lot number.

When reviewing the laboratory's maintenance and function check records as required in §493.1254, determine whether the laboratory performed calibration verification when major maintenance occurred or critical parts were replaced.

The actual measurement(s) taken, reactions and/or observations must be recorded.

Probes §493.1255(b)

If a laboratory does not perform calibration verification after a complete change of reagents, what data does the laboratory have to document that changing reagent lot numbers does not affect the reportable range of patient test results, and does not adversely affect control results?

**FED - D5441 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(a)(b)(c)(g)

**Type** Standard

**Regulation Definition**

- (a) For each test system, the laboratory is responsible for having control procedures that monitor the accuracy and precision of the complete analytic process.
- (b) The laboratory must establish the number, type, and frequency of testing control materials using, if applicable, the performance specifications verified or established by the laboratory as specified in §493.1253(b)(3).
- (c) The control procedures must--
  - (c)(1) Detect immediate errors that occur due to test system failure, adverse environmental conditions, and operator performance.

**Interpretive Guideline**

Interpretative Guidelines §493.1256(a)-(c)

For each test system, the laboratory is responsible for monitoring the accuracy and precision of each phase of the analytic testing process by using control procedures that will detect immediate errors and errors occurring over time. Errors may occur due to test system failure, change in environmental conditions, and operator performance.

**TEST SYSTEM**

Test system failures may result from reagent contamination or deterioration, reagent lot variation, reaction temperature fluctuations, inadequate sampling, improper or loss of calibration, electronic or mechanical failure, power supply variances, etc.

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(c)(2) Monitor over time the accuracy and precision of test performance that may be influenced by changes in test system performance and environmental conditions, and variance in operator performance.

(g) The laboratory must document all control procedures performed.

**ENVIRONMENT**

Environmental conditions that may affect test system performance include temperature, airflow, light intensity, humidity, altitude, etc.

**OPERATOR (TESTING PERSONNEL)**

Operator (testing personnel) performance that may affect testing includes improper specimen preparation and handling, incorrect test interpretation, failure to follow the manufacturer's test system instructions, etc. Operator training prior to testing is critical and competency assessment over time is necessary to ensure continued appropriate test performance. (See subpart M.)

Interpretative Guidelines §493.1256(c)

**CONTROL PROCEDURES**

In determining the control procedures, including the frequency of testing controls that detect immediate errors and monitor test performance over time, the laboratory needs to consider the following:

- o Control procedures specified by the test system's manufacturer;
- o Test system instrument reliability and reagent stability (e.g., relocation);
- o Frequency and volume of test performance;
- o Technique dependence of the method;
- o Frequency of quality control failures; and
- o Training, experience, and competency of person(s) performing the test.

Traditionally, laboratories have tested two levels of external control materials daily to monitor the accuracy and precision of the analytic test system components. External control materials having a similar matrix to that of patient

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specimens, are treated in the same manner as patient specimens, and go through all analytic phases of testing as applicable. External control materials may be provided as part of the test system, provided separately or prepared in-house. Testing external controls meets the requirement for monitoring test system components, environment, and operator performance. External control materials may be:

- o Commercially or in-house prepared controls;
- o Proficiency testing specimens for which results have been confirmed;
- o Reference or control strains of microorganisms;
- o Calibrators of different lot numbers and concentration than those used to calibrate the system; or
- o Previously tested patient specimens provided the laboratory determines the acceptable performance level for the patient specimens.

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

**FED - D5445 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(d)(1)(2)(g)

**Type** Standard

**Regulation Definition**

Unless CMS Approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub. 7), that provides equivalent quality testing, the laboratory must--

(d)(1) Perform control procedures as defined in this section unless otherwise specified in the additional specialty and subspecialty requirements at §§493.1261 through 493.1278.

(d)(2) For each test system, perform control procedures using the number and frequency specified by the manufacturer or

**Interpretive Guideline**

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established by the laboratory when they meet or exceed the requirements in paragraph (d)(3) of this section.

(g) The laboratory must document all control procedures performed.

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{f97\froman\fcharset128\fprq2{\\*\panose 02020600040205080304}MS PMincho;}  
{f98\froman\fcharset128\fprq2{\\*\panose 02020600040205080304}@MS PMincho;}  
{f99\fnil\fcharset0\fprq2{\\*\panose 02000500030200090000}MV Boli;}{f100\swiss\fcharset0\fprq2{\\*\panose  
020b0502040204020203}Microsoft New Tai Lue;}  
{f101\fnil\fcharset0\fprq2{\\*\panose 02000504070300020003}Nyala;}{f102\swiss\fcharset0\fprq2{\\*\panose  
020b0502040204020203}Microsoft PhagsPa;}{f103\froman\fcharset0\fprq2{\\*\panose  
02020602070100000000}Plantagenet Cherokee;}  
{f104\swiss\fcharset0\fprq2{\\*\panose 020b0504020000000003}Segoe Script;}  
{f105\swiss\fcharset0\fprq2{\\*\panose 020b0702040204020203}Segoe UI Semibold;}  
{f106\swiss\fcharset0\fprq2{\\*\panose 020b0502040204020203}Segoe UI Light;}  
{f107\swiss\fcharset0\fprq2{\\*\panose 020b0502040204020203}Segoe UI Symbol;}  
{f108\fnil\fcharset134\fprq2{\\*\panose 02010600030101010101}@SimSun;}  
{f109\modern\fcharset134\fprq1{\\*\panose 02010609030101010101}NSimSun;}  
{f110\modern\fcharset134\fprq1{\\*\panose 02010609030101010101}@NSimSun;}  
{f111\modern\fcharset134\fprq1{\\*\panose 02010609060101010101}SimSun-ExtB;}  
{f112\modern\fcharset134\fprq1{\\*\panose 02010609060101010101}@SimSun-ExtB;}  
{f113\swiss\fcharset0\fprq2{\\*\panose 020b0502040204020203}Microsoft Tai Le;}  
{f114\swiss\fcharset0\fprq2{\\*\panose 020b0502040204020203}Shonar Bangla;}  
{f115\fscrip\t\fcharset0\fprq2{\\*\panose 03000500000000000000}Microsoft Yi Baiti;}  
{f116\swiss\fcharset0\fprq2{\\*\panose 020b0604020202020204}Microsoft Sans Serif;}

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{\f117\fwiss\fcharset0\fprq2 {\\*\panose 020b06040202020204} Aparajita;} {\f118\fnil\fcharset0\fprq2 {\\*\panose 020000000000000000} Ebrima;};

{\f119\fwiss\fcharset0\fprq2 {\\*\panose 020b0502040204020203} Gisha;} {\f120\fwiss\fcharset0\fprq2 {\\*\panose 020b06040202020204} Kokila;} {\f121\fwiss\fcharset0\fprq2 {\\*\panose 020b0502040204020203} Leelawadee;};

{\f122\fnil\fcharset0\fprq2 {\\*\panose 020000000000000000} Microsoft Uighur;};

{\f123\fwiss\fcharset0\fprq2 {\\*\panose 020b0100010101010101} MoolBoran;};

{\f124\fwiss\fcharset0\fprq2 {\\*\panose 020b0604020202020204} Utsaah;};

{\f125\fwiss\fcharset0\fprq2 {\\*\panose 020b0604020202020204} Vijaya;} {\f126\froman\fcharset0\fprq2 {\\*\panose 02020603050405020304} Andalus;} {\f127\fscript\fcharset0\fprq2 {\\*\panose 03020402040406030203} Arabic Typesetting;};

{\f128\froman\fcharset0\fprq2 {\\*\panose 02020603050405020304} Simplified Arabic;};

{\f129\fmodern\fcharset0\fprq1 {\\*\panose 02070309020205020404} Simplified Arabic Fixed;};

{\f130\fnil\fcharset0\fprq2 {\\*\panose 020000000000000000} Sakkal Majalla;};

{\f131\froman\fcharset0\fprq2 {\\*\panose 02020603050405020304} Traditional Arabic;}; {\f132\fbidi \fnil\fcharset177\fprq2 {\\*\panose 02010803020104030203} Aharoni;}; {\f133\fbidi \fwiss\fcharset177\fprq2 {\\*\panose 020e0502060401010101} David;};

{\f134\fbidi \fwiss\fcharset177\fprq2 {\\*\panose 020e0503060101010101} FrankRuehl;}; {\f135\fbidi \fnil\fcharset177\fprq2 {\\*\panose 02010502060101010101} Levenim MT;}; {\f136\fbidi \fwiss\fcharset177\fprq2 {\\*\panose 020b0502050101010101} Miriam;};

{\f137\fbidi \fmodern\fcharset177\fprq1 {\\*\panose 020b0509050101010101} Miriam Fixed;}; {\f138\fbidi \fwiss\fcharset177\fprq2 {\\*\panose 020e0502050101010101} Narkisim;}; {\f139\fbidi \fmodern\fcharset177\fprq1 {\\*\panose 02030509050101010101} Rod;};

{\f140\fmodern\fcharset134\fprq1 {\\*\panose 02010609060101010101} FangSong;};

{\f141\fmodern\fcharset134\fprq1 {\\*\panose 02010609060101010101} @FangSong;};

{\f142\fmodern\fcharset134\fprq1 {\\*\panose 02010609060101010101} @SimHei;};

{\f143\fmodern\fcharset134\fprq1 {\\*\panose 02010609060101010101} KaiTi;};

{\f144\fmodern\fcharset134\fprq1 {\\*\panose 02010609060101010101} @KaiTi;};

{\f145\froman\fcharset0\fprq2 {\\*\panose 02020603050405020304} AngsanaUPC;};

{\f146\fwiss\fcharset0\fprq2 {\\*\panose 020b0604020202020204} Browallia New;};

{\f147\fwiss\fcharset0\fprq2 {\\*\panose 020b0604020202020204} BrowalliaUPC;};

{\f148\fwiss\fcharset0\fprq2 {\\*\panose 020b0304020202020204} CordiaUPC;};

{\f149\froman\fcharset0\fprq2 {\\*\panose 02020603050405020304} DilleniaUPC;};

{\f150\froman\fcharset0\fprq2 {\\*\panose 02020603050405020304} EucrosiaUPC;};

{\f151\fwiss\fcharset0\fprq2 {\\*\panose 020b0604020202020204} FreesiaUPC;};

{\f152\fwiss\fcharset0\fprq2 {\\*\panose 020b0604020202020204} IrisUPC;}; {\f153\froman\fcharset0\fprq2 {\\*\panose

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02020603050405020304} JasmineUPC; } {\f154\froman\fcharset0\fpqr2 {\\*\panose  
02020603050405020304} KodchiangUPC; }  
{\f155\fwiss\ffchar\fpqr2 {\\*\panose 020b06040202020204} LilyUPC; }  
{\f156\fscrip\ffchar\fpqr1 {\\*\panose 03000509000000000000} DFKai-SB; }  
{\f157\fscrip\ffchar\fpqr1 {\\*\panose 03000509000000000000} @DFKai-SB; }  
{\f158\fwiss\ffchar\fpqr2 {\\*\panose 020b0602030504020204} Lucida Sans Unicode; }  
{\f159\fwiss\ffchar\fpqr2 {\\*\panose 020b0a04020102020204} Arial Black; }  
{\f160\fwiss\ffchar\fpqr2 {\\*\panose 020e0502030303020204} Candara; }  
{\f161\fscrip\ffchar\fpqr2 {\\*\panose 030f0702030302020204} Comic Sans MS; }  
{\f162\fmodyn\ffchar\fpqr1 {\\*\panose 020b0609020204030204} Consolas; }  
{\f163\froman\ffchar\fpqr2 {\\*\panose 02030602050306030303} Constantia {\\*\falt Constantia}; }  
{\f164\fwiss\ffchar\fpqr2 {\\*\panose 020b0503020204020204} Corbel; } {\f165\fwiss\ffchar\fpqr2 {\\*\panose  
020b0603020102020204} Franklin Gothic Medium; } {\f166\fdcor\ffchar\fpqr2 {\\*\panose  
04040605051002020d02} Gabriola; }  
{\f167\froman\ffchar\fpqr2 {\\*\panose 02040502050405020303} Georgia; }  
{\f168\froman\ffchar\fpqr2 {\\*\panose 02040502050505030304} Palatino Linotype; }  
{\f169\fnil\ffchar\fpqr2 {\\*\panose 02000600000000000000} Segoe Print; }  
{\f170\fwiss\ffchar\fpqr2 {\\*\panose 020b0603020202020204} Trebuchet MS; }  
{\f171\fwiss\ffchar\fpqr2 {\\*\panose 020b0604030504040204} Verdana; } {\f172\froman\ffchar\fpqr2 {\\*\panose  
05030102010509060703} Webdings; }  
{\f173\froman\ffchar\fpqr2 {\\*\panose 02040602050505020304} Lucida Bright; }  
{\f174\fwiss\ffchar\fpqr2 {\\*\panose 020b0602030504020204} Lucida Sans; }  
{\f175\fdcor\ffchar\fpqr2 {\\*\panose 04020705040a02060702} Algerian; }  
{\f176\fwiss\ffchar\fpqr2 {\\*\panose 020b0604020202020204} @Arial Unicode MS; }  
{\f177\froman\ffchar\fpqr2 {\\*\panose 02020602080505020303} Baskerville Old Face; }  
{\f178\fdcor\ffchar\fpqr2 {\\*\panose 04030905020b02020c02} Bauhaus 93; }  
{\f179\froman\ffchar\fpqr2 {\\*\panose 02020503060305020303} Bell MT; }  
{\f180\fwiss\ffchar\fpqr2 {\\*\panose 020e0602020502020306} Berlin Sans FB; }  
{\f181\froman\ffchar\fpqr2 {\\*\panose 02050806060905020404} Bernard MT Condensed; }  
{\f182\froman\ffchar\fpqr2 {\\*\panose 02070706080601050204} Bodoni MT Poster Compressed; }  
{\f183\froman\ffchar\fpqr2 {\\*\panose 02040602050305030304} Book Antiqua; }  
{\f184\froman\ffchar\fpqr2 {\\*\panose 02050604050505020204} Bookman Old Style; }  
{\f185\fwiss\ffchar\fpqr2 {\\*\panose 020b0903060703020204} Britannic Bold; }  
{\f186\fdcor\ffchar\fpqr2 {\\*\panose 04040905080b02020502} Broadway; }  
{\f187\fscrip\ffchar\fpqr2 {\\*\panose 03060802040406070304} Brush Script MT; }

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{\f188\froman\fcharset0\fprq2{\\*\panose 0207040306080b030204} Californian FB;}  
{\f189\froman\fcharset0\fprq2{\\*\panose 02030504050205020304} Centaur;}{\f190\fwiss\fcharset0\fprq2{\\*\panose 020b05020202020204} Century Gothic;}  
{\f191\fdcor\fcharset0\fprq2{\\*\panose 04020404031007020602} Chiller;}{\f192\fdcor\fcharset0\fprq2{\\*\panose 04020805060202030203} Colonna MT;}{\f193\froman\fcharset0\fprq2{\\*\panose 0208090404030b020404} Cooper Black;}  
{\f194\froman\fcharset0\fprq2{\\*\panose 0204060206030a020304} Footlight MT Light;}  
{\f195\fscrip\fcharset0\fprq2{\\*\panose 030804020302050b0404} Freestyle Script;}  
{\f196\froman\fcharset0\fprq2{\\*\panose 02020404030301010803} Garamond;}  
{\f197\fdcor\fcharset0\fprq2{\\*\panose 04030604020f02020d02} Harlow Solid Italic;}  
{\f198\fdcor\fcharset0\fprq2{\\*\panose 04040505050a02020702} Harrington;}  
{\f199\froman\fcharset0\fprq2{\\*\panose 02040502050506030303} High Tower Text;}  
{\f200\fdcor\fcharset0\fprq2{\\*\panose 04090605060d06020702} Jokerman;}  
{\f201\fdcor\fcharset0\fprq2{\\*\panose 04040403040a02020202} Juice ITC;}  
{\f202\fscrip\fcharset0\fprq2{\\*\panose 03050502040202030202} Kristen ITC;}  
{\f203\fscrip\fcharset0\fprq2{\\*\panose 030304020206070d0d06} Kunstler Script;}  
{\f204\fscrip\fcharset0\fprq2{\\*\panose 03010101010101010101} Lucida Calligraphy;}  
{\f205\froman\fcharset0\fprq2{\\*\panose 02060602050505020204} Lucida Fax;}  
{\f206\fscrip\fcharset0\fprq2{\\*\panose 03010101010101010101} Lucida Handwriting;}  
{\f207\fdcor\fcharset0\fprq2{\\*\panose 04030805050802020d02} Magneto;}{\f208\fscrip\fcharset0\fprq2{\\*\panose 03020802060602070202} Matura MT Script Capitals;}  
{\f209\fscrip\fcharset0\fprq2{\\*\panose 03090702030407020403} Mistral;}{\f210\froman\fcharset0\fprq2{\\*\panose 02070704070505020303} Modern No. 20;}{\f211\fscrip\fcharset0\fprq2{\\*\panose 03010101010201010101} Monotype Corsiva;}  
{\f212\fdcor\fcharset0\fprq2{\\*\panose 04020502070703030202} Niagara Engraved;}  
{\f213\fdcor\fcharset0\fprq2{\\*\panose 04020502070702020202} Niagara Solid;}  
{\f214\fscrip\fcharset0\fprq2{\\*\panose 03040902040508030806} Old English Text MT;}  
{\f215\fdcor\fcharset0\fprq2{\\*\panose 04050602080702020203} Onyx;}{\f216\fscrip\fcharset0\fprq2{\\*\panose 03040602040708040804} Parchment;}{\f217\fdcor\fcharset0\fprq2{\\*\panose 040506030a0602020202} Playbill;}  
{\f218\froman\fcharset0\fprq2{\\*\panose 02080502050505020702} Poor Richard;}  
{\f219\fdcor\fcharset0\fprq2{\\*\panose 04040805050809020602} Ravie;}{\f220\fscrip\fcharset0\fprq2{\\*\panose 030604020304060b0204} Informal Roman;}  
{\f221\fdcor\fcharset0\fprq2{\\*\panose 04020904020102020604} Showcard Gothic;}  
{\f222\fdcor\fcharset0\fprq2{\\*\panose 04040a07060a02020202} Snap ITC;}  
{\f223\fdcor\fcharset0\fprq2{\\*\panose 040409050d0802020404} Stencil;}

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{f224\fdcor\fcharset0\fprq2 {\\*\panose 04020404030d07020202} Tempus Sans ITC;};  
{f225\fscrip\fcharset0\fprq2 {\\*\panose 03070502030502020203} Viner Hand ITC;};  
{f226\fscrip\fcharset0\fprq2 {\\*\panose 03020602050506090804} Vivaldi;};  
{f227\fscrip\fcharset0\fprq2 {\\*\panose 03050402040407070305} Vladimir Script;};  
{f228\froman\fcharset0\fprq2 {\\*\panose 020a0a07050505020404} Wide Latin;};  
{f229\froman\fcharset2\fprq2 {\\*\panose 05020102010507070707} Wingdings 2;};  
{f230\froman\fcharset2\fprq2 {\\*\panose 05040102010807070707} Wingdings 3;};  
{f231\fwiss\fcharset0\fprq2 {\\*\panose 020e0802020502020306} Berlin Sans FB Demi;};  
{f232\fwiss\fcharset0\fprq2 {\\*\panose 020b0604030504040204} MS Reference Sans Serif;};  
{f233\fnil\fcharset2\fprq2 {\\*\panose 05000500000000000000} MS Reference Specialty;};  
{f234\froman\fcharset2\fprq2 {\\*\panose 05050102010205020202} MT Extra;};  
{f235\fwiss\fcharset0\fprq2 {\\*\panose 020b0506020202030204} Arial Narrow;};  
{f236\fnil\fcharset2\fprq2 {\\*\panose 05010101010101010101} Bookshelf Symbol 7;};  
{f237\fwiss\fcharset0\fprq2 {\\*\panose 020f0302020204030204} Calibri Light;};  
{f238\fwiss\fcharset0\fprq2 {\\*\panose 020b0503020202020204} Agency FB;};  
{f239\fwiss\fcharset0\fprq2 {\\*\panose 020f0704030504030204} Arial Rounded MT Bold;};  
{f240\fdcor\fcharset0\fprq2 {\\*\panose 04020505051007020d02} Blackadder ITC;};  
{f241\froman\fcharset0\fprq2 {\\*\panose 02070603080606020203} Bodoni MT;};  
{f242\froman\fcharset0\fprq2 {\\*\panose 02070a03080606020203} Bodoni MT Black;};  
{f243\froman\fcharset0\fprq2 {\\*\panose 02070606080606020203} Bodoni MT Condensed;};  
{f244\fscrip\fcharset0\fprq2 {\\*\panose 03070402050302030203} Bradley Hand ITC;};  
{f245\froman\fcharset0\fprq2 {\\*\panose 02040603050505030304} Calisto MT;};  
{f246\froman\fcharset0\fprq2 {\\*\panose 020a0402060406010301} Castellar;};  
{f247\froman\fcharset0\fprq2 {\\*\panose 02040604050505020304} Century Schoolbook;};  
{f248\fwiss\fcharset0\fprq2 {\\*\panose 020e0705020206020404} Copperplate Gothic Bold;};  
{f249\fwiss\fcharset0\fprq2 {\\*\panose 020e0507020206020404} Copperplate Gothic Light;};  
{f250\fdcor\fcharset0\fprq2 {\\*\panose 04040404050702020202} Curlz MT;};  
{f251\fscrip\fcharset0\fprq2 {\\*\panose 030303020407070d0804} Edwardian Script ITC;};  
{f252\froman\fcharset0\fprq2 {\\*\panose 02020904090505020303} Elephant;};  
{f253\froman\fcharset0\fprq2 {\\*\panose 02090707080505020304} Engravers MT;};  
{f254\fwiss\fcharset0\fprq2 {\\*\panose 020b0907030504020204} Eras Bold ITC;};  
{f255\fwiss\fcharset0\fprq2 {\\*\panose 020b0805030504020804} Eras Demi ITC;};  
{f256\fwiss\fcharset0\fprq2 {\\*\panose 020b0402030504020804} Eras Light ITC;};  
{f257\fwiss\fcharset0\fprq2 {\\*\panose 020b0602030504020804} Eras Medium ITC;};  
{f258\fdcor\fcharset0\fprq2 {\\*\panose 04060505060202020a04} Felix Titling;};

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{\f259\fscrip\t\fccharset0\fpqr2 {\\*\panose 03060902040502070203} Forte;};  
{\f260\fwiss\fccharset0\fpqr2 {\\*\panose 020b0503020102020204} Franklin Gothic Book;};  
{\f261\fwiss\fccharset0\fpqr2 {\\*\panose 020b0703020102020204} Franklin Gothic Demi;};  
{\f262\fwiss\fccharset0\fpqr2 {\\*\panose 020b0706030402020204} Franklin Gothic Demi Cond;};  
{\f263\fwiss\fccharset0\fpqr2 {\\*\panose 020b0903020102020204} Franklin Gothic Heavy;};  
{\f264\fwiss\fccharset0\fpqr2 {\\*\panose 020b0606030402020204} Franklin Gothic Medium Cond;};  
{\f265\fscrip\t\fccharset0\fpqr2 {\\*\panose 03020402040607040605} French Script MT;};  
{\f266\fdcor\fccharset0\fpqr2 {\\*\panose 04040504061007020d02} Gigi;}; {\f267\fwiss\fccharset0\fpqr2 {\\*\panose 020b0902020104020203} Gill Sans MT Ext Condensed Bold;}; {\f268\fwiss\fccharset0\fpqr2 {\\*\panose 020b0502020104020203} Gill Sans MT;};  
{\f269\fwiss\fccharset0\fpqr2 {\\*\panose 020b0506020104020203} Gill Sans MT Condensed;};  
{\f270\fwiss\fccharset0\fpqr2 {\\*\panose 020b0a02020104020203} Gill Sans Ultra Bold;};  
{\f271\fwiss\fccharset0\fpqr2 {\\*\panose 020b0a06020104020203} Gill Sans Ultra Bold Condensed;};  
{\f272\froman\fccharset0\fpqr2 {\\*\panose 02030808020601010101} Gloucester MT Extra Condensed;};  
{\f273\froman\fccharset0\fpqr2 {\\*\panose 02020502050305020303} Goudy Old Style;};  
{\f274\froman\fccharset0\fpqr2 {\\*\panose 0202090407030b020401} Goudy Stout;};  
{\f275\fwiss\fccharset0\fpqr2 {\\*\panose 020b0706040902060204} Haettenschweiler;};  
{\f276\fdcor\fccharset0\fpqr2 {\\*\panose 04020605060303030202} Imprint MT Shadow;};  
{\f277\modern\fccharset0\fpqr1 {\\*\panose 020b0509030504030204} Lucida Sans Typewriter;};  
{\f278\fnil\fccharset2\fpqr2 {\\*\panose 05010100010000000000} MS Outlook;};  
{\f279\fwiss\fccharset0\fpqr2 {\\*\panose 020e0502030308020204} Maiandra GD;};  
{\f280\modern\fccharset0\fpqr2 {\\*\panose 02010509020102010303} OCR A Extended;};  
{\f281\fscrip\t\fccharset0\fpqr2 {\\*\panose 030303020206070c0b05} Palace Script MT;};  
{\f282\fscrip\t\fccharset0\fpqr2 {\\*\panose 03070502060502030205} Papyrus;}; {\f283\froman\fccharset0\fpqr2 {\\*\panose 02020502060401020303} Perpetua;}; {\f284\froman\fccharset0\fpqr2 {\\*\panose 02020502060505020804} Perpetua Titling MT;};  
{\f285\fscrip\t\fccharset0\fpqr2 {\\*\panose 03060402040406080204} Pristina;}; {\f286\fscrip\t\fccharset0\fpqr2 {\\*\panose 03070502040507070304} Rage Italic;}; {\f287\froman\fccharset0\fpqr2 {\\*\panose 02060603020205020403} Rockwell;};  
{\f288\froman\fccharset0\fpqr2 {\\*\panose 02060603050405020104} Rockwell Condensed;};  
{\f289\froman\fccharset0\fpqr2 {\\*\panose 02060903040505020403} Rockwell Extra Bold;};  
{\f290\fscrip\t\fccharset0\fpqr2 {\\*\panose 03040602040607080904} Script MT Bold;};  
{\f291\fwiss\fccharset0\fpqr2 {\\*\panose 020b0602020104020603} Tw Cen MT;};  
{\f292\fwiss\fccharset0\fpqr2 {\\*\panose 020b0606020104020203} Tw Cen MT Condensed;};  
{\f293\fwiss\fccharset0\fpqr2 {\\*\panose 020b0803020202020204} Tw Cen MT Condensed Extra Bold;};

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{f294\fwiss\charset0\frq2 {\\*\panose 020b0606030504040204}Nina;}{f295\fwiss\charset0\frq2 {\\*\panose 020b0606040200020203}Segoe Condensed;}{f296\fwiss\charset0\frq2 {\\*\panose 020f0502020204030204}Z@R54CC.tmp;}{f297\froman\charset0\frq0 {\\*\panose 000000000000000000}inherit;}{f298\froman\charset0\frq2 {\\*\panose 020205000000000000}Reveal Line Draw;}{f299\modern\charset0\frq1 {\\*\panose 02070309020205020404}Quest Font;}{f300\finl\charset0\frq2 {\\*\panose 000000000000000000}ZWAdobeF;}{f301\modern\charset0\frq2 {\\*\panose 000000000000000000}MMa Century;}{f302\modern\charset0\frq2 {\\*\panose 000000000000000000}MMa Fermat;}{f303\modern\charset0\frq2 {\\*\panose 000000000000000000}MMa Fraktur;}{f304\modern\charset0\frq2 {\\*\panose 000000000000000000}MMa Gauss;}{f305\modern\charset0\frq2 {\\*\panose 000000000000000000}MMa Pascal;}{f306\modern\charset0\frq2 {\\*\panose 000000000000000000}MMArrow;}{f307\modern\charset0\frq2 {\\*\panose 000000000000000000}MMa TextBook;}{f308\modern\charset0\frq2 {\\*\panose 000000000000000000}MMBinary;}{f309\modern\charset0\frq2 {\\*\panose 000000000000000000}MMCenturyNew;}{f310\modern\charset0\frq2 {\\*\panose 000000000000000000}MMCenturyOld;}{f311\modern\charset0\frq2 {\\*\panose 000000000000000000}MMCenturyOldGreek;}{f312\modern\charset0\frq2 {\\*\panose 000000000000000000}MMetc;}{f313\modern\charset0\frq2 {\\*\panose 000000000000000000}MMExtra;}{f314\modern\charset0\frq2 {\\*\panose 000000000000000000}MMGreek;}{f315\modern\charset0\frq2 {\\*\panose 000000000000000000}MMNegate;}{f316\modern\charset0\frq2 {\\*\panose 000000000000000000}MMRelation;}{f317\modern\charset0\frq2 {\\*\panose 000000000000000000}MMTextBook;}{f318\modern\charset0\frq2 {\\*\panose 000000000000000000}MMTimes;}{f319\modern\charset0\frq2 {\\*\panose 000000000000000000}MMVariable2;}{f320\modern\charset0\frq2 {\\*\panose 000000000000000000}MMVariableF;}{f321\modern\charset0\frq2 {\\*\panose 000000000000000000}MMSchoolRD;}{f322\decor\charset0\frq2 {\\*\panose 000000000000000000}ZapfDingbats;}{f323\fwiss\charset163\frq2 {\\*\panose 000000000000000000}Microsoft Sans Serif (Vietnames;){f324\fbidi \modern\charset178\frq1 {\\*\panose 000000000000000000}Simplified Arabic Fixed (Arabic;){f325\fwiss\charset163\frq2 {\\*\panose 000000000000000000}MS Reference Sans Serif (Vietna;){f326\fwiss\charset186\frq2 {\\*\panose 000000000000000000}Franklin Gothic Demi Cond Balti;}{f327\fwiss\charset161\frq2 {\\*\panose 000000000000000000}Franklin Gothic Medium Cond Gre;}{f328\fwiss\charset186\frq2 {\\*\panose 000000000000000000}Franklin Gothic Medium Cond Bal;}

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{f329\fwiss\fcharset238\fpqr2 {\\*\panose 000000000000000000} Gill Sans Ultra Bold Condensed ;}  
{f330\fwiss\fcharset238\fpqr2 {\\*\panose 000000000000000000} Tw Cen MT Condensed Extra Bold ;}  
{f331\fwiss\fcharset238\fpqr1 {\\*\panose 000000000000000000} Lucida Sans Typewriter CE ;}  
{f332\fwiss\fcharset204\fpqr1 {\\*\panose 000000000000000000} Lucida Sans Typewriter Cyr ;}  
{f333\fwiss\fcharset161\fpqr1 {\\*\panose 000000000000000000} Lucida Sans Typewriter Greek ;}  
{f334\fwiss\fcharset162\fpqr1 {\\*\panose 000000000000000000} Lucida Sans Typewriter Tur ;} {f335\fbidi  
\fwiss\fcharset177\fpqr1 {\\*\panose 000000000000000000} Lucida Sans Typewriter (Hebrew) ;}  
{f336\fbidi \fwiss\fcharset178\fpqr1 {\\*\panose 000000000000000000} Lucida Sans Typewriter (Arabic) ;}  
{f337\fwiss\fcharset186\fpqr1 {\\*\panose 000000000000000000} Lucida Sans Typewriter Baltic ;}  
{f338\fwiss\fcharset222\fpqr1 {\\*\panose 000000000000000000} Lucida Sans Typewriter (Thai) ;}  
{f339\fmmodern\fcharset238\fpqr1 {\\*\panose 000000000000000000} Quest Font CE ;}  
{f340\fmmodern\fcharset204\fpqr1 {\\*\panose 000000000000000000} Quest Font Cyr ;}  
{f341\fmmodern\fcharset161\fpqr1 {\\*\panose 000000000000000000} Quest Font Greek ;}  
{f342\fmmodern\fcharset162\fpqr1 {\\*\panose 000000000000000000} Quest Font Tur ;} {f343\fbidi  
\fmmodern\fcharset177\fpqr1 {\\*\panose 000000000000000000} Quest Font (Hebrew) ;}  
{f344\fbidi \fmmodern\fcharset178\fpqr1 {\\*\panose 000000000000000000} Quest Font (Arabic) ;}  
{f345\fmmodern\fcharset186\fpqr1 {\\*\panose 000000000000000000} Quest Font Baltic ;}  
{f346\fmmodern\fcharset163\fpqr1 {\\*\panose 000000000000000000} Quest Font (Vietnamese) ;}  
{f347\fnil\fcharset238\fpqr2 {\\*\panose 000000000000000000} ZWAdobeF CE ;}  
{f348\fnil\fcharset204\fpqr2 {\\*\panose 000000000000000000} ZWAdobeF Cyr ;}  
{f349\fnil\fcharset161\fpqr2 {\\*\panose 000000000000000000} ZWAdobeF Greek ;}  
{f350\fnil\fcharset162\fpqr2 {\\*\panose 000000000000000000} ZWAdobeF Tur ;} {f351\fbidi  
\fnil\fcharset177\fpqr2 {\\*\panose 000000000000000000} ZWAdobeF (Hebrew) ;}  
{f352\fbidi \fnil\fcharset178\fpqr2 {\\*\panose 000000000000000000} ZWAdobeF (Arabic) ;}  
{f353\fnil\fcharset186\fpqr2 {\\*\panose 000000000000000000} ZWAdobeF Baltic ;}  
{f354\fnil\fcharset163\fpqr2 {\\*\panose 000000000000000000} ZWAdobeF (Vietnamese) ;}  
{f355\fwiss\fcharset238\fpqr2 {\\*\panose 000000000000000000} Z@R54CC.tmp CE ;}  
{f356\fwiss\fcharset204\fpqr2 {\\*\panose 000000000000000000} Z@R54CC.tmp Cyr ;}  
{f357\fwiss\fcharset161\fpqr2 {\\*\panose 000000000000000000} Z@R54CC.tmp Greek ;}  
{f358\fwiss\fcharset162\fpqr2 {\\*\panose 000000000000000000} Z@R54CC.tmp Tur ;}  
{f359\fwiss\fcharset186\fpqr2 {\\*\panose 000000000000000000} Z@R54CC.tmp Baltic ;}  
{f360\fwiss\fcharset163\fpqr2 {\\*\panose 000000000000000000} Z@R54CC.tmp ( Vietnamese) ;}  
{f361\froman\fcharset238\fpqr2 Times New Roman CE {\\*\falt Times New Roman} ;}  
{f362\froman\fcharset204\fpqr2 Times New Roman Cyr {\\*\falt Times New Roman} ;}  
{f364\froman\fcharset161\fpqr2 Times New Roman Greek {\\*\falt Times New Roman} ;}



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{\f514\fmmodern\fcharset161\fprq1 MS Gothic Greek {\\*\falt ?!r ?S?V?b?N};} {\f515\fmmodern\fcharset162\fprq1 MS Gothic Tur {\\*\falt ?!r ?S?V?b?N};} {\f518\fmmodern\fcharset186\fprq1 MS Gothic Baltic {\\*\falt ?!r ?S?V?b?N};}

{\f523\fwissw\fcharset0\fprq2 Dotum Western {\\*\falt  
 \a2\aeE\ a1\cbci\ a2\aeE\ a1\cbce\ a2\aeE\ a1\cbce\ a1\ a7I\ a2\ae\ a8\cf\ a2\aeE\ a1\cbce?o};}

{\f521\fwissw\fcharset238\fprq2 Dotum CE {\\*\falt  
 \a2\aeE\ a1\cbci\ a2\aeE\ a1\cbce\ a2\aeE\ a1\cbce\ a1\ a7I\ a2\ae\ a8\cf\ a2\aeE\ a1\cbce?o};}

{\f522\fwissw\fcharset204\fprq2 Dotum Cyr {\\*\falt  
 \a2\aeE\ a1\cbci\ a2\aeE\ a1\cbce\ a2\aeE\ a1\cbce\ a1\ a7I\ a2\ae\ a8\cf\ a2\aeE\ a1\cbce?o};}

{\f524\fwissw\fcharset161\fprq2 Dotum Greek {\\*\falt  
 \a2\aeE\ a1\cbci\ a2\aeE\ a1\cbce\ a2\aeE\ a1\cbce\ a1\ a7I\ a2\ae\ a8\cf\ a2\aeE\ a1\cbce?o};}

{\f525\fwissw\fcharset162\fprq2 Dotum Tur {\\*\falt  
 \a2\aeE\ a1\cbci\ a2\aeE\ a1\cbce\ a2\aeE\ a1\cbce\ a1\ a7I\ a2\ae\ a8\cf\ a2\aeE\ a1\cbce?o};}

{\f528\fwissw\fcharset186\fprq2 Dotum Baltic {\\*\falt  
 \a2\aeE\ a1\cbci\ a2\aeE\ a1\cbce\ a2\aeE\ a1\cbce\ a1\ a7I\ a2\ae\ a8\cf\ a2\aeE\ a1\cbce?o};}

{\f533\fmmodern\fcharset0\fprq1 SimHei Western {\\*\falt o??\ a1\ a7???\ a1\ a7???\};}

{\f543\fmmodern\fcharset0\fprq1 MingLiU Western {\\*\falt 2OcuAe};}

{\f563\fwissw\fcharset0\fprq2 Gulim Western {\\*\falt  
 \a1\cb\ a2\ e7E\ a2\aeEc\ a1\cb\ a2\ e7\ a2\ae\ a1\bfu\ a2\aeE\ a1\cbc\ a1\cb\ a2\ e7\ a2\ae\ a2\afu\ a2\aeE\ a1\cbce\ a  
 2\aeE\ a1\cbce\ a1\ a7I};}

{\f561\fwissw\fcharset238\fprq2 Gulim CE {\\*\falt  
 \a1\cb\ a2\ e7E\ a2\aeEc\ a1\cb\ a2\ e7\ a2\ae\ a1\bfu\ a2\aeE\ a1\cbc\ a1\cb\ a2\ e7\ a2\ae\ a2\afu\ a2\aeE\ a1\cbce\ a  
 2\aeE\ a1\cbce\ a1\ a7I};}

{\f562\fwissw\fcharset204\fprq2 Gulim Cyr {\\*\falt  
 \a1\cb\ a2\ e7E\ a2\aeEc\ a1\cb\ a2\ e7\ a2\ae\ a1\bfu\ a2\aeE\ a1\cbc\ a1\cb\ a2\ e7\ a2\ae\ a2\afu\ a2\aeE\ a1\cbce\ a  
 2\aeE\ a1\cbce\ a1\ a7I};}

{\f564\fwissw\fcharset161\fprq2 Gulim Greek {\\*\falt  
 \a1\cb\ a2\ e7E\ a2\aeEc\ a1\cb\ a2\ e7\ a2\ae\ a1\bfu\ a2\aeE\ a1\cbc\ a1\cb\ a2\ e7\ a2\ae\ a2\afu\ a2\aeE\ a1\cbce\ a  
 2\aeE\ a1\cbce\ a1\ a7I};}

{\f565\fwissw\fcharset162\fprq2 Gulim Tur {\\*\falt  
 \a1\cb\ a2\ e7E\ a2\aeEc\ a1\cb\ a2\ e7\ a2\ae\ a1\bfu\ a2\aeE\ a1\cbc\ a1\cb\ a2\ e7\ a2\ae\ a2\afu\ a2\aeE\ a1\cbce\ a  
 2\aeE\ a1\cbce\ a1\ a7I};}

{\f568\fwissw\fcharset186\fprq2 Gulim Baltic {\\*\falt  
 \a1\cb\ a2\ e7E\ a2\aeEc\ a1\cb\ a2\ e7\ a2\ae\ a1\bfu\ a2\aeE\ a1\cbc\ a1\cb\ a2\ e7\ a2\ae\ a2\afu\ a2\aeE\ a1\cbce\ a  
 2\aeE\ a1\cbce\ a1\ a7I};}

{\f571\froman\fcharset238\fprq2 Century CE; } {\f572\froman\fcharset204\fprq2 Century Cyr; }

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{\f574\froman\fcharset161\fprq2 Century Greek;}{\f575\froman\fcharset162\fprq2 Century Tur;}  
{\f578\froman\fcharset186\fprq2 Century Baltic;}  
{\f590\froman\fcharset222\fprq2 Angsana New (Thai);}{\f600\fswiss\fcharset222\fprq2 Cordia New (Thai);}  
{\f621\froman\fcharset238\fprq2 Sylfaen CE;}{\f622\froman\fcharset204\fprq2 Sylfaen Cyr;}  
{\f624\froman\fcharset161\fprq2 Sylfaen Greek;}  
{\f625\froman\fcharset162\fprq2 Sylfaen Tur;}{\f628\froman\fcharset186\fprq2 Sylfaen Baltic;}  
{\f713\fswiss\fcharset0\fprq2 Arial Unicode MS Western;}{\f711\fswiss\fcharset238\fprq2 Arial Unicode MS CE;}  
{\f712\fswiss\fcharset204\fprq2 Arial Unicode MS Cyr;}{\f714\fswiss\fcharset161\fprq2 Arial Unicode MS Greek;}  
{\f715\fswiss\fcharset162\fprq2 Arial Unicode MS Tur;}{\f716\fbidi \fswiss\fcharset177\fprq2 Arial Unicode MS  
(Hebrew);}  
{\f717\fbidi \fswiss\fcharset178\fprq2 Arial Unicode MS (Arabic);}{\f718\fswiss\fcharset186\fprq2 Arial Unicode  
MS Baltic;}{\f719\fswiss\fcharset163\fprq2 Arial Unicode MS (Vietnamese);}{\f720\fswiss\fcharset222\fprq2 Arial  
Unicode MS (Thai);}  
{\f721\fswiss\fcharset238\fprq2 Tahoma CE;}{\f722\fswiss\fcharset204\fprq2 Tahoma Cyr;}  
{\f724\fswiss\fcharset161\fprq2 Tahoma Greek;}{\f725\fswiss\fcharset162\fprq2 Tahoma Tur;}{\f726\fbidi  
\fswiss\fcharset177\fprq2 Tahoma (Hebrew);}  
{\f727\fbidi \fswiss\fcharset178\fprq2 Tahoma (Arabic);}{\f728\fswiss\fcharset186\fprq2 Tahoma Baltic;}  
{\f729\fswiss\fcharset163\fprq2 Tahoma (Vietnamese);}{\f730\fswiss\fcharset222\fprq2 Tahoma (Thai);}  
{\f731\froman\fcharset238\fprq2 Cambria Math CE;}  
{\f732\froman\fcharset204\fprq2 Cambria Math Cyr;}{\f734\froman\fcharset161\fprq2 Cambria Math Greek;}  
{\f735\froman\fcharset162\fprq2 Cambria Math Tur;}{\f738\froman\fcharset186\fprq2 Cambria Math Baltic;}  
{\f739\froman\fcharset163\fprq2 Cambria Math (Vietnamese);}{\f741\fswiss\fcharset238\fprq2 Calibri CE;}  
{\f742\fswiss\fcharset204\fprq2 Calibri Cyr;}{\f744\fswiss\fcharset161\fprq2 Calibri Greek;}  
{\f745\fswiss\fcharset162\fprq2 Calibri Tur;}  
{\f748\fswiss\fcharset186\fprq2 Calibri Baltic;}{\f749\fswiss\fcharset163\fprq2 Calibri (Vietnamese);}  
{\f751\fswiss\fcharset238\fprq2 Segoe UI CE;}{\f752\fswiss\fcharset204\fprq2 Segoe UI Cyr;}  
{\f754\fswiss\fcharset161\fprq2 Segoe UI Greek;}  
{\f755\fswiss\fcharset162\fprq2 Segoe UI Tur;}{\f757\fbidi \fswiss\fcharset178\fprq2 Segoe UI (Arabic);}  
{\f758\fswiss\fcharset186\fprq2 Segoe UI Baltic;}{\f759\fswiss\fcharset163\fprq2 Segoe UI (Vietnamese);}  
{\f761\froman\fcharset238\fprq2 Cambria CE;}  
{\f762\froman\fcharset204\fprq2 Cambria Cyr;}{\f764\froman\fcharset161\fprq2 Cambria Greek;}  
{\f765\froman\fcharset162\fprq2 Cambria Tur;}{\f768\froman\fcharset186\fprq2 Cambria Baltic;}  
{\f769\froman\fcharset163\fprq2 Cambria (Vietnamese);}  
{\f771\froman\fcharset238\fprq2 Times CE;}{\f772\froman\fcharset204\fprq2 Times Cyr;}  
{\f774\froman\fcharset161\fprq2 Times Greek;}{\f775\froman\fcharset162\fprq2 Times Tur;}{\f776\fbidi

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\froman\fcharset177\fprq2 Times (Hebrew);}  
{\f777\fbidi \froman\fcharset178\fprq2 Times (Arabic);}{\f778\froman\fcharset186\fprq2 Times Baltic;}  
{\f779\froman\fcharset163\fprq2 Times (Vietnamese);}{\f793\froman\fcharset0\fprq2 MingLiU\_HKSCS Western;}  
{\f833\froman\fcharset0\fprq2 @Batang Western;}  
{\f831\froman\fcharset238\fprq2 @Batang CE;}{\f832\froman\fcharset204\fprq2 @Batang Cyr;}  
{\f834\froman\fcharset161\fprq2 @Batang Greek;}{\f835\froman\fcharset162\fprq2 @Batang Tur;}  
{\f838\froman\fcharset186\fprq2 @Batang Baltic;}  
{\f843\modern\fcharset0\fprq1 BatangChe Western;}{\f841\modern\fcharset238\fprq1 BatangChe CE;}  
{\f842\modern\fcharset204\fprq1 BatangChe Cyr;}{\f844\modern\fcharset161\fprq1 BatangChe Greek;}  
{\f845\modern\fcharset162\fprq1 BatangChe Tur;}  
{\f848\modern\fcharset186\fprq1 BatangChe Baltic;}{\f853\modern\fcharset0\fprq1 @BatangChe Western;}  
{\f851\modern\fcharset238\fprq1 @BatangChe CE;}{\f852\modern\fcharset204\fprq1 @BatangChe Cyr;}  
{\f854\modern\fcharset161\fprq1 @BatangChe Greek;}  
{\f855\modern\fcharset162\fprq1 @BatangChe Tur;}{\f858\modern\fcharset186\fprq1 @BatangChe Baltic;}  
{\f863\froman\fcharset0\fprq2 Gungsuh Western;}{\f861\froman\fcharset238\fprq2 Gungsuh CE;}  
{\f862\froman\fcharset204\fprq2 Gungsuh Cyr;}  
{\f864\froman\fcharset161\fprq2 Gungsuh Greek;}{\f865\froman\fcharset162\fprq2 Gungsuh Tur;}  
{\f868\froman\fcharset186\fprq2 Gungsuh Baltic;}{\f873\froman\fcharset0\fprq2 @Gungsuh Western;}  
{\f871\froman\fcharset238\fprq2 @Gungsuh CE;}  
{\f872\froman\fcharset204\fprq2 @Gungsuh Cyr;}{\f874\froman\fcharset161\fprq2 @Gungsuh Greek;}  
{\f875\froman\fcharset162\fprq2 @Gungsuh Tur;}{\f878\froman\fcharset186\fprq2 @Gungsuh Baltic;}  
{\f883\modern\fcharset0\fprq1 GungsuhChe Western;}  
{\f881\modern\fcharset238\fprq1 GungsuhChe CE;}{\f882\modern\fcharset204\fprq1 GungsuhChe Cyr;}  
{\f884\modern\fcharset161\fprq1 GungsuhChe Greek;}{\f885\modern\fcharset162\fprq1 GungsuhChe Tur;}  
{\f888\modern\fcharset186\fprq1 GungsuhChe Baltic;}  
{\f893\modern\fcharset0\fprq1 @GungsuhChe Western;}{\f891\modern\fcharset238\fprq1 @GungsuhChe CE;}  
{\f892\modern\fcharset204\fprq1 @GungsuhChe Cyr;}{\f894\modern\fcharset161\fprq1 @GungsuhChe Greek;}  
{\f895\modern\fcharset162\fprq1 @GungsuhChe Tur;}  
{\f898\modern\fcharset186\fprq1 @GungsuhChe Baltic;}{\f920\swiss\fcharset222\fprq2 DokChampa (Thai);}  
{\f943\swiss\fcharset0\fprq2 @Gulim Western;}{\f941\swiss\fcharset238\fprq2 @Gulim CE;}  
{\f942\swiss\fcharset204\fprq2 @Gulim Cyr;}  
{\f944\swiss\fcharset161\fprq2 @Gulim Greek;}{\f945\swiss\fcharset162\fprq2 @Gulim Tur;}  
{\f948\swiss\fcharset186\fprq2 @Gulim Baltic;}{\f953\modern\fcharset0\fprq1 GulimChe Western;}  
{\f951\modern\fcharset238\fprq1 GulimChe CE;}  
{\f952\modern\fcharset204\fprq1 GulimChe Cyr;}{\f954\modern\fcharset161\fprq1 GulimChe Greek;}

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{\f955\fmmodern\fcharset162\fprq1 GulimChe Tur;} {\f958\fmmodern\fcharset186\fprq1 GulimChe Baltic;}  
{\f963\fmmodern\fcharset0\fprq1 @GulimChe Western;}  
{\f961\fmmodern\fcharset238\fprq1 @GulimChe CE;} {\f962\fmmodern\fcharset204\fprq1 @GulimChe Cyr;}  
{\f964\fmmodern\fcharset161\fprq1 @GulimChe Greek;} {\f965\fmmodern\fcharset162\fprq1 @GulimChe Tur;}  
{\f968\fmmodern\fcharset186\fprq1 @GulimChe Baltic;}  
{\f973\fwiss\fcharset0\fprq2 @Dotum Western;} {\f971\fwiss\fcharset238\fprq2 @Dotum CE;}  
{\f972\fwiss\fcharset204\fprq2 @Dotum Cyr;} {\f974\fwiss\fcharset161\fprq2 @Dotum Greek;}  
{\f975\fwiss\fcharset162\fprq2 @Dotum Tur;}  
{\f978\fwiss\fcharset186\fprq2 @Dotum Baltic;} {\f983\fmmodern\fcharset0\fprq1 DotumChe Western;}  
{\f981\fmmodern\fcharset238\fprq1 DotumChe CE;} {\f982\fmmodern\fcharset204\fprq1 DotumChe Cyr;}  
{\f984\fmmodern\fcharset161\fprq1 DotumChe Greek;}  
{\f985\fmmodern\fcharset162\fprq1 DotumChe Tur;} {\f988\fmmodern\fcharset186\fprq1 DotumChe Baltic;}  
{\f993\fmmodern\fcharset0\fprq1 @DotumChe Western;} {\f991\fmmodern\fcharset238\fprq1 @DotumChe CE;}  
{\f992\fmmodern\fcharset204\fprq1 @DotumChe Cyr;}  
{\f994\fmmodern\fcharset161\fprq1 @DotumChe Greek;} {\f995\fmmodern\fcharset162\fprq1 @DotumChe Tur;}  
{\f998\fmmodern\fcharset186\fprq1 @DotumChe Baltic;} {\f1001\fwiss\fcharset238\fprq2 Impact CE;}  
{\f1002\fwiss\fcharset204\fprq2 Impact Cyr;}  
{\f1004\fwiss\fcharset161\fprq2 Impact Greek;} {\f1005\fwiss\fcharset162\fprq2 Impact Tur;}  
{\f1008\fwiss\fcharset186\fprq2 Impact Baltic;} {\f1051\fmmodern\fcharset238\fprq1 Lucida Console CE;}  
{\f1052\fmmodern\fcharset204\fprq1 Lucida Console Cyr;}  
{\f1054\fmmodern\fcharset161\fprq1 Lucida Console Greek;} {\f1055\fmmodern\fcharset162\fprq1 Lucida Console Tur;}  
{\f1063\fwiss\fcharset0\fprq2 Malgun Gothic Western;} {\f1073\fwiss\fcharset0\fprq2 @Malgun Gothic Western;}  
{\f1083\fwiss\fcharset0\fprq2 Meiryu Western;} {\f1081\fwiss\fcharset238\fprq2 Meiryu CE;}  
{\f1082\fwiss\fcharset204\fprq2 Meiryu Cyr;} {\f1084\fwiss\fcharset161\fprq2 Meiryu Greek;}  
{\f1085\fwiss\fcharset162\fprq2 Meiryu Tur;}  
{\f1088\fwiss\fcharset186\fprq2 Meiryu Baltic;} {\f1093\fwiss\fcharset0\fprq2 @Meiryu Western;}  
{\f1091\fwiss\fcharset238\fprq2 @Meiryu CE;} {\f1092\fwiss\fcharset204\fprq2 @Meiryu Cyr;}  
{\f1094\fwiss\fcharset161\fprq2 @Meiryu Greek;}  
{\f1095\fwiss\fcharset162\fprq2 @Meiryu Tur;} {\f1098\fwiss\fcharset186\fprq2 @Meiryu Baltic;}  
{\f1103\fwiss\fcharset0\fprq2 Meiryu UI Western;} {\f1101\fwiss\fcharset238\fprq2 Meiryu UI CE;}  
{\f1102\fwiss\fcharset204\fprq2 Meiryu UI Cyr;}  
{\f1104\fwiss\fcharset161\fprq2 Meiryu UI Greek;} {\f1105\fwiss\fcharset162\fprq2 Meiryu UI Tur;}  
{\f1108\fwiss\fcharset186\fprq2 Meiryu UI Baltic;} {\f1113\fwiss\fcharset0\fprq2 @Meiryu UI Western;}  
{\f1111\fwiss\fcharset238\fprq2 @Meiryu UI CE;}  
{\f1112\fwiss\fcharset204\fprq2 @Meiryu UI Cyr;} {\f1114\fwiss\fcharset161\fprq2 @Meiryu UI Greek;}

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{\f1115\fswiss\fcharset162\fpqr2 @Meiry UI Tur;} {\f1118\fswiss\fcharset186\fpqr2 @Meiry UI Baltic;}  
{\f1133\fswiss\fcharset0\fpqr2 Microsoft JhengHei Western;} {\f1134\fswiss\fcharset161\fpqr2 Microsoft JhengHei  
Greek;} {\f1143\fswiss\fcharset0\fpqr2 @Microsoft JhengHei Western;} {\f1144\fswiss\fcharset161\fpqr2 @Microsoft  
JhengHei Greek;}  
{\f1153\fswiss\fcharset0\fpqr2 Microsoft YaHei Western;} {\f1151\fswiss\fcharset238\fpqr2 Microsoft YaHei CE;}  
{\f1152\fswiss\fcharset204\fpqr2 Microsoft YaHei Cyr;} {\f1154\fswiss\fcharset161\fpqr2 Microsoft YaHei Greek;}  
{\f1155\fswiss\fcharset162\fpqr2 Microsoft YaHei Tur;} {\f1163\fswiss\fcharset0\fpqr2 @Microsoft YaHei Western;}  
{\f1161\fswiss\fcharset238\fpqr2 @Microsoft YaHei CE;} {\f1162\fswiss\fcharset204\fpqr2 @Microsoft YaHei Cyr;}  
{\f1164\fswiss\fcharset161\fpqr2 @Microsoft YaHei Greek;} {\f1165\fswiss\fcharset162\fpqr2 @Microsoft YaHei  
Tur;} {\f1173\fmmodern\fcharset0\fpqr1 @MingLiU Western;} {\f1183\froman\fcharset0\fpqr2 @PMingLiU Western;}  
{\f1193\froman\fcharset0\fpqr2 @MingLiU\_HKSCS Western;} {\f1203\froman\fcharset0\fpqr2 MingLiU-ExtB  
Western;} {\f1213\froman\fcharset0\fpqr2 @MingLiU-ExtB Western;} {\f1223\froman\fcharset0\fpqr2  
PMingLiU-ExtB Western;}  
{\f1233\froman\fcharset0\fpqr2 @PMingLiU-ExtB Western;} {\f1243\froman\fcharset0\fpqr2  
MingLiU\_HKSCS-ExtB Western;} {\f1253\froman\fcharset0\fpqr2 @MingLiU\_HKSCS-ExtB Western;}  
{\f1273\fmmodern\fcharset0\fpqr1 @MS Gothic Western;}  
{\f1271\fmmodern\fcharset238\fpqr1 @MS Gothic CE;} {\f1272\fmmodern\fcharset204\fpqr1 @MS Gothic Cyr;}  
{\f1274\fmmodern\fcharset161\fpqr1 @MS Gothic Greek;} {\f1275\fmmodern\fcharset162\fpqr1 @MS Gothic Tur;}  
{\f1278\fmmodern\fcharset186\fpqr1 @MS Gothic Baltic;}  
{\f1283\fswiss\fcharset0\fpqr2 MS PGothic Western;} {\f1281\fswiss\fcharset238\fpqr2 MS PGothic CE;}  
{\f1282\fswiss\fcharset204\fpqr2 MS PGothic Cyr;} {\f1284\fswiss\fcharset161\fpqr2 MS PGothic Greek;}  
{\f1285\fswiss\fcharset162\fpqr2 MS PGothic Tur;}  
{\f1288\fswiss\fcharset186\fpqr2 MS PGothic Baltic;} {\f1293\fswiss\fcharset0\fpqr2 @MS PGothic Western;}  
{\f1291\fswiss\fcharset238\fpqr2 @MS PGothic CE;} {\f1292\fswiss\fcharset204\fpqr2 @MS PGothic Cyr;}  
{\f1294\fswiss\fcharset161\fpqr2 @MS PGothic Greek;}  
{\f1295\fswiss\fcharset162\fpqr2 @MS PGothic Tur;} {\f1298\fswiss\fcharset186\fpqr2 @MS PGothic Baltic;}  
{\f1303\fswiss\fcharset0\fpqr2 MS UI Gothic Western;} {\f1301\fswiss\fcharset238\fpqr2 MS UI Gothic CE;}  
{\f1302\fswiss\fcharset204\fpqr2 MS UI Gothic Cyr;} {\f1304\fswiss\fcharset161\fpqr2 MS UI Gothic Greek;}  
{\f1305\fswiss\fcharset162\fpqr2 MS UI Gothic Tur;} {\f1308\fswiss\fcharset186\fpqr2 MS UI Gothic Baltic;}  
{\f1313\fswiss\fcharset0\fpqr2 @MS UI Gothic Western;} {\f1311\fswiss\fcharset238\fpqr2 @MS UI Gothic CE;}  
{\f1312\fswiss\fcharset204\fpqr2 @MS UI Gothic Cyr;} {\f1314\fswiss\fcharset161\fpqr2 @MS UI Gothic Greek;}  
{\f1315\fswiss\fcharset162\fpqr2 @MS UI Gothic Tur;} {\f1318\fswiss\fcharset186\fpqr2 @MS UI Gothic Baltic;}  
{\f1323\fmmodern\fcharset0\fpqr1 @MS Mincho Western;} {\f1321\fmmodern\fcharset238\fpqr1 @MS Mincho CE;}  
{\f1322\fmmodern\fcharset204\fpqr1 @MS Mincho Cyr;} {\f1324\fmmodern\fcharset161\fpqr1 @MS Mincho Greek;}  
{\f1325\fmmodern\fcharset162\fpqr1 @MS Mincho Tur;} {\f1328\fmmodern\fcharset186\fpqr1 @MS Mincho Baltic;}

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{\f1333\froman\charset0\prq2 MS PMincho Western;} {\f1331\froman\charset238\prq2 MS PMincho CE;}  
{\f1332\froman\charset204\prq2 MS PMincho Cyr;} {\f1334\froman\charset161\prq2 MS PMincho Greek;}  
{\f1335\froman\charset162\prq2 MS PMincho Tur;}  
{\f1338\froman\charset186\prq2 MS PMincho Baltic;} {\f1343\froman\charset0\prq2 @MS PMincho Western;}  
{\f1341\froman\charset238\prq2 @MS PMincho CE;} {\f1342\froman\charset204\prq2 @MS PMincho Cyr;}  
{\f1344\froman\charset161\prq2 @MS PMincho Greek;}  
{\f1345\froman\charset162\prq2 @MS PMincho Tur;} {\f1348\froman\charset186\prq2 @MS PMincho Baltic;}  
{\f1371\fnil\charset238\prq2 Nyala CE;} {\f1375\fnil\charset162\prq2 Nyala Tur;} {\f1378\fnil\charset186\prq2  
Nyala Baltic;}  
{\f1401\fwiss\charset238\prq2 Segoe Script CE;} {\f1402\fwiss\charset204\prq2 Segoe Script Cyr;}  
{\f1404\fwiss\charset161\prq2 Segoe Script Greek;} {\f1405\fwiss\charset162\prq2 Segoe Script Tur;}  
{\f1408\fwiss\charset186\prq2 Segoe Script Baltic;} {\f1411\fwiss\charset238\prq2 Segoe UI Semibold CE;}  
{\f1412\fwiss\charset204\prq2 Segoe UI Semibold Cyr;} {\f1414\fwiss\charset161\prq2 Segoe UI Semibold  
Greek;}  
{\f1415\fwiss\charset162\prq2 Segoe UI Semibold Tur;} {\f1418\fwiss\charset186\prq2 Segoe UI Semibold  
Baltic;} {\f1419\fwiss\charset163\prq2 Segoe UI Semibold (Vietnamese);} {\f1421\fwiss\charset238\prq2 Segoe  
UI Light CE;}  
{\f1422\fwiss\charset204\prq2 Segoe UI Light Cyr;} {\f1424\fwiss\charset161\prq2 Segoe UI Light Greek;}  
{\f1425\fwiss\charset162\prq2 Segoe UI Light Tur;} {\f1428\fwiss\charset186\prq2 Segoe UI Light Baltic;}  
{\f1429\fwiss\charset163\prq2 Segoe UI Light (Vietnamese);} {\f1443\fnil\charset0\prq2 @SimSun Western;}  
{\f1453\fmodern\charset0\prq1 NSimSun Western;} {\f1463\fmodern\charset0\prq1 @NSimSun Western;}  
{\f1473\fmodern\charset0\prq1 SimSun-ExtB Western;} {\f1483\fmodern\charset0\prq1 @SimSun-ExtB Western;}  
{\f1521\fwiss\charset238\prq2 Microsoft Sans Serif CE;} {\f1522\fwiss\charset204\prq2 Microsoft Sans Serif  
Cyr;}  
{\f1524\fwiss\charset161\prq2 Microsoft Sans Serif Greek;} {\f1525\fwiss\charset162\prq2 Microsoft Sans Serif  
Tur;} {\f1526\fbidi \fwiss\charset177\prq2 Microsoft Sans Serif (Hebrew);}  
{\f1527\fbidi \fwiss\charset178\prq2 Microsoft Sans Serif (Arabic);} {\f1528\fwiss\charset186\prq2 Microsoft  
Sans Serif Baltic;} {\f1529\fwiss\charset163\prq2 Microsoft Sans Serif (Vietnamese);}  
{\f1530\fwiss\charset222\prq2 Microsoft Sans Serif (Thai);} {\f1541\fnil\charset238\prq2 Ebrima CE;}  
{\f1545\fnil\charset162\prq2 Ebrima Tur;} {\f1548\fnil\charset186\prq2 Ebrima Baltic;} {\f1556\fbidi  
\fwiss\charset177\prq2 Gisha (Hebrew);}  
{\f1580\fwiss\charset222\prq2 Leelawadee (Thai);} {\f1587\fbidi \fnil\charset178\prq2 Microsoft Uighur  
(Arabic);} {\f1627\fbidi \froman\charset178\prq2 Andalus (Arabic);} {\f1631\fscript\charset238\prq2 Arabic  
Typesetting CE;}  
{\f1635\fscript\charset162\prq2 Arabic Typesetting Tur;} {\f1637\fbidi \fscript\charset178\prq2 Arabic Typesetting

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(Arabic);}{\f1638\fscrip\ fcharset186\ fprq2 Arabic Typesetting Baltic;}  
{\f1647\fbidi \froman\ fcharset178\ fprq2 Simplified Arabic (Arabic);}{\f1657\fbidi \fmodern\ fcharset178\ fprq1  
Simplified Arabic Fixed (Arabic);}{\f1661\fnil\ fcharset238\ fprq2 Sakkal Majalla CE;}{\f1665\fnil\ fcharset162\ fprq2  
Sakkal Majalla Tur;}  
{\f1667\fbidi \fnil\ fcharset178\ fprq2 Sakkal Majalla (Arabic);}{\f1668\fnil\ fcharset186\ fprq2 Sakkal Majalla Baltic;}  
{\f1677\fbidi \froman\ fcharset178\ fprq2 Traditional Arabic (Arabic);}{\f1763\ fmodern\ fcharset0\ fprq1 FangSong  
Western;}  
{\f1773\ fmodern\ fcharset0\ fprq1 @FangSong Western;}{\f1783\ fmodern\ fcharset0\ fprq1 @SimHei Western;}  
{\f1793\ fmodern\ fcharset0\ fprq1 KaiTi Western;}{\f1803\ fmodern\ fcharset0\ fprq1 @KaiTi Western;}  
{\f1820\ froman\ fcharset222\ fprq2 AngsanaUPC (Thai);}  
{\f1830\ fswiss\ fcharset222\ fprq2 Browallia New (Thai);}{\f1840\ fswiss\ fcharset222\ fprq2 BrowalliaUPC (Thai);}  
{\f1850\ fswiss\ fcharset222\ fprq2 CordiaUPC (Thai);}{\f1860\ froman\ fcharset222\ fprq2 DilleniaUPC (Thai);}  
{\f1870\ froman\ fcharset222\ fprq2 EucrosiaUPC (Thai);}{\f1880\ fswiss\ fcharset222\ fprq2 FreesiaUPC (Thai);}  
{\f1890\ fswiss\ fcharset222\ fprq2 IrisUPC (Thai);}{\f1900\ froman\ fcharset222\ fprq2 JasmineUPC (Thai);}  
{\f1910\ froman\ fcharset222\ fprq2 KodchiangUPC (Thai);}{\f1920\ fswiss\ fcharset222\ fprq2 LilyUPC (Thai);}  
{\f1923\ fscript\ fcharset0\ fprq1 DFKai-SB Western;}{\f1933\ fscript\ fcharset0\ fprq1 @DFKai-SB Western;}  
{\f1941\ fswiss\ fcharset238\ fprq2 Lucida Sans Unicode CE;}{\f1942\ fswiss\ fcharset204\ fprq2 Lucida Sans Unicode  
Cyr;}{\f1944\ fswiss\ fcharset161\ fprq2 Lucida Sans Unicode Greek;}{\f1945\ fswiss\ fcharset162\ fprq2 Lucida Sans  
Unicode Tur;}  
{\f1946\fbidi \fswiss\ fcharset177\ fprq2 Lucida Sans Unicode (Hebrew);}{\f1948\ fswiss\ fcharset186\ fprq2 Lucida  
Sans Unicode Baltic;}{\f1951\ fswiss\ fcharset238\ fprq2 Arial Black CE;}{\f1952\ fswiss\ fcharset204\ fprq2 Arial  
Black Cyr;}  
{\f1954\ fswiss\ fcharset161\ fprq2 Arial Black Greek;}{\f1955\ fswiss\ fcharset162\ fprq2 Arial Black Tur;}  
{\f1958\ fswiss\ fcharset186\ fprq2 Arial Black Baltic;}{\f1961\ fswiss\ fcharset238\ fprq2 Candara CE;}  
{\f1962\ fswiss\ fcharset204\ fprq2 Candara Cyr;}  
{\f1964\ fswiss\ fcharset161\ fprq2 Candara Greek;}{\f1965\ fswiss\ fcharset162\ fprq2 Candara Tur;}  
{\f1968\ fswiss\ fcharset186\ fprq2 Candara Baltic;}{\f1969\ fswiss\ fcharset163\ fprq2 Candara (Vietnamese);}  
{\f1971\ fscript\ fcharset238\ fprq2 Comic Sans MS CE;}  
{\f1972\ fscript\ fcharset204\ fprq2 Comic Sans MS Cyr;}{\f1974\ fscript\ fcharset161\ fprq2 Comic Sans MS Greek;}  
{\f1975\ fscript\ fcharset162\ fprq2 Comic Sans MS Tur;}{\f1978\ fscript\ fcharset186\ fprq2 Comic Sans MS Baltic;}  
{\f1981\ fmodern\ fcharset238\ fprq1 Consolas CE;}{\f1982\ fmodern\ fcharset204\ fprq1 Consolas Cyr;}  
{\f1984\ fmodern\ fcharset161\ fprq1 Consolas Greek;}{\f1985\ fmodern\ fcharset162\ fprq1 Consolas Tur;}  
{\f1988\ fmodern\ fcharset186\ fprq1 Consolas Baltic;}  
{\f1989\ fmodern\ fcharset163\ fprq1 Consolas (Vietnamese);}{\f1991\ froman\ fcharset238\ fprq2 Constantia CE{\*falt  
Constantia};}{\f1992\ froman\ fcharset204\ fprq2 Constantia Cyr{\*falt Constantia};}

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{\f1994\froman\fcharset161\fpqr2 Constantia Greek {\\*\falt Constantia};} {\f1995\froman\fcharset162\fpqr2 Constantia Tur {\\*\falt Constantia};} {\f1998\froman\fcharset186\fpqr2 Constantia Baltic {\\*\falt Constantia};} {\f1999\froman\fcharset163\fpqr2 Constantia (Vietnamese) {\\*\falt Constantia};} {\f2001\fswiss\fcharset238\fpqr2 Corbel CE;} {\f2002\fswiss\fcharset204\fpqr2 Corbel Cyr;} {\f2004\fswiss\fcharset161\fpqr2 Corbel Greek;} {\f2005\fswiss\fcharset162\fpqr2 Corbel Tur;} {\f2008\fswiss\fcharset186\fpqr2 Corbel Baltic;} {\f2009\fswiss\fcharset163\fpqr2 Corbel (Vietnamese);} {\f2011\fswiss\fcharset238\fpqr2 Franklin Gothic Medium CE;} {\f2012\fswiss\fcharset204\fpqr2 Franklin Gothic Medium Cyr;} {\f2014\fswiss\fcharset161\fpqr2 Franklin Gothic Medium Greek;} {\f2015\fswiss\fcharset162\fpqr2 Franklin Gothic Medium Tur;} {\f2018\fswiss\fcharset186\fpqr2 Franklin Gothic Medium Baltic;} {\f2021\fddecor\fcharset238\fpqr2 Gabriola CE;} {\f2022\fddecor\fcharset204\fpqr2 Gabriola Cyr;} {\f2024\fddecor\fcharset161\fpqr2 Gabriola Greek;} {\f2025\fddecor\fcharset162\fpqr2 Gabriola Tur;} {\f2028\fddecor\fcharset186\fpqr2 Gabriola Baltic;} {\f2031\froman\fcharset238\fpqr2 Georgia CE;} {\f2032\froman\fcharset204\fpqr2 Georgia Cyr;} {\f2034\froman\fcharset161\fpqr2 Georgia Greek;} {\f2035\froman\fcharset162\fpqr2 Georgia Tur;} {\f2038\froman\fcharset186\fpqr2 Georgia Baltic;} {\f2041\froman\fcharset238\fpqr2 Palatino Linotype CE;} {\f2042\froman\fcharset204\fpqr2 Palatino Linotype Cyr;} {\f2044\froman\fcharset161\fpqr2 Palatino Linotype Greek;} {\f2045\froman\fcharset162\fpqr2 Palatino Linotype Tur;} {\f2048\froman\fcharset186\fpqr2 Palatino Linotype Baltic;} {\f2049\froman\fcharset163\fpqr2 Palatino Linotype (Vietnamese);} {\f2051\fnil\fcharset238\fpqr2 Segoe Print CE;} {\f2052\fnil\fcharset204\fpqr2 Segoe Print Cyr;} {\f2054\fnil\fcharset161\fpqr2 Segoe Print Greek;} {\f2055\fnil\fcharset162\fpqr2 Segoe Print Tur;} {\f2058\fnil\fcharset186\fpqr2 Segoe Print Baltic;} {\f2061\fswiss\fcharset238\fpqr2 Trebuchet MS CE;} {\f2062\fswiss\fcharset204\fpqr2 Trebuchet MS Cyr;} {\f2064\fswiss\fcharset161\fpqr2 Trebuchet MS Greek;} {\f2065\fswiss\fcharset162\fpqr2 Trebuchet MS Tur;} {\f2068\fswiss\fcharset186\fpqr2 Trebuchet MS Baltic;} {\f2071\fswiss\fcharset238\fpqr2 Verdana CE;} {\f2072\fswiss\fcharset204\fpqr2 Verdana Cyr;} {\f2074\fswiss\fcharset161\fpqr2 Verdana Greek;} {\f2075\fswiss\fcharset162\fpqr2 Verdana Tur;} {\f2078\fswiss\fcharset186\fpqr2 Verdana Baltic;} {\f2079\fswiss\fcharset163\fpqr2 Verdana (Vietnamese);} {\f2091\froman\fcharset238\fpqr2 Lucida Bright CE;} {\f2095\froman\fcharset162\fpqr2 Lucida Bright Tur;} {\f2098\froman\fcharset186\fpqr2 Lucida Bright Baltic;} {\f2101\fswiss\fcharset238\fpqr2 Lucida Sans CE;} {\f2102\fswiss\fcharset204\fpqr2 Lucida Sans Cyr;} {\f2104\fswiss\fcharset161\fpqr2 Lucida Sans Greek;} {\f2105\fswiss\fcharset162\fpqr2 Lucida Sans Tur;} {\f2106\fbidi\fswiss\fcharset177\fpqr2 Lucida Sans (Hebrew);} {\f2107\fbidi\fswiss\fcharset178\fpqr2 Lucida Sans (Arabic);}

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{\f2108\fswiss\fcharset186\fpqr2 Lucida Sans Baltic;} {\f2110\fswiss\fcharset222\fpqr2 Lucida Sans (Thai);} {\f2123\fswiss\fcharset0\fpqr2 @Arial Unicode MS Western;} {\f2121\fswiss\fcharset238\fpqr2 @Arial Unicode MS CE;} {\f2122\fswiss\fcharset204\fpqr2 @Arial Unicode MS Cyr;} {\f2124\fswiss\fcharset161\fpqr2 @Arial Unicode MS Greek;} {\f2125\fswiss\fcharset162\fpqr2 @Arial Unicode MS Tur;} {\f2126\fbidi\fswiss\fcharset177\fpqr2 @Arial Unicode MS (Hebrew);} {\f2127\fbidi\fswiss\fcharset178\fpqr2 @Arial Unicode MS (Arabic);} {\f2128\fswiss\fcharset186\fpqr2 @Arial Unicode MS Baltic;} {\f2129\fswiss\fcharset163\fpqr2 @Arial Unicode MS (Vietnamese);} {\f2130\fswiss\fcharset222\fpqr2 @Arial Unicode MS (Thai);} {\f2185\froman\fcharset162\fpqr2 Bodoni MT Poster Compressed Tur;} {\f2191\froman\fcharset238\fpqr2 Book Antiqua CE;} {\f2192\froman\fcharset204\fpqr2 Book Antiqua Cyr;} {\f2194\froman\fcharset161\fpqr2 Book Antiqua Greek;} {\f2195\froman\fcharset162\fpqr2 Book Antiqua Tur;} {\f2198\froman\fcharset186\fpqr2 Book Antiqua Baltic;} {\f2201\froman\fcharset238\fpqr2 Bookman Old Style CE;} {\f2202\froman\fcharset204\fpqr2 Bookman Old Style Cyr;} {\f2204\froman\fcharset161\fpqr2 Bookman Old Style Greek;} {\f2205\froman\fcharset162\fpqr2 Bookman Old Style Tur;} {\f2208\froman\fcharset186\fpqr2 Bookman Old Style Baltic;} {\f2261\fswiss\fcharset238\fpqr2 Century Gothic CE;} {\f2262\fswiss\fcharset204\fpqr2 Century Gothic Cyr;} {\f2264\fswiss\fcharset161\fpqr2 Century Gothic Greek;} {\f2265\fswiss\fcharset162\fpqr2 Century Gothic Tur;} {\f2268\fswiss\fcharset186\fpqr2 Century Gothic Baltic;} {\f2321\froman\fcharset238\fpqr2 Garamond CE;} {\f2322\froman\fcharset204\fpqr2 Garamond Cyr;} {\f2324\froman\fcharset161\fpqr2 Garamond Greek;} {\f2325\froman\fcharset162\fpqr2 Garamond Tur;} {\f2328\froman\fcharset186\fpqr2 Garamond Baltic;} {\f2451\fscrip\fcharset238\fpqr2 Mistral CE;} {\f2452\fscrip\fcharset204\fpqr2 Mistral Cyr;} {\f2454\fscrip\fcharset161\fpqr2 Mistral Greek;} {\f2455\fscrip\fcharset162\fpqr2 Mistral Tur;} {\f2458\fscrip\fcharset186\fpqr2 Mistral Baltic;} {\f2471\fscrip\fcharset238\fpqr2 Monotype Corsiva CE;} {\f2472\fscrip\fcharset204\fpqr2 Monotype Corsiva Cyr;} {\f2474\fscrip\fcharset161\fpqr2 Monotype Corsiva Greek;} {\f2475\fscrip\fcharset162\fpqr2 Monotype Corsiva Tur;} {\f2478\fscrip\fcharset186\fpqr2 Monotype Corsiva Baltic;} {\f2681\fswiss\fcharset238\fpqr2 MS Reference Sans Serif CE;} {\f2682\fswiss\fcharset204\fpqr2 MS Reference Sans Serif Cyr;} {\f2684\fswiss\fcharset161\fpqr2 MS Reference Sans Serif Greek;} {\f2685\fswiss\fcharset162\fpqr2 MS Reference Sans Serif Tur;} {\f2688\fswiss\fcharset186\fpqr2 MS Reference Sans Serif Baltic;} {\f2689\fswiss\fcharset163\fpqr2 MS Reference Sans Serif (Vietnamese);} {\f2711\fswiss\fcharset238\fpqr2 Arial Narrow CE;}

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{\f2712\fswiss\fcharset204\fpqr2 Arial Narrow Cyr;} {\f2714\fswiss\fcharset161\fpqr2 Arial Narrow Greek;}  
{\f2715\fswiss\fcharset162\fpqr2 Arial Narrow Tur;} {\f2718\fswiss\fcharset186\fpqr2 Arial Narrow Baltic;}  
{\f2731\fswiss\fcharset238\fpqr2 Calibri Light CE;} {\f2732\fswiss\fcharset204\fpqr2 Calibri Light Cyr;}  
{\f2734\fswiss\fcharset161\fpqr2 Calibri Light Greek;} {\f2735\fswiss\fcharset162\fpqr2 Calibri Light Tur;}  
{\f2738\fswiss\fcharset186\fpqr2 Calibri Light Baltic;} {\f2739\fswiss\fcharset163\fpqr2 Calibri Light ( Vietnamese);}  
{\f2831\froman\fcharset238\fpqr2 Century Schoolbook CE;} {\f2832\froman\fcharset204\fpqr2 Century Schoolbook  
Cyr;}  
{\f2834\froman\fcharset161\fpqr2 Century Schoolbook Greek;} {\f2835\froman\fcharset162\fpqr2 Century  
Schoolbook Tur;} {\f2838\froman\fcharset186\fpqr2 Century Schoolbook Baltic;} {\f2961\fswiss\fcharset238\fpqr2  
Franklin Gothic Book CE;}  
{\f2962\fswiss\fcharset204\fpqr2 Franklin Gothic Book Cyr;} {\f2964\fswiss\fcharset161\fpqr2 Franklin Gothic Book  
Greek;} {\f2965\fswiss\fcharset162\fpqr2 Franklin Gothic Book Tur;} {\f2968\fswiss\fcharset186\fpqr2 Franklin  
Gothic Book Baltic;}  
{\f2971\fswiss\fcharset238\fpqr2 Franklin Gothic Demi CE;} {\f2972\fswiss\fcharset204\fpqr2 Franklin Gothic Demi  
Cyr;} {\f2974\fswiss\fcharset161\fpqr2 Franklin Gothic Demi Greek;} {\f2975\fswiss\fcharset162\fpqr2 Franklin  
Gothic Demi Tur;}  
{\f2978\fswiss\fcharset186\fpqr2 Franklin Gothic Demi Baltic;} {\f2981\fswiss\fcharset238\fpqr2 Franklin Gothic  
Demi Cond CE;} {\f2982\fswiss\fcharset204\fpqr2 Franklin Gothic Demi Cond Cyr;} {\f2984\fswiss\fcharset161\fpqr2  
Franklin Gothic Demi Cond Greek;}  
{\f2985\fswiss\fcharset162\fpqr2 Franklin Gothic Demi Cond Tur;} {\f2988\fswiss\fcharset186\fpqr2 Franklin Gothic  
Demi Cond Baltic;} {\f2991\fswiss\fcharset238\fpqr2 Franklin Gothic Heavy CE;} {\f2992\fswiss\fcharset204\fpqr2  
Franklin Gothic Heavy Cyr;}  
{\f2994\fswiss\fcharset161\fpqr2 Franklin Gothic Heavy Greek;} {\f2995\fswiss\fcharset162\fpqr2 Franklin Gothic  
Heavy Tur;} {\f2998\fswiss\fcharset186\fpqr2 Franklin Gothic Heavy Baltic;} {\f3001\fswiss\fcharset238\fpqr2  
Franklin Gothic Medium Cond CE;}  
{\f3002\fswiss\fcharset204\fpqr2 Franklin Gothic Medium Cond Cyr;} {\f3004\fswiss\fcharset161\fpqr2 Franklin  
Gothic Medium Cond Greek;} {\f3005\fswiss\fcharset162\fpqr2 Franklin Gothic Medium Cond Tur;}  
{\f3008\fswiss\fcharset186\fpqr2 Franklin Gothic Medium Cond Baltic;} {\f3031\fswiss\fcharset238\fpqr2 Gill Sans  
MT Ext Condensed Bold CE;} {\f3041\fswiss\fcharset238\fpqr2 Gill Sans MT CE;} {\f3051\fswiss\fcharset238\fpqr2  
Gill Sans MT Condensed CE;}  
{\f3061\fswiss\fcharset238\fpqr2 Gill Sans Ultra Bold CE;} {\f3071\fswiss\fcharset238\fpqr2 Gill Sans Ultra Bold  
Condensed CE;} {\f3111\fswiss\fcharset238\fpqr2 Haettenschweiler CE;} {\f3112\fswiss\fcharset204\fpqr2  
Haettenschweiler Cyr;}  
{\f3114\fswiss\fcharset161\fpqr2 Haettenschweiler Greek;} {\f3115\fswiss\fcharset162\fpqr2 Haettenschweiler Tur;}  
{\f3118\fswiss\fcharset186\fpqr2 Haettenschweiler Baltic;} {\f3271\fswiss\fcharset238\fpqr2 Tw Cen MT CE;}

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{f3281\fwiss\fcharset238\fpqr2 Tw Cen MT Condensed CE;} {f3291\fwiss\fcharset238\fpqr2 Tw Cen MT Condensed Extra Bold CE;} {f3301\fwiss\fcharset238\fpqr2 Nina CE;} {f3302\fwiss\fcharset204\fpqr2 Nina Cyr;} {f3304\fwiss\fcharset161\fpqr2 Nina Greek;} {f3305\fwiss\fcharset162\fpqr2 Nina Tur;} {f3308\fwiss\fcharset186\fpqr2 Nina Baltic;} {f3311\fwiss\fcharset238\fpqr2 Segoe Condensed CE;} {f3312\fwiss\fcharset204\fpqr2 Segoe Condensed Cyr;} {f3314\fwiss\fcharset161\fpqr2 Segoe Condensed Greek;} {f3315\fwiss\fcharset162\fpqr2 Segoe Condensed Tur;} {f3318\fwiss\fcharset186\fpqr2 Segoe Condensed Baltic;} {\colorbl\red0\green0\blue0;\red0\green0\blue255;\red0\green255\blue255;\red0\green255\blue0;\red255\green0\blue255;\red255\green0\blue0;\red255\green255\blue0;\red255\green255\blue255;\red0\green0\blue128;\red0\green128\blue128;\red0\green128\blue0;\red128\green0\blue128;\red128\green0\blue0;\red128\green128\blue0;\red128\green128\blue128;\red192\green192\blue192;} {\stylesheet{ \ql \li0\ri0\widctlpar\wrapdefault\aspalpha\aspnum\faauto\adjustright\ri0\lin0\itap0 \rtlch\fcs1 \af0\afs20\alang1025 \ltrch\fcs0 \fs24\lang1033\langfe1033\cgrid\langnp1033\langfenp1033 \snext0 \styrsid13782716 Normal;} {\s1\qc \li0\ri0\sl192\smult1 \keepn\widctlpar\tx475\tx950\tx1425\tx1900\tx2376\tx2851\tx3326\tx3801\tx4276\tx4752\tx5227\tx5702\tx6177\tx6652\tx7128\tx7603\tx8078\tx8553\tx9028\wrapdefault\aspalpha\aspnum\faauto\outlinelevel0\adjustright\ri0\lin0\itap0 \rtlch\fcs1 \af0\afs20\alang1025 \ltrch\fcs0 \b\fs24\ul\lang1033\langfe1033\cgrid\langnp1033\langfenp1033 \sbasedon0 \snext0 \slink15 \styrsid13782716 heading 1;} {\s2\ql \li0\ri0\sl192\smult1\keepn\nowidctlpar \tx475\tx950\tx1425\tx1900\tx2376\tx2851\tx3326\tx3801\tx4276\tx4752\tx5227\tx5702\tx6177\tx6652\tx7128\tx7603\tx8078\tx8553\tx9028\wrapdefault\aspalpha\aspnum\faauto\outlinelevel1\adjustright\ri0\lin0\itap0 \rtlch\fcs1 \af0\afs20\alang1025 \ltrch\fcs0 \b\fs24\ul\lang1033\langfe1033\cgrid\langnp1033\langfenp1033 \sbasedon0 \snext0 \slink16 \styrsid13782716 heading 2;} {\s3\ql \li0\ri0\sl192\smult1\keepn\widctlpar \tx475\tx950\tx1425\tx1900\tx2376\tx2851\tx3326\tx3801\tx4276\tx4752\tx5227\tx5702\tx6177\tx6652\tx7128\tx7603\tx8078\tx8553\tx9028\wrapdefault\aspalpha\aspnum\faauto\outlinelevel2\adjustright\ri0\lin0\itap0 \rtlch\fcs1 \af0\afs20\alang1025 \ltrch\fcs0 \b\fs52\lang1033\langfe1033\cgrid\langnp1033\langfenp1033 \sbasedon0 \snext0 \slink17 \styrsid13782716 heading 3;} {\s4\ql \li0\ri0\sl192\smult1\keepn\widctlpar \tx475\tx950\tx1425\tx1900\tx2376\tx2851\tx3326\tx3801\tx4276\tx4752\tx5227\tx5702\tx6177\tx6652\tx7128\tx7603\tx8078\tx8553\tx9028\wrapdefault\aspalpha\aspnum\faauto\outlinelevel3\adjustright\ri0\lin0\itap0 \rtlch\fcs1 \af0\afs20\alang1025 \ltrch\fcs0 \b\fs40\lang1033\langfe1033\cgrid\langnp1033\langfenp1033 \sbasedon0 \snext0 \slink18 \styrsid13782716 heading 4;} {\s5\ql \li0\ri0\keepn\widctlpar\wrapdefault\aspalpha\aspnum\faauto\outlinelevel4\adjustright\ri0\lin0\itap0

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**FED - D5447 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(d)(3)(i)(g)

**Type** Standard

**Regulation Definition**

Unless CMS Approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub. 7), that provides equivalent quality testing, the laboratory must--

At least once a day patient specimens are assayed or examined perform the following for--

Each quantitative procedure, include two control materials of different concentrations;

**Interpretive Guideline**

[Also see Interpretative guidelines under D5445 for Equivalent quality testing which apply to all of §493.1256(d)]

Interpretative Guidelines §493.1256(d)(3)

Laboratories generally need to follow manufacturers' test system instructions for control performance and meet the requirements in this section. The laboratory must determine if more extensive (e.g., number, frequency) control testing is necessary. Use D5425.

Immunology:

Determine which immunological methods the laboratory uses and how the laboratory tests quality control materials to check each test component of the test system. Examples of test systems that have multiple components are:

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(g) The laboratory must document all control procedures performed.

- o Complement Fixation (CF);
- o Hemagglutination inhibition (HAI);
- o Radio-immunoassay (RIA);
- o Enzyme immunoassay (EIA);
- o Indirect immunofluorescence (IFA);
- o Fluorescence Polarization Immunoassay (FPIA);
- o Radioimmunoprecipitin assay (RIPA); and
- o Radioallergosorbent test (RAST).

Use D5449 or D5451, as appropriate.

Syphilis Serology:

For FTA-ABS tests, does the laboratory employ:

- o Reactive control serum in Phosphatase Buffered Solution (PBS);
- o Reactive control serum in sorbent;
- o Minimally reactive control (1+);
- o Non-specific serum control in PBS;
- o Non-specific serum control in sorbent;
- o Non-specific staining control of PBS; and
- o Non-specific staining control of sorbent?

For MHATP or HATTS tests, does the laboratory employ:

- o Reactive reference control material;
- o Non-reactive reference control material;
- o Unsensitized erythrocyte with each specimen;
- o Unsensitized erythrocyte with buffer;
- o Sensitized erythrocyte with buffer;
- o Unsensitized erythrocyte with each reactive control serum; and
- o Unsensitized erythrocyte with non-reactive control serum?

Use D5451as appropriate.

Interpretative Guidelines §493.1256(d)(3)(i)

For monitoring the abnormal range, the laboratory must select controls that correlate with the patient values either in terms of specimen matrix or range to be evaluated. A laboratory must not use control materials outside the patient reportable range. Control samples not containing the analytes or substances to be controlled are not acceptable as control material.

Routine Chemistry:

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For monitoring the abnormal range, the laboratory should select control materials that correlate with the patient values both in terms of specimen matrix and range to be evaluated. For example, an elevated serum based bilirubin control should be employed when measuring neonatal bilirubins; a low level protein control or cerebrospinal fluid control should be used for monitoring cerebrospinal fluid protein.

**Hematology:**

For instruments which perform hemoglobin, hematocrit, red and white blood cell counts, platelets and/or differentials, acceptable controls are 2 levels of assayed materials, OR 1 level of assayed material and 1 patient specimen that was verified in the same batch of specimens with the assayed control material. The laboratory must establish criteria for an acceptable range of performance as required at D5481.

**EXCEPTION:**

Unless otherwise required by the test system's manufacturer or the laboratory's performance specifications, for instruments that perform white blood cell differentials directly from blood films (smears), a commercial control or patient specimen (differential) that has been verified through repetitive testing is an acceptable control and satisfies the requirements of §493.1256(d), as appropriate.

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

**FED - D5449 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(d)(3)(ii)(g)

**Type** Standard

**Regulation Definition**

Unless CMS Approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub. 7), that provides equivalent quality testing, the laboratory must--

At least once a day patient specimens are assayed or examined perform the following for--

Each qualitative procedure, include a negative and positive control material;

**Interpretive Guideline**

[Also see Interpretative guidelines under D5445 for Equivalent quality testing which apply to all of §493.1256(d)]

[Also see Interpretative Guidelines §493.1256(d)(3) under D5447]

Interpretative Guidelines §493.1256(d)(3)(ii)

Urinalysis - Photomicrographs or charts of all possible urine sediment components will meet the control requirement for manual microscopic urinalysis examinations.

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Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

(g) The laboratory must document all control procedures performed.

**FED - D5451 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(d)(3)(iii)(g)

**Type** Standard

**Regulation Definition**

Unless CMS Approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub. 7), that provides equivalent quality testing, the laboratory must--

At least once a day patient specimens are assayed or examined perform the following for--

Test procedures producing graded or titered results include a negative control material and a control material with graded or titered reactivity, respectively;

Interpretive Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

**Interpretive Guideline**

[Also see Interpretative guidelines under D5445 for Equivalent quality testing which apply to all of §493.1256(d)]

[Also see Interpretative Guidelines §493.1256(d)(3) under D5447]

Interpretative Guidelines §493.1256(d)(3)(iii)

For tests in which patient results are reported in terms of graded reactivity (1+, 2+, 3+, etc.) control(s) of graded reactivity must be used. For tests in which patient results are reported as a titer, controls of known titer must be used.

**EXCEPTIONS:**

A negative control is not required for anti-streptolysin O titer or anti-hyaluronidase titer tests. A positive control is not required for the cold agglutination test. For radial immuno-diffusion, one control or calibration material is required on each plate.

**FED - D5453 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(d)(3)(iv)(g)

**Type** Standard

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**Regulation Definition**

Unless CMS Approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub. 7), that provides equivalent quality testing, the laboratory must--

At least once a day patient specimens are assayed or examined perform the following for--

Each test system that has an extraction phase, include two control materials, including one that is capable of detecting errors in the extraction process;

(g) The laboratory must document all control procedures performed.

**Interpretive Guideline**

[Also see Interpretative guidelines under D5445 for Equivalent quality testing which apply to all of §493.1256(d)]

[Also see Interpretative Guidelines §493.1256(d)(3) under D5447]

Interpretative Guidelines §493.1256(d)(3)(iv)

Bacteriology:

For direct antigen systems, laboratories may use bacterial cell suspensions to meet the requirement for control organisms since the cell suspensions are subjected to both the extraction and reaction phases of the test. However, a matrix similar to patient specimens is preferred. For example, for direct antigen tests for group A streptococcal antigen, commercially prepared, dried (solid-shafted) swabs, one containing group A streptococcus (*S. pyogenes*) as a positive control and another with non-group A streptococcus and/or *Staphylococcus aureus* as a negative control may be used.

Additionally, if the manufacturer's instructions do not specify what the positive control contains, the laboratory should contact the manufacturer to ensure that the positive control contains a cell suspension of the organism. Otherwise, the laboratory must have an alternative mechanism for meeting this requirement (e.g., laboratory suspension stock American Type Culture Collection (ATCC) organism, commercially prepared organism controls).

Toxicology:

For comprehensive broad spectrum qualitative drug screening, procedures using gas chromatography, a control material containing one or more drugs representative of each drug class reported (e.g., tricyclic antidepressants, barbiturates), must go through each test phase, including the extraction process.

NOTE: For gas chromatography and mass spectrometry used for drug confirmations, an analyte-specific control is required for both qualitative and quantitative tests.

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

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**FED - D5455 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(d)(3)(v)(g)

**Type** Standard

**Regulation Definition**

Unless CMS Approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub. 7), that provides equivalent quality testing, the laboratory must--

At least once a day patient specimens are assayed or examined perform the following for--

Each molecular amplification procedure, include two control materials and, if reaction inhibition is a significant source of false negative results, a control material capable of detecting the inhibition.

(g) The laboratory must document all control procedures performed.

**Interpretive Guideline**

[Also see Interpretative guidelines under D5445 for Equivalent quality testing which apply to all of §493.1256(d)]

[Also see Interpretative Guidelines §493.1256(d)(3) under D5447]

Interpretative Guidelines §493.1256(d)(3)(iii)

The laboratory is also responsible for following the manufacturer's instructions concerning procedure limitations for detecting nucleic acid target amplification sequences, when provided by the manufacturer.

If the laboratory suspects the presence of interfering substances (inhibitors), the laboratory is responsible for using a control material (in addition to positive and negative control materials) capable of detecting interfering substances. Patient specimens may contain substances (inhibitors) that interfere with the enzymatic reaction of a molecular amplification procedure. These interfering substances could affect the assay's sensitivity causing a false negative result. Interfering substances may include, but are not limited to, components within the patient specimen or exogenous substances introduced during the preanalytic and/or analytic phase of testing.

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

**FED - D5457 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(d)(4)(g)

**Type** Standard

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**Regulation Definition**

Unless CMS Approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub. 7), that provides equivalent quality testing, the laboratory must--

For thin layer chromatography--

- (4)(i) Spot each plate or card, as applicable, with a calibrator containing all known substances or drug groups, as appropriate, which are identified by thin layer chromatography and reported by the laboratory; and
- (4)(ii) Include at least one control material on each plate or card, as applicable, which must be processed through each step of patient testing, including extraction processes.

(g) The laboratory must document all control procedures performed.

**Interpretive Guideline**

Interpretative Guidelines §493.1256(d)(4)

For qualitative urine drug screens performed by thin layer chromatography, a negative control is not required. However, a control containing one or more drugs representative of each drug group reported (e.g., tricyclic antidepressants, barbiturates) that goes through each test phase (including the extraction process) is required.

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

**FED - D5459 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(d)(5)(g)

**Type** Standard

**Regulation Definition**

Unless CMS Approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub. 7), that provides equivalent quality testing, the laboratory must--

Each electrophoretic procedure include, concurrent with patient specimens, at least one control material containing the substances being identified or measured.

**Interpretive Guideline**

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

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(g) The laboratory must document all control procedures performed.

**FED - D5461 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(d)(6)(g)

**Type** Standard

**Regulation Definition**

Unless CMS Approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub. 7), that provides equivalent quality testing, the laboratory must--

Perform control material testing as specified in this paragraph before resuming patient testing when a complete change of reagents is introduced; major preventive maintenance is performed; or any critical part that may influence test performance is replaced.

(g) The laboratory must document all control procedures performed.

**Interpretive Guideline**

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

**FED - D5463 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(d)(7)(g)

**Type** Standard

**Regulation Definition**

Unless CMS Approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub. 7), that provides equivalent quality testing, the

**Interpretive Guideline**

Interpretative Guidelines §493.1256(d)(7)

The laboratory may use this requirement to assist in competency assessment determinations specified in subpart M.

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laboratory must--

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

Over time, rotate control material testing among all operators who perform the test.

(g) The laboratory must document all control procedures performed.

**FED - D5465 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(d)(8)(g)

**Type** Standard

**Regulation Definition**

Unless CMS Approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub. 7), that provides equivalent quality testing, the laboratory must--

Test control materials in the same manner as patient specimens.

(g) The laboratory must document all control procedures performed.

**Interpretive Guideline**

Interpretative Guidelines §493.1256(d)(8)

Control materials of a similar matrix to that of patient specimens should be utilized, if available, and the control materials must be treated in the same manner as patient specimens and go through all analytic test phases.

Flow Cytometry

In cell surface phenotyping by flow cytometry or fluorescent microscopy, control samples must be analyzed within the same time period after staining as test specimens.

Probes §493.1256(d)(8)

Flow Cytometry

How did the laboratory establish the time period in which stained cells must be analyzed to avoid significant loss of any cell subpopulations or total cell numbers?

If analysis will be based on a population of cells selected by flow cytometry "gating" on size or density parameters, or selected by depletion or enrichment techniques, are controls tested with each patient to detect the presence of contaminating cells in the selected population? (e.g., Monocyte contamination of "lymphocytes" gated by forward angle or forward angle versus 90 degrees light scatter must be detected with a monocyte-specific antibody.) Use

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D5465 or D5425 as appropriate.

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

**FED - D5467 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(d)(9)(g)

**Type** Standard

**Regulation Definition**

Unless CMS Approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub. 7), that provides equivalent quality testing, the laboratory must--

When using calibration material as a control material, use calibration material from a different lot number than that used to establish a cut-off value or to calibrate the test system.

(g) The laboratory must document all control procedures performed.

**Interpretive Guideline**

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

**FED - D5469 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(d)(10)(g)

**Type** Standard

**Regulation Definition**

Unless CMS Approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub. 7), that provides equivalent quality testing, the

**Interpretive Guideline**

Interpretative Guidelines §493.1256(d)(10)

Acceptable ranges must be verified (assayed) or established (unassayed) by the laboratory for control materials and

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laboratory must--

Establish or verify the criteria for acceptability of all control materials.

(i) When control materials providing quantitative results are used, statistical parameters (for example, mean and standard deviation) for each batch and lot number of control materials must be defined and available.

(ii) The laboratory may use the stated value of a commercially assayed control material provided the stated value is for the methodology and instrumentation employed by the laboratory and is verified by the laboratory.

(iii) Statistical parameters for unassayed control materials must be established over time by the laboratory through concurrent testing of control materials having previously determined statistical parameters.

(g) The laboratory must document all control procedures performed.

any calibrators that are used in lieu of control materials.

For procedures in which a spiked sample is used as a control, an acceptable range must be established for the amount of recovery of the spiked sample, either in percentage or actual concentration.

If laboratories rely on commercial companies to establish statistical limits for controls, the laboratory must have documentation to verify that its control results correlate with the established limits.

When patient specimens are used to meet the control requirements, data must be evaluated in accordance with §493.1256(d)(10)(iii).

There are no specific guidelines for the number of times a material must be tested to establish statistical limits. In general, twenty replicate tests should be considered the minimum for determining a standard deviation.

Probes §493.1256(d)(10)

What statistics does the laboratory have to demonstrate the number of assays and the period of time in which the laboratory repetitively tested control materials to verify or establish control limits?

How does the laboratory evaluate control results to detect any outliers, shifts or trends in control values due to instrument malfunctions or changes in the analytical system?

If more than one test system is in use for a test procedure, did the laboratory evaluate the data for each test method in the establishment of control limits?

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

**FED - D5471 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(e)(1)(g)

**Type** Standard

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**Regulation Definition**

(e) For reagent, media, and supply checks, the laboratory must do the following:

(e)(i) Check each batch (prepared in-house), lot number (commercially prepared) and shipment of reagents, disks, stains, antisera, (except those specifically referenced in §493.1261 (a)(3)) and identification systems (systems using two or more substrates or two or more reagents, or a combination) when prepared or opened for positive and negative reactivity, as well as graded reactivity, if applicable.

(g) The laboratory must document all control procedures performed.

**Interpretive Guideline**

Interpretative Guidelines §493.1256(e)(1)

Review the laboratory's quality control records and note when lot numbers change.

NOTE: Media checks are defined under §493.1256(e)(4) Guidelines.

The laboratory must demonstrate that each reagent performs within the specifications established by the laboratory for the test procedure. Documentation of concurrent testing of reagents or acceptable quality control results will satisfy this requirement.

Reagents, disks, and test procedures used for identification purposes may include, but are not limited to, catalase, coagulase plasma, oxidase, bacitracin, optochin, CefinaseO, ONPG, X and V factor strips and disks, germ tube, yeast morphology media, and commercial identification systems.

A negative reactivity control is not required for the mycology germ tube test.

Test each batch, lot, and shipment for positive and negative reactivity for reagents such as:

- o Bacitracin;
- o Catalase;
- o Cefinase;
- o Coagulase plasma;
- o ONPG;
- o Optochin;
- o Oxidase;
- o Spot indole; and
- o X and V factor strips and disks.

For bacteriology, XV discs or strips need only be checked with an organism that produces a positive reaction.

Probes §493.1256(e)(1)

What records does the laboratory have to demonstrate that controls are tested when shipments of reagents, discs, stains, antisera or identification systems are opened or when the laboratory prepares these materials? Use D5471 for not recording performance and for nonperformance of quality control checks, and stain checks.

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

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**FED - D5473 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(e)(2)(g)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

(e) For reagent, media, and supply checks, the laboratory must do the following:

Interpretative Guidelines §493.1256(e)(2)

(e)(2) Each day of use (unless otherwise specified in this subpart), test staining materials for intended reactivity to ensure predictable staining characteristics. Control materials for both positive and negative reactivity must be included, as appropriate.

The laboratory must check routine stain Hematoxylin and Eosin each day for intended response, and predicted characteristics of the stain.

(g) The laboratory must document all control procedures performed.

Interpretative Guidelines §493.1256(e)(2)-(e)(3)

Acid-fast stains must be checked each day of use for positive and negative reactivity.  
Interpretative Guidelines §493.1256(e)(2)-(e)(3)

Acid-fast stains must be checked each day of use for positive and negative reactivity.

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

**FED - D5475 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(e)(3)(g)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

(e) For reagent, media, and supply checks, the laboratory must do the following:

Interpretative Guidelines §493.1256(e)(3)

(e)(3) Check fluorescent and immunohistochemical stains for positive and negative reactivity each time of use.

All fluorescent stains, including fluorochrome acid-fast stains, must be tested for positive and negative reactivity each time of use.

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Flow Cytometry

(g) The laboratory must document all control procedures performed.

Staining controls for cell surface immunophenotyping by flow cytometry should consist of either normal, cultured or abnormal cells known to be positive for selected standard antigens and must verify the proper performance of reagents. Frozen or other preserved cells may be used. A negative reagent control must be run for each test cell preparation, and is to consist of monoclonal antibody (ies) of the same species and isotype. Negative reagent controls will consist of:

- o For indirect stains, an irrelevant primary antibody, if available, and in all cases, the same secondary antibody(ies) conjugated with the same fluorochrome(s) used in all relevant test combinations; and
- o For direct stains, an irrelevant antibody conjugated to the same fluorochrome and at the same fluorochromes: protein ratio used in all relevant test combinations.

Probes §493.1256(e)(3)

For flow cell cytometric surface immunophenotyping, is a negative reagent control used to define a threshold for positive staining cells? If not, how does the laboratory define the threshold for positive staining cells?

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

**FED - D5477 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(e)(4)(g)

**Type** Standard

**Regulation Definition**

(e) For reagent, media, and supply checks, the laboratory must do the following:

(e)(4) Before, or concurrent with the initial use--

(e)(4)(i) Check each batch of media for sterility if sterility is required for testing;

(e)(4)(ii) Check each batch of media for its ability to support

**Interpretive Guideline**

Interpretative Guidelines §493.1256(e)(4)

A batch of media (solid, semi-solid, or liquid) consists of all tubes, plates, or containers of the same medium prepared at the same time and in the same laboratory; or, if received from an outside source or commercial supplier, consists of all of the plates, tubes or containers of the same medium that have the same lot numbers and are received in a single shipment.

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growth and, as appropriate, select or inhibit specific organisms or produce a biochemical response; and

(e)(4)(iii) Document the physical characteristics of the media when compromised and report any deterioration in the media to the manufacturer.

(g) The laboratory must document all control procedures performed.

A sample from each batch of media is sufficient as a check for:

- o Sterility, if it is autoclaved or filtered during preparation;
- o Ability to support growth, using at least one organism to demonstrate the ability of the media to support growth;
- o Selectivity and/or inhibition, using at least one organism to confirm its selective characteristic, and at least one organism to confirm its inhibitory characteristic; and
- o Biochemical response, using at least one organism which will produce the expected reaction (positive control) and with at least one organism which will not produce the expected reaction (negative control).

American Type Culture Collection (ATCC) control organisms are not necessarily required. However, if the laboratory uses "in-house" isolates for control organisms, it must have established reactivity for each organism. Use D5469 as appropriate.

Central laboratories that prepare media for satellite locations must either perform the same quality control checks required of commercial manufacturers and furnish documentation of media quality control checks to each satellite location, or each laboratory must continue to perform media checks as required under §493.1256(e)(4).

If a laboratory screens cultures for growth or no growth, reports "No growth" and refers all growth to a reference laboratory, the screening laboratory must perform applicable quality control of the media.

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

**FED - D5479 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(e)(5)(g)

**Type** Standard

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**Regulation Definition**

(e) For reagent, media, and supply checks, the laboratory must do the following:

(e)(5) Follow the manufacturer's specifications for using reagents, media, and supplies and be responsible for results.

(g) The laboratory must document all control procedures performed.

**Interpretive Guideline**

Interpretative Guidelines §493.1256(e)(5)

The laboratory must meet any and all regulatory requirements and comply with the manufacturer's recommendations and requirements for testing to the extent that the manufacturer's recommendations and requirements do not conflict with any regulatory requirements

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

**FED - D5481 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(f)(g)

**Type** Standard

**Regulation Definition**

(f) Results of control materials must meet the laboratory's and, as applicable, the manufacturer's test system criteria for acceptability before reporting patient test results.

(g) The laboratory must document all control procedures performed.

**Interpretive Guideline**

Interpretative Guidelines §493.1256(g)

The actual measurement(s) taken, reactions and/or observations must be recorded.

**FED - D5485 - CONTROL PROCEDURES**

**Title** CONTROL PROCEDURES

**CFR** 493.1256(h)

**Type** Standard

**Regulation Definition**

If control materials are not available, the laboratory must have an alternative mechanism to detect immediate errors and

**Interpretive Guideline**

Interpretative Guidelines §493.1256(h)

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monitor test system performance over time. The performance of alternative control procedures must be documented.

Laboratories may choose to split samples for testing by another method or in another laboratory to evaluate the results obtained. Previously tested patient specimens (include specimens across the reportable range) must be tested in duplicate. Precision is determined through replicate testing of a previously tested patient specimen. The duplicate tests may be performed by the same individual or by different people and the results compared to previously defined acceptable limits for differences between duplicates.

Public Health Laboratories Performing Newly Developed Assays/Test Systems for Agents for Emergent Public Health Significance

Screening and confirmation methods for agents of emergent public health significance require the rapid development and transfer of technology and expertise from federal agencies to public health laboratories (or other designee laboratories). Because of unique situations of emergent diseases or other public health threats, control and calibration materials for the assay or test system may not be immediately available. Under these circumstances, the laboratory must follow the assay or test system's protocol(s) without modification and document the alternative mechanisms employed to ensure accurate test results. Laboratories are encouraged to use multiple mechanisms (as described below) for ensuring accuracy.

When control and calibration materials are not available, examples of alternative control procedures that may be available include, but are not limited to, the following:

- o Split specimens for testing by another method or in another laboratory;
- o Include previously tested patient specimens (both positive and negative) tested in duplicate as surrogate controls;
- o Test each patient specimen in duplicate;
- o Test multiple specimen types from the same patient (e.g., saliva, urine, serum);
- o Perform serial dilutions of positive specimens to confirm positive reactions;
- o Provide additional supervisory review of results prior to release.

As soon as control and calibration materials become available, the applicable requirements in §493.1256 must be met.

For specific information regarding testing for agents of emergent public health significance and alternative

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methods/procedures for ensuring accuracy of this testing, refer to <<http://www.aphl.org/>>.

Probes §493.1256(h)

If control materials are not provided by the manufacturer, how does the laboratory ensure the validity of test results?

**FED - D5501 - BACTERIOLOGY**

**Title** BACTERIOLOGY

**CFR** 493.1261(a)(1)

**Type** Standard

**Regulation Definition**

(a) The laboratory must check the following for positive and negative reactivity using control organisms:

(a)(1) Each day of use for beta-lactamase methods other than Cefinase(trademark).

**Interpretive Guideline**

Interpretative Guidelines §493.1261(a)

When condition level deficiencies in Bacteriology are in any or all phases of testing, use D5002.

For direct antigen systems, laboratories may use bacterial cell suspensions to meet the requirement for control organisms since the cell suspensions are subjected to both the extraction and reaction phases of the test. However, a matrix similar to patient specimens is preferred. For example, for direct antigen tests for group A streptococcal antigen, already prepared, dried (solid-shafted) swabs, one containing group A streptococcus (*S. pyogenes*) as a positive control and another with non-group A streptococcus and/or *Staphylococcus aureus* as a negative control may be used. Use D5449 to cite a laboratory that fails both a negative and positive control. Use D5453 for deficiencies related to the extraction process.

Additionally, if the manufacturer's instructions do not specify what the positive control contains, the laboratory should contact the manufacturer to ensure that the positive control contains a cell suspension of the organism. Otherwise, the laboratory must have an alternative mechanism for meeting this requirement (e.g., laboratory suspension stock ATCC organism, commercially prepared organism controls).

For microbial identification systems utilizing two or more substrates, the laboratory must check each media using control organisms to verify positive and negative reactivity of each substrate. Use D5471 for deficiencies in this area.

If a laboratory utilizes primary isolation media (e.g., MacConkey, CLED, EMB), for presumptive identification of organisms, then the media should meet the quality control requirements at D5471 and D5477.

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For bacitracin, catalase, coagulase plasma, desoxycholate, oxidase, optochin, PYR disks, spot indole, staphylococcal latex reagents, streptococcal latex grouping reagents, and X and V factor strips and disks, use D5471.

For bacteriology, XV discs or strips need only be checked with an organism that produces a positive reaction. Use D5471.

For guidelines for molecular amplification testing, use D5455.

Interpretative Guidelines §493.1261(a)(1)

Beta-lactamase testing performed by acidometric, iodometric or chromogenic methodologies other than Cefinase (trademark) must have positive and negative reactivity checked each day of use.

For Cefinase (trademark), use D5471.

**FED - D5503 - BACTERIOLOGY**

**Title** BACTERIOLOGY

**CFR** 493.1261(a)(2)

**Type** Standard

**Regulation Definition**

(a) The laboratory must check the following for positive and negative reactivity using control organisms:

(a)(2) Each week of use for gram stains.

**Interpretive Guideline**

Interpretative Guidelines §493.1261(a)

When condition level deficiencies in Bacteriology are in any or all phases of testing, use D5002.

For direct antigen systems, laboratories may use bacterial cell suspensions to meet the requirement for control organisms since the cell suspensions are subjected to both the extraction and reaction phases of the test. However, a matrix similar to patient specimens is preferred. For example, for direct antigen tests for group A streptococcal antigen, already prepared, dried (solid-shafted) swabs, one containing group A streptococcus (*S. pyogenes*) as a positive control and another with non-group A streptococcus and/or *Staphylococcus aureus* as a negative control may be used. Use D5449 to cite a laboratory that fails both a negative and positive control. Use D5453 for deficiencies related to the extraction process.

Additionally, if the manufacturer's instructions do not specify what the positive control contains, the laboratory should contact the manufacturer to ensure that the positive control contains a cell suspension of the organism. Otherwise, the

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laboratory must have an alternative mechanism for meeting this requirement (e.g., laboratory suspension stock ATCC organism, commercially prepared organism controls).

For microbial identification systems utilizing two or more substrates, the laboratory must check each media using control organisms to verify positive and negative reactivity of each substrate. Use D5471 for deficiencies in this area.

If a laboratory utilizes primary isolation media (e.g., MacConkey, CLED, EMB), for presumptive identification of organisms, then the media should meet the quality control requirements at D5471 and D5477.

For bacitracin, catalase, coagulase plasma, desoxycholate, oxidase, optochin, PYR disks, spot indole, staphylococcal latex reagents, streptococcal latex grouping reagents, and X and V factor strips and disks, use D5471.

For bacteriology, XV discs or strips need only be checked with an organism that produces a positive reaction. Use D5471.

For guidelines for molecular amplification testing, use D5455.

**FED - D5505 - BACTERIOLOGY**

**Title** BACTERIOLOGY

**CFR** 493.1261(a)(3)

**Type** Standard

**Regulation Definition**

(a) The laboratory must check the following for positive and negative reactivity using control organisms:

(a)(3) When each batch (prepared in-house), lot number (commercially prepared), and shipment of antisera is prepared or opened, and once every 6 months thereafter.

**Interpretive Guideline**

Interpretative Guidelines §493.1261(a)

When condition level deficiencies in Bacteriology are in any or all phases of testing, use D5002.

For direct antigen systems, laboratories may use bacterial cell suspensions to meet the requirement for control organisms since the cell suspensions are subjected to both the extraction and reaction phases of the test. However, a matrix similar to patient specimens is preferred. For example, for direct antigen tests for group A streptococcal antigen, already prepared, dried (solid-shafted) swabs, one containing group A streptococcus (*S. pyogenes*) as a positive control and another with non-group A streptococcus and/or *Staphylococcus aureus* as a negative control may be used. Use D5449 to cite a laboratory that fails both a negative and positive control. Use D5453 for deficiencies related to the extraction process.

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Additionally, if the manufacturer's instructions do not specify what the positive control contains, the laboratory should contact the manufacturer to ensure that the positive control contains a cell suspension of the organism. Otherwise, the laboratory must have an alternative mechanism for meeting this requirement (e.g., laboratory suspension stock ATCC organism, commercially prepared organism controls).

For microbial identification systems utilizing two or more substrates, the laboratory must check each media using control organisms to verify positive and negative reactivity of each substrate. Use D5471 for deficiencies in this area.

If a laboratory utilizes primary isolation media (e.g., MacConkey, CLED, EMB), for presumptive identification of organisms, then the media should meet the quality control requirements at D5471 and D5477.

For bacitracin, catalase, coagulase plasma, desoxycholate, oxidase, optochin, PYR disks, spot indole, staphylococcal latex reagents, streptococcal latex grouping reagents, and X and V factor strips and disks, use D5471.

For bacteriology, XV discs or strips need only be checked with an organism that produces a positive reaction. Use D5471.

For guidelines for molecular amplification testing, use D5455. Interpretative Guidelines §493.1261(a)(3) In addition to Salmonella and Shigella antisera, antisera used for serotyping of homologous isolates, (i.e., streptococcal serotyping systems) must be checked for positive and negative reactivity. Polyvalent antisera should be tested with at least one organism from each polyvalent group.

Requirements for antisera QC apply to testing that has a direct impact on patient care.

Interpretative Guidelines §493.1261(a)(3)

In addition to Salmonella and Shigella antisera, antisera used for serotyping of homologous isolates, (i.e., streptococcal serotyping systems) must be checked for positive and negative reactivity. Polyvalent antisera should be tested with at least one organism from each polyvalent group.

Requirements for antisera QC apply to testing that has a direct impact on patient care.

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**FED - D5507 - BACTERIOLOGY**

**Title** BACTERIOLOGY

**CFR** 493.1261(b)(c)

**Type** Standard

**Regulation Definition**

(b) For antimicrobial susceptibility tests, the laboratory must check each batch of media and each lot number and shipment of antimicrobial agent(s) before, or concurrent with, initial use, using approved control organisms.

(b)(1) Each day tests are performed, the laboratory must use the appropriate control organism(s) to check the procedure.

(b)(2) The laboratory's zone sizes or minimum inhibitory concentration for control organisms must be within established limits before reporting patient results.

(c) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1261(b)(1-2)

"Approved control organism(s)" means either an appropriate control strain or an equivalent strain as defined below.

The laboratory must ensure proper standardization of the inoculum (e.g., use a 0.5 McFarland standard or its optical equivalent, or follow manufacturer's instructions for a commercially available system).

Antimicrobial Disk Diffusion Susceptibility  
(Bauer, Kirby, Sherris and Turk Method)

Each new batch of medium and each new lot/shipment of antimicrobial disks must be checked as follows:

**ANTIMICROBIAL DISK SUSCEPTIBILITY TEST**

Appropriate Control Strain	Each New Batch of Media and Disks	Each Day If Isolates Are:
S. aureus ATCC 25923 or equivalent**	X	Staphylococcus spp.
E. coli ATCC 25922 or equivalent**	X	Enterobacteriaceae
P. aeruginosa ATCC 27853 and E. coli ATCC 25922 or equivalent**	X	Pseudomonas aeruginosa Acintebacter spp .

The above table provides guidance to surveyors of the checks required for each new batch of medium and each new lot/shipment of antimicrobial disks. These must be checked as follows:

1. S.aureus ATCC 25923 or equivalent must be used to test each new batch of media or disks and it must be used each day if the isolate is Staphylococcus spp.
2. E. coli ATCC 25922 or equivalent must be used to test each new batch of media or disks and it must be used each day if the isolate is Enterobacteriaceae spp.

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3. P.aeruginosa ATCC 27853 and E.coli ATCC 25922 or equivalent must be used to test each new batch of media or disks and it must be used each day if the isolate is Pseudomonas aeruginosa and/or Acetobacter spp.

Zone sizes must be recorded for each antimicrobial control and limits must be established.

\*\*An equivalent strain is one which demonstrates reactivity similar to an ATCC strain and for which limits have been established. Organisms which manufacturers recommend or require for use in their systems are acceptable strains of control organisms.

When testing is performed daily, for each antimicrobial agent/organism combination, 1 out of every 20 consecutive results may be out of the acceptable range. Any more than 1 out-of-control result in 20 consecutive tests requires corrective action.

Direct susceptibility testing is a modification of the standardized disk diffusion susceptibility testing method. Therefore, the laboratory must establish the interpretative zone diameters for patient specimens, as well as establish the zone diameters for quality control organisms.

**MINIMUM INHIBITORY CONCENTRATION (MIC)**

Each new batch of macrodilution tubes, microdilution trays, or agar dilution plates must be checked as follows:

**MINIMUM INHIBITORY CONCENTRATION (MIC)**

Appropriate Control Strain	Each New Batch of Media	Each Day If Isolates are:
S. aureus ATCC 29213 or equivalent**	X	Staphylococcus spp.
E. coli ATCC 25922 or equivalent**	X	Enterobacteriaceae
P. aeruginosa ATCC 27853 and E. coli ATCC 25922 or equivalent **	X	Non-Enterobacteriaceae
	to include	Acetobacter spp.,
Pseudomonas spp.	Stenotrophomonas maltophilia,	

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	and other nonfastidious, glucose nonfermenting, gram-negative bacilli	
E. faecalis ATCC 29212 or equivalent**	X	Enterococcus spp.

The above table provides guidance to surveyors of the checks required for each new batch of macrodilution tubes, microdilution trays, or agar dilution plates. These must be checked as follows:

1. S.aureus ATCC 29213 or equivalent must be used to test each new batch of media and it must be used each day if the isolate is Staphylococcus spp.
2. E.coli ATCC 25922 or equivalent must be used to test each new batch of media and it must be used each day if the isolate is Enterobacteriaceae spp.
3. P.aeruginosa ATCC 27853 and E.coli ATCC 25922 or equivalent must be used to test each new batch of media and it must be used each day if the isolate is Non-Enterobacteriaceae to include Acinetobacter spp., Stenotrophomonas maltophilia, Pseudomonas spp. and/or other nonfastidious, glucose nonfermenting, gram-negative bacilli.
4. E.faecalis ATCC 29212 or equivalent must be used to test each new batch of media and it must be used each day if the isolate is Enterococcus spp.

\*\*An equivalent strain is one which demonstrates reactivity similar to an ATCC strain and for which limits have been established. Organisms which manufacturers recommend or require for use in their systems are acceptable strains of control organisms.

Each day the test is performed, the appropriate control strain(s) must be included to check the test system.

When testing is performed daily, for each antimicrobial agent/organism combination, 1 out of every 20 consecutive results may be out of the acceptable range. Any more than 1 out-of-control result in 20 consecutive tests requires corrective action.

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Interpretative Guidelines §493.1261(c)

QC records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reactions, and/or observations and demonstrate that controls were tested when shipments of reagents, disks, stains, or antisera for identification systems were opened or when the laboratory prepared these materials.

**FED - D5511 - MYCOBACTERIOLOGY**

**Title** MYCOBACTERIOLOGY

**CFR** 493.1262(a)(c)

**Type** Standard

**Regulation Definition**

Each day of use, the laboratory must check all reagents or test procedures used for mycobacteria identification with at least one acid-fast organism that produces a positive reaction and an acid-fast organism that produces a negative reaction.

(c) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1262(a)

When condition level deficiencies in Mycobacteriology are identified in any or all phases of testing, use D5004.

For acid-fast stains (i.e., Ziehl-Neelsen, Kinyoun), use positive and negative stain controls each day of testing patient samples. Use D5473 for deficiencies in these practices. For fluorochrome acid-fast stains, use positive and negative stain controls each time of use. Use D5475 for deficiencies in these practices.

Controls for acid-fast and fluorochrome stains for clinical specimens may include previously processed specimens that contain confirmed acid-fast organisms such as *Mycobacterium fortuitum* or other non-tuberculous mycobacteria for the positive control, and a negative sputum seeded with *Escherichia coli* for a negative control. Control smears should be heat-fixed and stored in a protective box.

For controls when staining mycobacteriology cultures, use a previously confirmed acid-fast organism such as *Mycobacterium fortuitum* for the positive control, and a non-mycobacterial species such as *Escherichia coli* for the negative control.

For the BACTEC NAP test, positive and negative control organisms must be tested each week of use. Controls should include *M. tuberculosis* ATCC 27294 and *M. kansasii* ATCC 35775. *M. tuberculosis* should be inhibited by NAP, while *M. kansasii* should have increasing growth index values in the presence of NAP.

For molecular amplification testing guidelines, use D5455.

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Interpretative Guidelines §493.1262(c)

QC records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reactions, and/or observations and demonstrate that controls were tested when shipments of reagents, disks, stains, or antisera for identification systems were opened or when the laboratory prepared these materials.

Probes §493.1262(a)

How often are mycobacteriology cultures checked for growth prior to the issuance of final patient reports? How long are negative cultures held before a final patient report is issued (e.g., minimum of six weeks)? Use D5411 and D5413 as appropriate.

**FED - D5513 - MYCOBACTERIOLOGY**

**Title** MYCOBACTERIOLOGY

**CFR** 493.1262(b)(c)

**Type** Standard

**Regulation Definition**

(b) For antimycobacterial susceptibility tests, the laboratory must check each batch of media and each lot number and shipment of antimycobacterial agent(s) before, or concurrent with, initial use, using an appropriate control organism(s).

(b)(1) The laboratory must establish limits for acceptable control results.

(b)(2) Each week tests are performed, the laboratory must use the appropriate control organism(s) to check the procedure.

(b)(3) The results for the control organism(s) must be within established limits before reporting patient results.

(c) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1262(b)

A susceptible control strain of Mycobacterium tuberculosis, such as H37Rv or other appropriate control strain, must be used to check the susceptibility procedure.

For automated mycobacterial susceptibility testing, organisms which manufacturers recommend or require for use in their systems are acceptable strains of control organisms.

Probes §493.1262(b)

Are quality control samples tested at the same time specimens are tested? For example, a growth control without antimycobacterial agent should be inoculated at the time of patient testing.

Probes §493.1262(b)(1)

Which control strains are used and how did the laboratory establish acceptable control limits for susceptibility tests?

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Interpretative Guidelines 493.1262(b)(3)

The laboratory must ensure that it performs and documents all corrective action(s) taken whenever the test results do not meet the laboratory control limits for susceptibility. Use D5783.

Interpretative Guidelines §493.1262(c)

QC records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reactions, and/or observations and demonstrate that controls were tested when shipments of reagents, disks, stains, or antisera for identification systems were opened or when the laboratory prepared these materials.

**FED - D5517 - MYCOLOGY**

**Title** MYCOLOGY

**CFR** 493.1263(a)(c)

**Type** Standard

**Regulation Definition**

The laboratory must check each batch (prepared in-house), lot number (commercially prepared), and shipment of lactophenol cotton blue when prepared or opened for intended reactivity with a control organism(s).

(c) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1263(a)

When condition-level deficiencies in Mycology are identified in any or all phases of testing, use D5006.

For non-culture identification systems (e.g., direct antigen) use D5449 and/or D5453 as appropriate.

For mycology identification systems utilizing two or more substrates, the laboratory must check each media using control organisms to verify positive and negative reactivity of substrate. Use D5471.

A filamentous fungus such as *Aspergillus* species should be used to check staining of lactophenol cotton blue.

Interpretative Guidelines §493.1263(c)

QC records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reactions, and/or observations and demonstrate that controls were tested when shipments of reagents, discs, stains, or antisera for identification systems were opened or when the laboratory prepared these materials.

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**FED - D5519 - MYCOLOGY**

**Title** MYCOLOGY

**CFR** 493.1263(b)(c)

**Type** Standard

**Regulation Definition**

(b)For antifungal susceptibility tests, the laboratory must check each batch of media and each lot number and shipment of antifungal agent(s) before, or concurrent with, initial use, using an appropriate control organism(s).

(b)(1) The laboratory must establish limits for acceptable control results.

(b)(2) Each day tests are performed, the laboratory must use the appropriate control organism(s) to check the procedure.

(b)(3) The results for the control organism(s) must be within established limits before reporting patient results.

(c) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Probes §493.1263(b)(1)

Which control strains are used and how did the laboratory establish acceptable control limits for susceptibility tests?

Probes §493.1263(b)(2)

Are quality control samples tested at the same time specimens are tested?

Interpretative Guidelines §493.1263(c)

QC records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reactions, and/or observations and demonstrate that controls were tested when shipments of reagents, discs, stains, or antisera for identification systems were opened or when the laboratory prepared these materials.

**FED - D5523 - PARASITOLOGY**

**Title** PARASITOLOGY

**CFR** 493.1264(a)(d)

**Type** Standard

**Regulation Definition**

The laboratory must have available a reference collection of slides or photographs and, if available, gross specimens for identification of parasites and use these references in the laboratory for appropriate comparison with diagnostic

**Interpretive Guideline**

Interpretative Guidelines §493.1264(a)

When condition level deficiencies in Parasitology are identified in any or all phases of testing, use D5008.

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specimens.

(d) The laboratory must document all control procedures performed, as specified in this section.

The laboratory must have adequate reference material, but does not have to maintain several different reference systems. Textbooks with photographs, previously stained slide preparations, preserved specimens, or slides from proficiency testing programs are some acceptable systems.

If the laboratory uses zinc sulfate for concentration of fecal specimens for ova and parasite examinations, the acceptable specific gravity of the zinc sulfate solution is 1.18 for fresh fecal samples and 1.20 for formalinized fecal samples. Use D5411 as applicable.

For non-culture identification systems (e.g., direct antigen) use D5449 and/or D5453 as appropriate.

Interpretative Guidelines §493.1264(d)

QC records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reactions, and/or observations and demonstrate that controls were tested when shipments of reagents, disks, stains, or antisera for identification systems were opened or when the laboratory prepared these materials. QC records should also include documentation of the measurements and calculations for calibration of each objective (low, high, oil immersion) of the ocular micrometer, and demonstrate that permanent stain controls were tested with a fecal sample control material each month of use.

**FED - D5525 - PARASITOLOGY**

**Title** PARASITOLOGY

**CFR** 493.1264(b)(d)

**Type** Standard

**Regulation Definition**

The laboratory must calibrate and use the calibrated ocular micrometer for determining the size of ova and parasites, if size is a critical parameter.

(d) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1264(b)

Check for the following:

- o Presence of an ocular micrometer for the microscope(s) used;
- o Availability of a stage micrometer;
- o Instructions for calibration. Use D5403;

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- o Records of the measurements and calculations used to show that each objective (high, oil, low) has been calibrated; and
- o Criteria for the use of the micrometer for determining the size of ova and parasites. Use D5403.

Interpretative Guidelines §493.1264(d)

QC records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reactions, and/or observations and demonstrate that controls were tested when shipments of reagents, disks, stains, or antisera for identification systems were opened or when the laboratory prepared these materials. QC records should also include documentation of the measurements and calculations for calibration of each objective (low, high, oil immersion) of the ocular micrometer, and demonstrate that permanent stain controls were tested with a fecal sample control material each month of use.

Probes §493.1264(b)

How has the laboratory determined the accuracy of the ocular calibration and that the staff has the knowledge for proper use?

**FED - D5527 - PARASITOLOGY**

**Title** PARASITOLOGY

**CFR** 493.1264(c)(d)

**Type** Standard

**Regulation Definition**

(c) Each month of use, the laboratory must check permanent stains using a fecal sample control material that will demonstrate staining characteristics.

(d) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1264(c)

The fecal sample control may contain either parasites or added leukocytes sufficient to demonstrate staining characteristics. A commercially prepared quality control slide for intestinal parasites is also an acceptable control for checking permanent stains.

While a wet mount preparation may not be sufficiently sensitive to detect small numbers of ova or parasites in fecal specimens, or to render a final species identification, the regulations do not require use of concentrated and permanent stain techniques to identify fecal parasites. It is the laboratory's responsibility to ensure that it can accurately and reliably identify the organisms it claims to be able to identify. Use D3007 and/or D5411 as applicable.

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Upon request, the laboratory must specify the method employed by the laboratory for screening fecal specimens and provide information to clients on the test report that may affect the interpretation of test results. Use D5807 and/or D5809 as applicable.

The working iodine solution is stable for approximately two weeks. If the laboratory does not prepare fresh working iodine solution at least every two weeks, it must ensure that the iodine solution has not deteriorated by observing positive clinical specimens or formalin-fixed specimens. Use D5417. Protozoan cysts stained with iodine contain golden yellow cytoplasm, brown glycogen material and have refractile nuclei.

Interpretative Guidelines §493.1264(d)

QC records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reactions, and/or observations and demonstrate that controls were tested when shipments of reagents, disks, stains, or antisera for identification systems were opened or when the laboratory prepared these materials. QC records should also include documentation of the measurements and calculations for calibration of each objective (low, high, oil immersion) of the ocular micrometer, and demonstrate that permanent stain controls were tested with a fecal sample control material each month of use.

**FED - D5531 - VIROLOGY**

**Title** VIROLOGY

**CFR** 493.1265(a)(b)

**Type** Standard

**Regulation Definition**

(a) When using cell culture to isolate or identify viruses, the laboratory must simultaneously incubate a cell substrate control or uninoculated cells as a negative control material.

(b) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1265(a)

When condition level deficiencies in Virology are identified in any or all phases of testing, use D5010.

Any laboratory testing patient specimens for the Human Papillomavirus (HPV) must enroll and successfully participate in a CMS-approved proficiency testing program for HPV beginning in 2008. Laboratories should refer to Subpart H for further information. The laboratory's CLIA certificate must include the subspecialty of Virology. The laboratory must also be in compliance with all of the CLIA regulations governing the preanalytic, analytic, and post analytic phases of testing including proficiency testing and personnel requirement.

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Cell Culture

For commercially purchased cell culture media, the requirement for media quality control checks is satisfied by visually examining the media for sterility and ensuring the ability of the media to sustain cell life. If the media is prepared or produced in the laboratory, use D5477:

- o Each component of cell culture media should be checked for sterility using bacterial culture techniques. In addition, fetal bovine serum must be checked for toxicity using cell culture systems;
- o The combined product (e.g., Hanks, Eagles and Earles) should be checked for sterility using bacterial culture techniques and the ability to propagate growth with cell cultures; and
- o Cell culture systems should be checked for mycoplasma contamination at regular intervals established by the laboratory.

Non-Culture Methods

1o For other non-culture identification (e.g., antigen identification) systems that are used for viral identification, the laboratory is not required to maintain live viral cultures for quality control purposes. However, positive and negative controls are required to evaluate the detection phase, if such controls are available commercially or in the laboratory. Use D5449 and/or D5453 as appropriate.

2o If organism controls are not available, a previously extracted viral antigen as the positive control plus a previously confirmed negative control of the same matrix as the patient sample may be used. Use D5485. A positive organism control must be subjected to the extraction process if such a control is available in the laboratory. Use D5453.

3o For fluorescent stains, the control requirements are met by using virus-infected cells for a positive control among uninfected cells for a negative control. Use D5475.

The intent of the regulations is for the laboratory to have methodologies available to isolate and identify the viruses that are etiologically related to the clinical disease for which services are offered. For example, if a laboratory offers services only for Herpes testing, it must have available host systems for the isolation and/or test methods for the identification of the Herpes virus. If the laboratory is not using the appropriate host system, use D3007.

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"Host system" is defined as the animal, egg or cell culture model, which supports the propagation of viruses.

Clinical information important for the determination and selection of the proper host system should include (Use D5305):

- o Clinical symptoms of the patient;
- o Age of the patient;
- o Source of the specimen;
- o Date of onset of clinical symptoms;
- o Recent travel information of patient;
- o Test request; and
- o Date of specimen collection.

Cell culture is the host system used most frequently. The specific cell line (type) is usually selected based upon its known sensitivity and susceptibility to different viruses. For example, the cell lines to be used as host systems for the following clinical specimens could be:

- o Upper respiratory infection specimens: Primary Monkey Kidney (PMK), Human Fetal Diploid Lung (HFDL), or equivalent;
- o Enteric specimens: PMK, Human Fetal Diploid Kidney (HFDK), or equivalent;
- o Urine specimens: HFDL, PMK, or equivalent;
- o Genital specimens: Human Foreskin (HFD), Vero (Continuous Monkey Kidney), or equivalent;
- o Vesicular lesions: HFDL, PMK, BSC-1 (Monkey Cell Line), or equivalent; and
- o Tissues or Spinal fluids: PMK, Vero, BSC-1, HFDK or HFDL, or equivalent.

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Prior to the inoculation of the cell cultures, the laboratory should check the cell culture systems for the following:

- o The age of the cell culture monolayer (no more than 7-10 days post "seeding") (Use D5417);
- o Maintenance media that is free from inhibitory substances (Use D5477); and
- o Sterility (visual observation for turbidity) (Use D5477).

Uninoculated cell substrate controls are used to determine whether the specificity of a test system has been ensured. Generally, an uninoculated cell control for each cell line that is inoculated is used per inoculation day to determine whether the consequent cytopathic effect (CPE) in the cells inoculated with patient specimen was caused by specific etiologic agent(s), or caused by the nonspecific deterioration of the cells themselves. Often, as monolayer host cells age, the cells deteriorate, exhibiting "rounding" and "pulling-apart." This cell change may be confused with CPE if uninoculated cells are not available to compare with the inoculated cells.

Probes §493.1265(a)

How does the laboratory determine the specific cell line to be used as the host system? Use D3007 or D5411 as applicable.

When reviewing the laboratory's identification procedures for the clinical diseases for which services are offered, how does the laboratory rule out the presence of *Clostridium difficile* toxin in those cell cultures in which the patient specimen exhibits non-specific effects unrelated to viral cytopathic effect (CPE)? Use D3007 or D5411 as applicable.

If presumptive reports are issued based on CPE, how does the laboratory confirm the identification reported? Use D3007 or D5411 as applicable.

For tests such as hemagglutination inhibition and viral neutralization in which antisera must be standardized, how has the laboratory determined the optimum dilution of the antisera to ensure maximum sensitivity and specificity? Use D5437.

Neutralization Tests

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How does the laboratory standardize its dilution of the viral isolate and control virus to the appropriate Tissue Culture Dose 50 or equivalent, each time the test is performed? Use D5437.

How many varieties of uninoculated cell cultures does the laboratory use to check each new lot of anti-serum or serum pool for toxicity? Use D5477 or D5479 as applicable.

**Hemagglutination Inhibition Tests**

After having determined the hemagglutination titer, how does the laboratory determine the working dilution of the viral isolate (i.e., usually 4 Hemagglutination units)? How does the laboratory ensure that this working dilution is correct for isolates and controls? Use D5421 or D5423 as applicable.

How often and for which hemagglutination inhibition tests does the laboratory include a serum/cell/buffer control and a cell/buffer control? Use D5425.

Does the laboratory include one known virus or viral antigen specific to each antisera used in the test procedure? Use D5449.

**Direct Immunofluorescence Tests**

How does the laboratory determine which immune serum conjugate(s) to use when identifying viruses using antisera that react with viruses that are etiologically similar (e.g., an antigen test for specimens from patients with flu-like symptoms that identifies Respiratory Syncytial Virus, Influenza, and Parainfluenza)? How does the laboratory ensure the specificity of this conjugate for the specific virus being identified? Use D5421 or D5423 as applicable.

How does the laboratory rule out non-specific reactivity for each conjugate used? Use D5421 or D5423 as applicable.

**Indirect Immunofluorescence Tests**

Has the laboratory determined the optimum dilution of its anti-species, e.g., antibody to host system or cell culture (such as anti-PMK, conjugated immune serum)? Use D5421 or D5423 as applicable.

Has the laboratory determined the optimum dilution of the virus specific immune serum? Use D5421 or D5423 as applicable.

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Determine whether the laboratory is checking positive and negative reactivity using (Use D5475):

- o Uninoculated cells plus immune serum plus anti-species conjugate (negative control); and
- o Viral antigen or known virus infected cells plus immune serum plus anti-species conjugate (positive control).

Determine whether the laboratory checks each new batch or shipment of conjugate using known virus infected cells plus PBS plus anti-species conjugate. Use D5471.

Interpretative Guidelines §493.1265(b)

QC records must identify the host cell cultures employed, the number of tubes or plates inoculated or uninoculated, maintenance medium used, the number of times the patient specimen was sub-cultured, the specific sub-culture or passage in which the virus was identified, the CPE observed, and post inoculation date of observations. If the deficiency is due to absence of dates of testing and observations, use D5787.

**FED - D5535 - ROUTINE CHEMISTRY**

**Title** ROUTINE CHEMISTRY

**CFR** 493.1267(a)(d)

**Type** Standard

**Regulation Definition**

For blood gas analyses, the laboratory must perform the following:

- (a) Calibrate or verify calibration according to the manufacturer's specifications and with at least the frequency recommended by the manufacturer.
- (d) Document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1267(a)-(d)

When condition-level deficiencies in Routine Chemistry are identified in one or more phases of testing, use D5016.

Control materials generally are not available to verify the reportable range at the very high range of patient results. When necessary, the laboratory may verify the results by splitting patient samples and assaying them on two different blood gas analyzers.

Quality control records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reaction and/or observations and demonstrate that controls were tested as required.

Do not dictate the acceptable format for documentation.

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Probes §493.1267(a)-(d)

For blood gas testing, do the records include barometric pressure and room temperature, as necessary?

Do the records of a laboratory that moves from testing site to testing site demonstrate the performance of control samples following transport of equipment when such activity affects test performance specifications and/or instrument calibration?

Interpretative Guidelines §493.1267(a)

For blood gas analysis, the laboratory must perform calibration and calibration verification in accordance with the manufacturer's instructions. If the laboratory meets the manufacturer's instructions, and the requirements at this section, the laboratory does not have to adhere to calibration and calibration verification requirements at §493.1255.

**FED - D5537 - ROUTINE CHEMISTRY**

**Title** ROUTINE CHEMISTRY

**CFR** 493.1267(b)(d)

**Type** Standard

**Regulation Definition**

For blood gas analyses, the laboratory must perform the following:

- (b) Test one sample of control material each 8 hours of testing using a combination of control materials that include both low and high values on each day of testing.
- (d) Document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1267(a)-(d)

When condition-level deficiencies in Routine Chemistry are identified in one or more phases of testing, use D5016.

Control materials generally are not available to verify the reportable range at the very high range of patient results. When necessary, the laboratory may verify the results by splitting patient samples and assaying them on two different blood gas analyzers.

Quality control records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reaction and/or observations and demonstrate that controls were tested as required.

Do not dictate the acceptable format for documentation.

Probes §493.1267(a)-(d)

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For blood gas testing, do the records include barometric pressure and room temperature, as necessary?

Do the records of a laboratory that moves from testing site to testing site demonstrate the performance of control samples following transport of equipment when such activity affects test performance specifications and/or instrument calibration?

Interpretative Guideline §493.1267(b)

"Each 8 hours of testing" is defined as each shift of 8 consecutive hours the laboratory is in operation, including "on-call" shifts. When documenting standards/controls results, the laboratory must identify the shifts in which controls are tested with patients.

For a laboratory that is only open 8 hours/day and the instrument auto calibrates, the laboratory must test both a low and high value in the eight hours to meet the requirement.

In addition to testing one control each eight hours, the combination of controls and calibrators used each day of testing must include a high and low value. Controls should be rotated to check normal, alkalosis and acidosis levels.

**FED - D5539 - ROUTINE CHEMISTRY**

**Title** ROUTINE CHEMISTRY

**CFR** 493.1267(c)(d)

**Type** Standard

**Regulation Definition**

For blood gas analyses, the laboratory must perform the following:

(c) Test one sample of control material each time specimens are tested unless automated instrumentation internally verifies calibration at least every 30 minutes.

(d) Document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1267(a)-(d)

When condition-level deficiencies in Routine Chemistry are identified in one or more phases of testing, use D5016.

Control materials generally are not available to verify the reportable range at the very high range of patient results. When necessary, the laboratory may verify the results by splitting patient samples and assaying them on two different blood gas analyzers.

Quality control records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reaction and/or observations and demonstrate that controls were tested as required.

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Do not dictate the acceptable format for documentation.

Probes §493.1267(a)-(d)

For blood gas testing, do the records include barometric pressure and room temperature, as necessary?

Do the records of a laboratory that moves from testing site to testing site demonstrate the performance of control samples following transport of equipment when such activity affects test performance specifications and/or instrument calibration?

Interpretative Guidelines §493.1267(c)

If blood gas analysis is performed with an instrument that does not internally verify the calibration at least every thirty minutes, then a calibrator or control must be tested each time patient specimens are tested. It is not the intent of this requirement to require the laboratory to maintain records of each auto-calibration.

**FED - D5543 - HEMATOLOGY**

**Title** HEMATOLOGY

**CFR** 493.1269(a)(d)

**Type** Standard

**Regulation Definition**

- (a) For manual cell counts performed using a hemocytometer--
- (a)(1) One control material must be tested each 8 hours of operation; and
- (a)(2) Patient specimens and control materials must be tested in duplicate.
- (d) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1269(a)

When condition-level deficiencies in Hematology are identified in any or all phases of testing, use D5024.

For all manual cell counts performed using a hemocytometer (e.g., synovial fluids, CSF, semen) the laboratory may meet the requirement for duplicate testing by counting two chambers from one dilution.

"Hours of operation" is defined as each shift of 8 consecutive hours the laboratory is in operation, including "on-call" shifts. When documenting standards/controls results, the laboratory must identify the shifts in which controls are tested with patients.

If the manufacturer of an instrument that performs automated differentials does not give criteria for when to perform a

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manual differential, the laboratory must establish criteria indicating when to perform a manual differential including instructions for reporting the results. Use D5423.

Control requirements for automated instruments that perform hemoglobin, hematocrit, red and white cell counts and differentials are found at §493.1256(d)(3)(i). Use D5447. The calibration verification exception for automated cell counters is found at D5439.

Interpretative Guidelines §493.1269(d)

Quality control records should include lot numbers, date prepared/opened, expiration dates, the actual measurement(s) taken, reactions and/or observations and demonstrate that controls were tested when shipments of reagents or stains were opened or when the laboratory prepared these materials. However, do not dictate the acceptable format for documentation.

**FED - D5545 - HEMATOLOGY**

**Title** HEMATOLOGY

**CFR** 493.1269(b)(d)

**Type** Standard

**Regulation Definition**

(b) For all nonmanual coagulation test systems, the laboratory must include two levels of control material each 8 hours of operation and each time a reagent is changed.

(d) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1269(b)-(c)

The laboratory performing nonmanual coagulation tests subject to §493.1269 must either establish criteria or verify manufacturer's criteria for an acceptable range of performance as required in §493.1253(b). Use D5421 or D5423 as appropriate.

An automated (nonmanual) coagulation test system samples the plasma, combines the plasma with the reagents, detects the end point or clot formation and displays the test results without operator intervention.

The International Sensitivity Index (ISI) is the correction factor for variable sensitivities of thromboplastins. The International Normalized Ratio (INR) is a calculation primarily used for monitoring a patient's oral anticoagulant therapy. The INR corrects for the variability in Prothrombin Time (PT) results attributable to the ISI. Therefore, this allows all PT's to be corrected to the international standard.

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INR Calculation

The INR is equal to the ratio of the patient's PT (in seconds) to the laboratory's established normal mean PT (in seconds), then raised to the power of the ISI.

$$\text{INR} = (\text{Patient PT} / \text{Mean Normal Range PT})^{\text{ISI}}$$

NOTE: A scientific calculator is needed to calculate the INR.

Example:

Patient PT (in seconds) = 18.5

Normal mean PT (in seconds) = 12.9

ISI value (obtain from the package insert of the laboratory's current lot of thromboplastin reagent) = 2.002

1.  $18.5 / 12.9 = 1.434$  (Patient Ratio)
2.  $1.434^{2.002} = 2.056$  (INR Result)
3. Report the INR as: INR = 2.1

For International Normalized Ratio (INR) calculations, ensure that the laboratory:

- o Establishes a normal patient Prothrombin time mean with each new thromboplastin lot number;
- o Verifies that the normal patient Prothrombin time mean study has been performed according to the manufacturer's instructions;
- o Incorporates the current and pertinent normal patient Prothrombin time mean and ISI value for each lot of thromboplastin (manual, instrument, or LIS);
- o Documents the manual check of the INR calculation for each new lot number;
- o Documents each thromboplastin lot number, with the normal patient Prothrombin time mean and the ISI value provided by the manufacturer (manual or instrument);
- o Periodically verifies, for each thromboplastin lot number in use, the correct normal patient Prothrombin time mean and the International Sensitivity Index (ISI) value are being used for calculating the INR value; and

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- o Periodically verifies the accuracy of the INR calculation (manual, instrument or LIS).

To verify prothrombin time testing with INR calculations:

- o Check the accuracy of normal patient Prothrombin time mean calculation (manual, instrument or LIS).
- o Verify that the ISI used in the calculation correlates with the ISI specified in the reagent package insert. Select an abnormal low or abnormal high prothrombin time result and verify the calculation.

Probes §493.1269(b)-(c)

Is the laboratory using the ISI value from the current manufacturer's package insert in calculating the INR values?

How does the laboratory ensure that the ISI values are changed with each change of thromboplastin lot number?

Has the laboratory established its own normal patient mean with each lot of thromboplastin?

For coagulation testing, do the records include timer checks and temperature checks as necessary?

Interpretative Guidelines §493.1269(d)

Quality control records should include lot numbers, date prepared/opened, expiration dates, the actual measurement(s) taken, reactions and/or observations and demonstrate that controls were tested when shipments of reagents or stains were opened or when the laboratory prepared these materials. However, do not dictate the acceptable format for documentation.

**FED - D5547 - HEMATOLOGY**

**Title** HEMATOLOGY

**CFR** 493.1269(c)(d)

**Type** Standard

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**Regulation Definition**

- (c) For manual coagulation tests--
- (c)(1) Each individual performing tests must test two levels of control materials before testing patient samples and each time a reagent is changed; and
- (c)(2) Patient specimens and control materials must be tested in duplicate.
- (d) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1269(b)-(c)

The laboratory performing nonmanual coagulation tests subject to §493.1269 must either establish criteria or verify manufacturer's criteria for an acceptable range of performance as required in §493.1253(b). Use D5421 or D5423 as appropriate.

An automated (nonmanual) coagulation test system samples the plasma, combines the plasma with the reagents, detects the end point or clot formation and displays the test results without operator intervention.

The International Sensitivity Index (ISI) is the correction factor for variable sensitivities of thromboplastins. The International Normalized Ratio (INR) is a calculation primarily used for monitoring a patient's oral anticoagulant therapy. The INR corrects for the variability in Prothrombin Time (PT) results attributable to the ISI. Therefore, this allows all PT's to be corrected to the international standard.

**INR Calculation**

The INR is equal to the ratio of the patient's PT (in seconds) to the laboratory's established normal mean PT (in seconds), then raised to the power of the ISI.

$$\text{INR} = (\text{Patient PT} / \text{Mean Normal Range PT})^{\text{ISI}}$$

NOTE: A scientific calculator is needed to calculate the INR.

Example:

Patient PT (in seconds) =18.5

Normal mean PT (in seconds) =12.9

ISI value (obtain from the package insert of the laboratory's current lot of thromboplastin reagent) =2.002

1.  $18.5 / 12.9 = 1.434$  (Patient Ratio)
2.  $1.434^{2.002} = 2.056$  (INR Result)
3. Report the INR as: INR = 2.1

For International Normalized Ratio (INR) calculations, ensure that the laboratory:

- o Establishes a normal patient Prothrombin time mean with each new thromboplastin lot number;

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- o Verifies that the normal patient Prothrombin time mean study has been performed according to the manufacturer's instructions;
- o Incorporates the current and pertinent normal patient Prothrombin time mean and ISI value for each lot of thromboplastin (manual, instrument, or LIS);
- o Documents the manual check of the INR calculation for each new lot number;
- o Documents each thromboplastin lot number, with the normal patient Prothrombin time mean and the ISI value provided by the manufacturer (manual or instrument);
- o Periodically verifies, for each thromboplastin lot number in use, the correct normal patient Prothrombin time mean and the International Sensitivity Index (ISI) value are being used for calculating the INR value; and
- o Periodically verifies the accuracy of the INR calculation (manual, instrument or LIS).

To verify prothrombin time testing with INR calculations:

- o Check the accuracy of normal patient Prothrombin time mean calculation (manual, instrument or LIS).
- o Verify that the ISI used in the calculation correlates with the ISI specified in the reagent package insert. Select an abnormal low or abnormal high prothrombin time result and verify the calculation.

Probes §493.1269(b)-(c)

Is the laboratory using the ISI value from the current manufacturer's package insert in calculating the INR values?

How does the laboratory ensure that the ISI values are changed with each change of thromboplastin lot number?

Has the laboratory established its own normal patient mean with each lot of thromboplastin?

For coagulation testing, do the records include timer checks and temperature checks as necessary?

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Interpretative Guidelines §493.1269(d)

Quality control records should include lot numbers, date prepared/opened, expiration dates, the actual measurement(s) taken, reactions and/or observations and demonstrate that controls were tested when shipments of reagents or stains were opened or when the laboratory prepared these materials. However, do not dictate the acceptable format for documentation.

**FED - D5551 - IMMUNOHEMATOLOGY**

**Title** IMMUNOHEMATOLOGY

**CFR** 493.1271(a)(f)

**Type** Standard

**Regulation Definition**

- (a) Patient testing. (a)(1) The laboratory must perform ABO grouping, D (Rho) typing, unexpected antibody detection, antibody identification, and compatibility testing by following the manufacturer's instructions, if provided, and as applicable, 21 CFR 606.151(a) through (e).
- (a)(2) The laboratory must determine ABO group by concurrently testing unknown red cells with, at a minimum, anti-A and anti-B grouping reagents. For confirmation of ABO group, the unknown serum must be tested with known A1 and B red cells.
- (a)(3) The laboratory must determine the D (Rho) type by testing unknown red cells with anti-D (anti-Rho) blood typing reagent.
- (f) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1271(a)(1)

21 CFR §606.151 requires the following standard operating procedures for compatibility testing:

- (a) A method of collecting and identifying the blood samples of recipients to ensure positive identification.
- (b) The use of fresh recipient serum or plasma samples less than 3 days old for all pretransfusion testing if the recipient has been pregnant or transfused within the previous 3 months. If information on the patient's history of transfusion or pregnancy is not available, then a fresh specimen is to be used.
- (c) Procedures to demonstrate incompatibility between the donor's cell type and the recipient's serum or plasma type. These procedures may consist of a serologic crossmatch, or a computer crossmatch. The computer crossmatch is a process of ensuring that a unit of blood is compatible with a specified recipient by means of electronically matching patient pretransfusion test results (ABO/Rh, etc.) with information about the blood donor that is stored in the LIS. The computer crossmatch is not strictly a "test" under CLIA; however, laboratories using this procedure must ensure that the LIS functions as intended. Refer to FDA Guidance for Industry: "Computer Crossmatch" (Computerized Analysis of the Compatibility between the Donor's Cell Type and the Recipient's Serum or Plasma Type). <<http://www.fda.gov/BiologicsBloodVaccines/GuidanceComplianceRegulatoryInformation/Guidances/Blood/ucm25829.htm>>.
- Laboratories using an immediate spin or computer crossmatch should have policies on the use of an antiglobulin crossmatch when warranted.

- (d) A provision that, if the unit of donor's blood has not been screened by a method that will demonstrate

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agglutinating, coating and hemolytic antibodies, the recipient's cells shall be tested with the donor's serum (minor crossmatch) by a method that will so demonstrate.

A minor crossmatch when the donor unit has not been screened for unexpected antibodies. Because all blood collected in FDA registered facilities is required to be screened for unexpected antibodies, this requirement is rarely applicable.

(e) Procedures to expedite transfusion in life-threatening emergencies. Records of all such incidents shall be maintained, including complete documentation justifying the emergency action, which shall be signed by a physician

The laboratory must maintain complete documentation, signed by a physician, which justifies the emergency action.

When condition-level deficiencies in Immunohematology are identified in any or all phases of testing, use D5026. Transfusion-related immunohematology testing performed on blood donors and recipients to determine compatibility is considered high complexity testing. When performed on blood donors or recipients, the following analytes are always high complexity: ABO group/ D (Rho) typing/antigen typing, direct antiglobulin tests, tests for unexpected antibody detection and identification, and crossmatch procedures. If personnel do not meet the qualifications or fulfill the responsibilities for high complexity testing, cite under subpart M-Personnel for Nonwaived Testing. There generally are no daily quality control requirements for reagent red cell panels used in antibody identification. However, the manufacturer's recommendations for QC are to be followed.

For laboratories using multiple racks of reagent typing sera and cells, laboratories should perform quality control on a representative sample of each lot of reagent in use on each day of testing. In addition, quality control needs to be performed on each new lot of reagent when first used.

When in-date reagents are unavailable, it may become necessary to frame written policies for their temporary use beyond their expiration dates until non-expired supplies become available. Under no circumstances, however, should a laboratory adopt policies that would allow for the regular use of expired reagents.

Determine if the laboratory has policies regarding:

- o Compatibility testing for patients with a history of a prior antibody;
- o Compatibility testing for patients with no history of a prior antibody; and

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- o Course of action to be taken for positive antibody screening and/or incompatible crossmatch.

Probes §493.1271(a)(1)

If the patient has been previously tested, how are results of current testing compared with interpretations of previous testing? When the results of current testing are discrepant with results of previous testing, how has the laboratory resolved the difference? Use D5777o

Interpretative Guidelines §493.1271(a)(2)

Determine if the laboratory has a policy to detect and resolve ABO discrepancies. If the laboratory does not have such procedures, use D5401. If the laboratory does not use patient records to confirm ABO group (i.e., current testing compared with historical records when available), use D5777.

Interpretative Guidelines §493.1271(a)(3)

Determine if the laboratory has established a policy specifying when testing for weak D must be performed.

Probes §493.1271(a)(3)

Is the laboratory following this policy?

Interpretative Guidelines §493.1271(f)

All non-transfusion related immunohematology QC records must be retained for at least 2 years. Use D3035.

Transfusion-related immunohematology QC records, including but not limited to, donor processing, compatibility testing, and transfusion reaction investigations, must be retained for the timeframe stated at 21 CFR §606.160(d) <<http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=606.160>>.

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**FED - D5553 - IMMUNOHEMATOLOGY**

**Title** IMMUNOHEMATOLOGY

**CFR** 493.1271(b)(f)

**Type** Standard

**Regulation Definition**

(b) Immunohematological testing and distribution of blood and blood products. Blood and blood product testing and distribution must comply with 21 CFR 606.100(b)(12); 606.160(b)(3)(ii) and (b)(3)(v); 610.40; 640.5(a), (b), (c), and (e); and 640.11(b).  
(f) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1271(b)

Refer to the current version of 21 CFR Parts 600 through 799

<<http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=606.160>> for the specified sections:

- o §606.100(b)(12) - Criteria for determining whether returned blood is suitable for reissue;
- o §606.160(b)(3)(ii) - Visual inspection of whole blood and red blood cells during storage and immediately before distribution;
- o §606.160(b)(3)(v) - Emergency release of blood, including signature of requesting physician obtained before or after release;
- o §610.40 Testing for communicable diseases;
- o §640.5(a) Syphilis testing;
- o §640.5(b) Determination of Blood group;
- o §640.5(c) Determination of Rh factor;
- o §640.5(e) Inspection of whole blood during storage and immediately prior to issue; and
- o §640.11(b) Inspection of RBC during storage and at the time of issue.

Probes §493.1271

If equipment and reagents are used in mobile or temporary testing sites, how are they protected from extreme

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temperature fluctuations when not in use (e.g., evenings, weekends, and holidays)?

Interpretative Guidelines §493.1271(f)

All non-transfusion related immunohematology QC records must be retained for at least 2 years. Use D3035.

Transfusion-related immunohematology QC records, including but not limited to, donor processing, compatibility testing, and transfusion reaction investigations, must be retained for the timeframe stated at 21 CFR §606.160(d) <<http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=606.160>>.

**FED - D5555 - IMMUNOHEMATOLOGY**

**Title** IMMUNOHEMATOLOGY

**CFR** 493.1271(c)(f)

**Type** Standard

**Regulation Definition**

(c) Blood and blood products storage. Blood and Blood products must be stored under appropriate conditions that include an adequate temperature alarm system that is regularly inspected.

(c)(1) An audible alarm system must monitor proper blood and blood product storage temperature over a 24-hour period.

(c)(2) Inspections of the alarm system must be documented.

(f) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1271(c)

Blood shall be stored in a clean and orderly environment in a manner to prevent mix-ups. No expired blood should be in the routine inventory. Unacceptable units should be segregated from routine inventory.

Acceptable temperature ranges must be established and actual readings of temperature-controlled storage areas must be recorded during the time that blood or blood products for transfusion are stored. Whole Blood, Red Blood Cells, and Liquid Plasma should be stored between 1 and 6° C; room temperature Platelets and Platelet Rich Plasma between 20 and 24° C or 1 and 6° C as indicated on the product label. Fresh Frozen Plasma, Plasma, and Cryoprecipitated AHF should be stored at -18° C or colder. Temperatures continuously monitored by a recording thermograph or central monitoring system are acceptable. The charts or central monitoring system must be retained to document that temperatures are maintained within acceptable limits as stated on the blood component label.

Verify that the laboratory regularly inspects the alarm system(s) according to its established policy. When the facility performs alarm checks, the temperature at which the alarm sounds should be compared to the temperature on the recording chart. Verify that the alarm activates at the appropriate temperature(s).

Reissue requirements are as follows: The container must have a tamper-proof seal which remains unbroken; records

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should indicate that the blood was maintained at 1 - 10° C while outside the control of the establishment; and the unit must be inspected prior to reissue. The laboratory must have a process for ensuring that blood components are maintained within acceptable limits while out of control of the laboratory.

Probes §493.1271(c)

Does the laboratory ensure that the freezer(s) used to store blood products is maintained at the recommended temperature(s) on a continuous basis?

Does the laboratory document and explain unacceptable storage temperatures? Use D5793.

What are the laboratory's criteria for determining blood or blood product suitability for reissue? Are they following their policy?

How are untested autologous units, potentially infectious units and reagents stored and segregated to prevent contamination?

If the laboratory does not have an emergency power source for the blood storage equipment and temperature alarm system, how does the laboratory ensure that blood is maintained at the appropriate temperature when a power failure occurs?

If the laboratory is not staffed 24 hours a day, seven days a week, how does it ensure prompt response to an activated alarm (evenings, weekends, and holidays)?

Interpretative Guidelines §493.1271(f)

All non-transfusion related immunohematology QC records must be retained for at least 2 years. Use D3035.

Transfusion-related immunohematology QC records, including but not limited to, donor processing, compatibility testing, and transfusion reaction investigations, must be retained for the timeframe stated at 21 CFR §606.160(d) <<http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=606.160>>.

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**FED - D5557 - IMMUNOHEMATOLOGY**

**Title** IMMUNOHEMATOLOGY

**CFR** 493.1271(d)(f)

**Type** Standard

**Regulation Definition**

(d) Retention of samples of transfused blood. According to the laboratory's established procedures, samples of each unit of transfused blood must be retained for further testing in the event of transfusion reactions. The laboratory must promptly dispose of blood not retained for further testing that has passed its expiration date.

(f) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1271(d)

There is no specific timeframe for retaining donor and recipient blood samples. However, it is common practice to keep these samples for a minimum of seven days after each transfusion in case there is a need for retesting.

Interpretative Guidelines §493.1271(f)

All non-transfusion related immunohematology QC records must be retained for at least 2 years. Use D3035.

Transfusion-related immunohematology QC records, including but not limited to, donor processing, compatibility testing, and transfusion reaction investigations, must be retained for the timeframe stated at 21 CFR §606.160(d) <<http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=606.160>>.

**FED - D5559 - IMMUNOHEMATOLOGY**

**Title** IMMUNOHEMATOLOGY

**CFR** 493.1271(e)(f)

**Type** Standard

**Regulation Definition**

(e) Investigation of transfusion reactions.

(e)(1) According to its established procedures, the laboratory that performs compatibility testing, or issues blood or blood products, must promptly investigate all transfusion reactions occurring in facilities for which it has investigational

**Interpretive Guideline**

Interpretative Guidelines §493.1271(e)(2):

Examine records of transfusion reaction investigations for completeness, accuracy, and promptness. Verify that investigations of transfusion reactions are conducted in accordance with the facility's established protocols. Records must include each step of the investigation, including conclusions and any follow-up.

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responsibility and make recommendations to the medical staff regarding improvements in transfusion procedures.

(e)(2) The laboratory must document, as applicable, that all necessary remedial actions are taken to prevent recurrences of transfusion reactions and that all policies and procedures are reviewed to assure they are adequate to ensure the safety of individuals being transfused.

(f) Documentation. The laboratory must document all control procedures performed, as specified in this section.

Probes §493.1271(e)(2):

If problems or technical errors are identified during a transfusion reaction investigation, are corrective actions taken and, as applicable, procedures instituted to prevent a recurrence?

Did the laboratory assess the adequacy of the procedures implemented? Use D5793

Interpretative Guidelines §493.1271(f)

All non-transfusion related immunohematology QC records must be retained for at least 2 years. Use D3035.

Transfusion-related immunohematology QC records, including but not limited to, donor processing, compatibility testing, and transfusion reaction investigations, must be retained for the timeframe stated at 21 CFR §606.160(d) <<http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=606.160>>.

**FED - D5601 - HISTOPATHOLOGY**

**Title** HISTOPATHOLOGY

**CFR** 493.1273(a)(f)

**Type** Standard

**Regulation Definition**

(a) As specified in §493.1256(e)(3), fluorescent and immunohistochemical stains must be checked for positive and negative reactivity each time of use. For all other differential or special stains, a control slide of known reactivity must be stained with each patient slide or group of patient slides. Reactions of the control slide with each special stain must be documented.

(f) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1273(a)

When condition-level deficiencies in Histopathology are identified in any or all phases of testing, use D5028.

The technical component, preparation of slides (TC) can be prepared in one laboratory and the finished product sent to another laboratory for professional interpretation (PC). Both laboratories should show documentation of adequate slide preparation which is processing, and processing includes the QC of the stain at both locations.

The laboratory must demonstrate that each reagent performs within the specifications established by the laboratory for the test procedure. Documentation of concurrent testing of reagents or acceptable quality control results will satisfy this requirement.

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When the laboratory uses a manufacturer's kit, the reagents of the kit must not be combined, mixed, or replaced with components of another kit from a different lot number, unless otherwise permitted and specified by the manufacturer in the package insert. Use D5419.

Laboratories which use automated staining methodologies must follow the manufacturer's instructions. Use D5411.

Flow Cytometry

Staining controls for cell surface immunophenotyping by flow cytometry should consist of either normal, cultured or abnormal cells known to be positive for selected standard antigens and must verify the proper performance of reagents. Frozen or other preserved cells may be used. A negative reagent control must be run for each test cell preparation, and is to consist of monoclonal antibody(ies) of the same species and isotype or equivalent. Negative reagent controls will consist of:

- (a) For indirect stains, an irrelevant primary antibody and the same secondary antibody(ies) conjugated with the same fluorochrome(s) used in all relevant test combinations; and
- (b) For direct stains, an irrelevant antibody conjugated to the same fluorochrome and at the same fluorochromes: protein ratio used in all relevant test combinations.

Probes §493.1273(a)

For flow cell cytometric surface immunophenotyping, is a negative reagent control used to define a threshold for positive staining cells? If not, how does the laboratory define the threshold for positive staining cells?

Is a quality control slide with the appropriate differential or special stain tested at the same time patient specimens are tested?

Interpretative Guidelines §493.1273(f)

QC records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reactions, and/or observations and demonstrate that controls were tested when shipments of reagents, stains, or kits were opened or when the laboratory prepared these materials.

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**FED - D5603 - HISTOPATHOLOGY**

**Title** HISTOPATHOLOGY

**CFR** 493.1273(b)(f)

**Type** Standard

**Regulation Definition**

(b) The laboratory must retain stained slides, specimen blocks, and tissue remnants as specified in §493.1105. The remnants of tissue specimens must be maintained in a manner that ensures proper preservation of the tissue specimens until the portions submitted for microscopic examination have been examined and a diagnosis made by an individual qualified under §§493.1449(b), (l), or (m).

(f) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1273(f)

QC records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reactions, and/or observations and demonstrate that controls were tested when shipments of reagents, stains, or kits were opened or when the laboratory prepared these materials.

**FED - D5605 - HISTOPATHOLOGY**

**Title** HISTOPATHOLOGY

**CFR** 493.1273(c)(f)

**Type** Standard

**Regulation Definition**

(c) An individual who has successfully completed a training program in neuromuscular pathology approved by HHS may examine and provide reports for neuromuscular pathology.

(f) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1273(c)

HHS approves the American Academy of Neurology Committee for Neuromuscular Pathology Training Program.

Interpretative Guidelines §493.1273(f)

QC records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reactions, and/or observations and demonstrate that controls were tested when shipments of reagents, stains, or kits were opened

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or when the laboratory prepared these materials.

**FED - D5607 - HISTOPATHOLOGY**

**Title** HISTOPATHOLOGY

**CFR** 493.1273(d)(f)

**Type** Standard

**Regulation Definition**

(d) Tissue pathology reports must be signed by an individual qualified as specified in paragraph (b) or, as appropriate, paragraph (c) of this section. If a computer report is generated with an electronic signature, it must be authorized by the individual who performed the examination and made the diagnosis.

(f) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1273(d)

The laboratory must ensure that only those individuals qualified to evaluate histopathology specimens can release his or her electronic signature for reporting purposes.

The tests in histopathology include both gross examination (macroscopic) and microscopic examination of the slide(s) with evaluation and diagnostic interpretation, and diagnostic findings reported.

In the event of a computer-generated signature, the laboratory must ensure that the system is protected from use by unauthorized individuals.

If the technical supervisor who performed the examination and diagnosis is not available to sign the report, an individual, also qualified as a technical supervisor in Histopathology, must reexamine and diagnose in order to sign out the report.

Interpretative Guidelines §493.1273(f)

QC records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reactions, and/or observations and demonstrate that controls were tested when shipments of reagents, stains, or kits were opened or when the laboratory prepared these materials.

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**FED - D5609 - HISTOPATHOLOGY**

**Title** HISTOPATHOLOGY

**CFR** 493.1273(e)(f)

**Type** Standard

**Regulation Definition**

(e) The laboratory must use acceptable terminology of a recognized system of disease nomenclature in reporting results.

(f) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1273(e)

"SNOMED (Registered)" - Systemized Nomenclature of Medicine is an example of a recognized system of disease nomenclature.

Interpretative Guidelines §493.1273(f)

QC records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reactions, and/or observations and demonstrate that controls were tested when shipments of reagents, stains, or kits were opened or when the laboratory prepared these materials.

**FED - D5613 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(a)

**Type** Standard

**Regulation Definition**

Cytology slide examination site. All cytology slide preparations must be evaluated on the premises of a laboratory certified to conduct testing in the subspecialty of cytology.

**Interpretive Guideline**

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**FED - D5615 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(b)(1)

**Type** Standard

**Regulation Definition**

(b) Staining. The laboratory must have available and follow written policies and procedures for each of the following, if applicable:

(b)(1) All gynecologic slide slide preparations must be stained using a Papanicolaou or modified Papanicolaou staining method.

**Interpretive Guideline**

Interpretative Guidelines §493.1274(b)(1)

The Papanicolaou staining procedure is a polychrome method that enhances differences in cellular morphology. The procedure utilizes a nuclear stain, hematoxylin and two cytoplasmic counterstains, OG-6 and EA. The Papanicolaou method is used for staining cytologic preparations because it provides well-defined nuclear detail, stains cytoplasm of various cell types different colors, and renders transparent cytoplasm. There are a variety of formulas for making hematoxylin, OG-6, and EA stains. The actual staining technique may vary among laboratories depending on the type of stains used and the laboratories' modification of the staining method. Modifications of the staining procedure must include the four main steps of the standard Papanicolaou staining method: fixation, nuclear staining, cytoplasmic staining, and clearing.

Cytology laboratories may receive reagents, solutions, and stains from a manufacturer in large volume stock containers. For ease in handling, portions of these reagents are usually decanted into smaller working containers, which must be labeled in accordance with §493.1252(c). Some manufacturers do not label stain or reagent containers with the expiration date; however, lot numbers and package inserts refer to this information.

If the laboratory uses a manufacturer's kit, the reagents of the kit must not be combined, mixed, or replaced with components of another kit from a different lot number, unless otherwise permitted and specified by the manufacturer in the package insert (use D5419). Laboratories which use automated staining methodologies must follow the manufacturer's instructions (use D5411).

The cytology laboratory must document the expiration date of stock reagents, working stains, and solutions made in the laboratory. Use D5415.

Laboratories may use staining procedures, other than the Papanicolaou method, for staining nongynecologic specimens.

Review the written staining procedure for staining gynecologic specimens. Confirm that the written procedures

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reflect:

- o Stains used (i.e., Harris, Gill or other type of hematoxylin, OG-6, modified OG-6, EA36, EA50, EA65, modified EA) or the identity of a combination counterstain;
- o Solutions used (water, alcohol, clearing reagent, acid and bluing agent);
- o Concentration of each solution used (i.e., percentage (%) of alcohol, acid, ammonium hydroxide or lithium carbonate solution);
- o Length of time or number of dips slides are placed in each stain or solution;
- o The staining dishes must be labeled to reflect content (not just lids); and
- o Procedure for coverslipping slides.

Current time frames must be specified in the procedure manual for each step in the staining of cytology specimens using the Papanicolaou staining method. Adjustments to time frame changes must be documented.

Step-by-step written procedures must be available and followed to prepare nongynecologic specimens.

Use D5403 if any of the above findings is not met.

The laboratory must ensure that the gynecologic and non-gynecologic stains have been tested to ensure predictable staining characteristics on a daily basis. Use D5473.

NOTE: Any fixatives, reagents, or preservatives intended to be used on one liquid-based manufacturer's instrument must not be used on another manufacturer's instrument.

**FED - D5617 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(b)(2)

**Type** Standard

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**Regulation Definition**

(b) Staining. The laboratory must have available and follow written policies and procedures for each of the following, if applicable:

(b)(2) Effective measures to prevent cross-contamination between gynecologic and nongynecologic specimens during the staining process must be used.

**Interpretive Guideline**

Interpretative Guidelines §493.1274(b)(2)

The laboratory must develop its own policies and procedures for the prevention of cross-contamination between gynecologic and nongynecologic specimens. The majority of gynecologic specimens are fixed prior to transport to the laboratory. Staining times may differ between gynecologic and nongynecologic specimens. Commonly used methods include separate staining dishes for various specimens (i.e., gynecologic specimens, CSF, sputa, other body fluids), or separate staining times (i.e., gynecologic specimens in the morning and nongynecologic specimens in the afternoon), with the staining dishes washed and stains filtered between staining times.

Probes §493.1274(b)(2)

What does the laboratory do to ensure that cross-contamination between gynecologic and nongynecologic specimens does not occur?

**FED - D5619 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(b)(3)

**Type** Standard

**Regulation Definition**

(b) Staining. The laboratory must have available and follow written policies and procedures for each of the following, if applicable:

(b)(3) Nongynecologic specimens that have a high potential for cross-contamination must be stained separately from other nongynecologic specimens, and the stains must be filtered or changed following staining.

**Interpretive Guideline**

Interpretative Guidelines §493.1274(b)(3)

A monochromatic stain such as toluidine blue may be used to determine the cellularity of nongynecologic specimens. Once a specimen has been concentrated, usually by centrifugation, a small drop of specimen is placed on a slide. A drop of stain is placed next to the specimen, allowed to mix, and coverslipped. Cellularity is evaluated microscopically. Highly cellular specimens have a high potential for cross-contamination. One option would be for the laboratory to stain these specimens after routine staining has been completed.

Laboratories which use automated staining methodologies must follow the manufacturer's instructions. Use D5411.

Probes §493.1274(b)(3)

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How is the cellularity of nongynecologic specimens checked prior to cytopreparation (staining)?

What procedure does the laboratory use to determine which specimens must be stained separately?

**FED - D5621 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(c)(1)

**Type** Standard

**Regulation Definition**

(c) Control procedures. The laboratory must establish and follow written policies and procedures for a program designed to detect errors in the performance of cytologic examinations and the reporting of results. The program must include the following:

(c)(1) A review of slides from at least 10 percent of the gynecologic cases interpreted by individuals qualified under §§493.1469 or 493.1483, to be negative for epithelial cell abnormalities and other malignant neoplasms (as defined in paragraph (e)(1) of this section).

(c)(1)(i) The review must be performed by an individual who meets one of the following qualifications:

(c)(1)(i)(A) A technical supervisor qualified under §§493.1449(b) or (k).

(c)(1)(i)(B) A cytology general supervisor qualified under §493.1469.

(c)(1)(i)(C) A cytotechnologist qualified under §493.1483 who has the experience specified in §493.1469(b)(2).

(c)(1)(ii) Cases must be randomly selected from the total caseload and include negatives and those from patients or groups of patients that are identified as having a higher than average probability of developing cervical cancer based on available patient information.

(c)(1)(iii) The review of those cases selected must be completed before reporting patient results.

**Interpretive Guideline**

Interpretative Guidelines §493.1274(c)(1)

The 10 percent rescreen of negative cases is not required for a one-person laboratory consisting of a technical supervisor or a laboratory which only employs pathologists qualified as technical supervisors. However, these laboratories must establish and follow a program to detect errors. This program must include, but is not limited to, cytologic/histologic correlations, retrospective review of negative cases, documentation of initial and rescreening results, and statistics [(c)(2)-(5) of this section].

The laboratory must review all slides from each case selected for rescreen.

Interpretative Guidelines §493.1274(c)(1)(i)

The laboratory must document which individual(s) are qualified to conduct the 10 percent rescreen. Slides reviewed as part of the 10 percent rescreen must be included in the workload limit of the cytology general supervisor or the cytotechnologist performing the review. Use D5639.

Interpretative Guidelines §493.1274(c)(1)(ii)

The laboratory must have a procedure to determine which slides are rescreened. This procedure should ensure that individuals screening the slides do not know which slides will be chosen for rescreen.

The laboratory must establish criteria to ensure that random negative gynecological cases selected for rescreening include, when possible, cases from patients that are identified as having a higher than average probability for developing cervical cancer.

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**FED - D5623 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(c)(2)

**Type** Standard

**Regulation Definition**

(c) Control procedures. The laboratory must establish and follow written policies and procedures for a program designed to detect errors in the performance of cytologic examinations and the reporting of results. The program must include the following:

(c)(2) Laboratory comparison of clinical information, when available, with cytology reports and comparison of all gynecologic cytology reports with a diagnosis of high-grade squamous intraepithelial lesion (HSIL), adenocarcinoma, or other malignant neoplasms with the histopathology report, if available in the laboratory (either on-site or in storage), and determination of the causes of any discrepancies.

**Interpretive Guideline**

Interpretative Guidelines §493.1274(c)(2)

The laboratory must compare clinical information with cytology final reports. For example, an atrophic smear (usually characteristic of a post-menopausal woman) from a 21-year-old female with an LMP (last menstrual period) of 2-weeks-ago constitutes inconsistent findings and must be resolved.

The laboratory must define criteria to determine a discrepancy between a final cytological diagnosis of High Grade Squamous Intraepithelial Lesion (HSIL) or squamous carcinoma, adenocarcinoma or other malignant neoplasias and the correlating histology report.

Cases considered HSIL include: moderate and severe dysplasia, carcinoma in-situ (CIS)/Cervical Intraepithelial Neoplasia (CIN) 2 and CIN 3 or with features suspicious for invasion.

Probes §493.1274(c)(2)

How does the laboratory identify and resolve discrepancies for:

- o Clinical information vs. cytology report; and
- o Gynecologic cytology report vs. histopathology report?

**FED - D5625 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(c)(3)

**Type** Standard

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**Regulation Definition**

(c) Control procedures. The laboratory must establish and follow written policies and procedures for a program designed to detect errors in the performance of cytologic examinations and the reporting of results. The program must include the following:

(c)(3) For each patient with a current HSIL, adenocarcinoma, or other malignant neoplasm, laboratory review of all normal or negative gynecologic specimens received within the previous 5 years, if available in the laboratory (either on-site or in storage). If significant discrepancies are found that will affect current patient care, the laboratory must notify the patient's physician and issue an amended report.

**Interpretive Guideline**

Probes §493.1274(c)(3)

How does the laboratory track previous cases on an individual patient?

What criteria does the laboratory use to determine discrepancies when reviewing normal or negative slides from the past five years? How does the laboratory document the review?

How does the laboratory use the retrospective review to assess the analytic system and communicate findings to the appropriate staff? Use D5793.

**FED - D5627 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(c)(4)

**Type** Standard

**Regulation Definition**

Records of initial examinations and all rescreening results must be documented.

**Interpretive Guideline**

**FED - D5629 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(c)(5)

**Type** Standard

**Regulation Definition**

(c) Control procedures. The laboratory must establish and follow written policies and procedures for a program designed

**Interpretive Guideline**

Interpretative Guidelines §493.1274(c)(5)(vi)

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to detect errors in the performance of cytologic examinations and the reporting of results. The program must include the following:

(c)(5) An annual statistical laboratory evaluation of the number of -

(c)(5)(i) Cytology cases examined;

(c)(5)(ii) Specimens processed by specimen type;

(c)(5)(iii) Patient cases reported by diagnosis (including the number reported as unsatisfactory for diagnostic interpretation);

(c)(5)(iv) Gynecologic cases with a diagnosis of HSIL, adenocarcinoma, or other malignant neoplasm for which histology results were available for comparison;

(c)(5)(v) Gynecologic cases where cytology and histology are discrepant; and

(c)(5)(vi) Gynecologic cases where any rescreen of a normal or negative specimen results in reclassification as low-grade squamous intraepithelial lesion (LSIL), HSIL, adenocarcinoma, or other malignant neoplasms.

Low-grade Squamous Intraepithelial Lesions (LSIL) encompasses all lesions that demonstrate cellular changes consistent with human papillomavirus, mild dysplasia, or CIN 1.

**FED - D5631 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(c)(6)

**Type** Standard

**Regulation Definition**

(c) Control procedures. The laboratory must establish and follow written policies and procedures for a program designed to detect errors in the performance of cytologic examinations and the reporting of results. The program must include the following:

(c)(6) An evaluation of the case reviews of each individual examining slides against the laboratory's overall statistical values, documentation of any discrepancies, including reasons for the deviation, and, if appropriate, corrective actions taken.

**Interpretive Guideline**

Probes §493.1274(c)(6)

How does the laboratory evaluate each individual's case reviews against the overall laboratory statistics?

What corrective actions are taken to resolve discrepancies?

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**FED - D5633 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(d)(1)

**Type** Standard

**Regulation Definition**

(d) Workload limits. The laboratory must establish and follow written policies and procedures that ensure the following:  
(d)(1) The technical supervisor establishes a maximum workload limit for each individual who performs primary screening.

**Interpretive Guideline**

Interpretative Guidelines §493.1274(d)(1)

The maximum workload limit established by the technical supervisor must be based on each individual's capabilities. A generic workload limit for the laboratory as a whole does not meet this requirement.

Probes §493.1274(d)(1)

What criteria does the technical supervisor use to determine the slide limit for each person who examines slides?

**FED - D5635 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(d)(1)(i)

**Type** Standard

**Regulation Definition**

(d) Workload limits. The laboratory must establish and follow written policies and procedures that ensure the following:  
(d)(1)(i) The workload limit is based on the individual's performance using evaluations of the following:  
(d)(1)(i)(A) Review of 10 percent of the cases interpreted as negative for the conditions defined in paragraph (e)(1) of this section.  
(d)(1)(i)(B) Comparison of the individual's interpretation with the technical supervisor's confirmation of patient smears specified in paragraphs (e)(1) and (e)(3) of this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1274(d)(1)(i)

The technical supervisor maintains documentation of the slide performance and provides feedback.

Probes §493.1274(d)(1)(i)

What records are maintained to document the technical supervisor's evaluation of the slide performance of each individual?

Probes §493.1274(d)(1)(i)(B)

How does the technical supervisor ensure that feedback is provided on slide examination performance to each person evaluating slides?

What mechanism is used to allow individuals an opportunity to discuss instances of misdiagnosis?

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**FED - D5637 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(d)(1)(ii)

**Type** Standard

**Regulation Definition**

(d) Workload limits. The laboratory must establish and follow written policies and procedures that ensure the following:  
(d)(1)(ii) Each individual's workload limit is reassessed at least every 6 months and adjusted when necessary.

**Interpretive Guideline**

Probes §493.1274(d)(1)(ii)

What criteria does the technical supervisor use to determine when a workload adjustment is needed?

How are records maintained to document that workload records are reassessed at least every six months and adjusted when necessary?

**FED - D5639 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(d)(2)(i)

**Type** Standard

**Regulation Definition**

(d) Workload limits. The laboratory must establish and follow written policies and procedures that ensure the Following:  
(d)(2) The maximum number of slides examined by an individual in each 24-hour period does not exceed 100 slides (one patient specimen per slide; gynecologic, nongynecologic, or both) irrespective of the site or laboratory. This limit represents an absolute maximum number of slides and must not be employed as an individual's performance target. In addition--  
(d)(2)(i) The maximum number of 100 slides is examined in no less than an 8-hour workday;

**Interpretive Guideline**

Interpretative Guidelines §493.1274(d)(2)

The maximum total number of slides an individual may screen is 100 per 24 hours regardless of site or laboratory. Although the regulation establishes this maximum number, not every individual will be able to accurately examine 100 slides in 24 hours. The laboratory must establish how many slides can be screened per day for each individual. Refer to §493.1274(d)(1) to ensure that the technical supervisor has established a maximum number of slides that each individual is capable of evaluating. The laboratory must ensure that persons employed at other sites or locations do not exceed the maximum of 100 slides in 24 hours.

This 100-slide limit is also applicable to those technical supervisors who examine previously unevaluated cytology specimens.

Probes §493.1274(d)(2)

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How does the laboratory ensure that each individual examining slides (cytotechnologists, cytology general supervisors and technical supervisors in cytology, as applicable) examines no more than 100 slides in a 24-hour period regardless of site or location?

Probes §493.1274(d)(2)(i)

What records are used to verify that the maximum number of 100 slides is examined in no less than 8 hours, especially in the situation in which individuals screen slides at different sites or locations?

**FED - D5641 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(d)(2)(ii)

**Type** Standard

**Regulation Definition**

(d) Workload limits. The laboratory must establish and follow written policies and procedures that ensure the following:

(d)(2)(ii) For the purposes of establishing workload limits for individuals examining slides in less than an 8-hour workday (includes full-time employees with duties other than slide examination and part-time employees), a period of 8 hours is used to prorate the number of slides that may be examined.

The formula--

Number of hours examining slides X 100 / 8

is used to determine maximum slide volume to be examined;

**Interpretive Guideline**

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**FED - D5643 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(d)(2)(iii)

**Type** Standard

**Regulation Definition**

(d) Workload limits. The laboratory must establish and follow written policies and procedures that ensure the following:  
(d)(2)(iii) Nongynecologic slide preparations made using liquid-based slide preparatory techniques that result in cell dispersion over one-half or less of the total available slide may be counted as one-half slide; and  
(d)(2)(iv) Technical supervisors who perform primary screening are not required to include tissue pathology slides and previously examined cytology slides (gynecologic and nongynecologic) in the 100 slide workload limit.

**Interpretive Guideline**

Interpretative Guidelines §493.1274(d)(2)(iii)  
Nongynecologic slide preparations made using automated, semi-automated or other liquid-based slide preparatory techniques include specimens prepared by centrifugation, cyto centrifugation, filtering techniques or monolayering techniques. Any instrument used to assist in the adherence of cells to the slide is considered to meet this requirement. This requirement refers to slide preparatory techniques, not liquid based coverslips. Slides prepared by traditional methods (usually smears prepared by hand) are not included.

Maximum Workload Limits for Nongynecologic Specimens:

Traditional Smear Technique 100 Slides  
Automated, Semi-Automated, Liquid-Based 200 Slides  
Combination of Techniques 100 - 200 Slides  
(Based on Prorated Time)

**FED - D5645 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(d)(3)

**Type** Standard

**Regulation Definition**

(d) Workload limits. The laboratory must establish and follow written policies and procedures that ensure the following:  
(d)(3) The laboratory must maintain records of the total number of slides examined by each individual during each 24-hour period and the number of hours spent examining

**Interpretive Guideline**

Interpretative Guidelines §493.1274(d)(3)  
Verify that the laboratory monitors the number of slides examined by each individual and the number of hours spent examining slides.

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slides in the 24-hour period irrespective of the site or laboratory.

**FED - D5647 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(d)(4)

**Type** Standard

**Regulation Definition**

(d) Workload limits. The laboratory must establish and follow written policies and procedures that ensure the following:  
(d)(4) Records are available to document the workload limit for each individual.

**Interpretive Guideline**

Probes §493.1274(d)(4)  
What records are maintained of each individual's workload limit when various types of slides are evaluated?

**FED - D5649 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(e)(1)

**Type** Standard

**Regulation Definition**

(e) Slide examination and reporting. The laboratory must establish and follow written policies and procedures that ensure the following:  
(e)(1) A technical supervisor confirms each gynecologic slide preparation interpreted to exhibit reactive or reparative changes or any of the following epithelial cell abnormalities:  
(e)(1)(i) Squamous cell.  
(e)(1)(i)(A) Atypical squamous cells of undetermined significance (ASC-US) or cannot exclude HSIL (ASC-H).  
(e)(1)(i)(B) LSIL-Human papillomavirus (HPV)/mild dysplasia/cervical intraepithelial neoplasia 1 (CIN 1).  
(e)(1)(i)(C) HSIL-moderate and severe dysplasia, carcinoma

**Interpretive Guideline**

Interpretative Guidelines §493.1274(e)(1)(i)  
Note: This requirement is in addition to the review and confirmation by a technical supervisor of all nongynecologic preparations as described under §493.1274(e)(3).

Probes §493.1274(e)(1)(i)  
How does the laboratory ensure that the technical supervisor confirms every slide containing cells exhibiting reactive, reparative, atypical squamous/glandular cells, LSIL, HSIL, and all carcinomas?

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in situ (CIS)/CIN 2 and CIN 3 or with features suspicious for invasion.

(e)(1)(i)(D) Squamous cell carcinoma.

(e)(1)(ii) Glandular Cell

(e)(1)(ii)(A) Atypical cells not otherwise specified (NOS) or specified in comments (endocervical, endometrial, or glandular).

(e)(1)(ii)(B) Atypical cells favor neoplastic (endocervical or glandular).

(e)(1)(ii)(C) Endocervical adenocarcinoma in situ.

(e)(1)(ii)(D) Adenocarcinoma endocervical, adenocarcinoma endometrial, adenocarcinoma extrauterine, and adenocarcinoma NOS.

(e)(1)(iii) Other malignant neoplasms.

**FED - D5651 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(e)(2)

**Type** Standard

**Regulation Definition**

(e) Slide examination and reporting. The laboratory must establish and follow written policies and procedures that ensure the following:

(e)(2) The report of gynecologic slide preparations with conditions specified in paragraph (e)(1) of this section must be signed to reflect the technical supervisory review or, if a computer report is generated with signature, it must reflect an electronic signature authorized by the technical supervisor who performed the review.

**Interpretive Guideline**

Interpretative Guidelines §493.1274(e)(2)

The laboratory must ensure that the technical supervisor is the only individual to release his or her electronic signature for reports requiring technical supervisory review.

If an electronic signature is used, the laboratory must ensure that the system is protected from use by unauthorized individuals.

If the technical supervisor who performed the examination and diagnosis is not available to sign the report, an individual, also qualified as a technical supervisor in Cytology, must reexamine and confirm the findings prior to signing the report.

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**FED - D5653 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(e)(3)

**Type** Standard

**Regulation Definition**

(e) Slide examination and reporting. The laboratory must establish and follow written policies and procedures that ensure the following:

(e)(3) All nongynecologic preparations are reviewed by a technical supervisor. The report must be signed to reflect technical supervisory review or, if a computer report is generated with signature, it must reflect an electronic signature authorized by the technical supervisor who performed the review.

**Interpretive Guideline**

Interpretative Guidelines §493.1274(e)(3)

The laboratory must ensure that the technical supervisor:

- o Is the only individual to release his or her electronic signature for reports requiring technical supervisory review; and
- o Reviews all nongynecologic cytological preparations.

If an electronic signature is used, the laboratory must ensure that the system is protected from use by unauthorized individuals.

If the technical supervisor who performed the examination and diagnosis is not available to sign the report, an individual, also qualified as a technical supervisor in Cytology, must reexamine and confirm the findings prior to signing the report.

**FED - D5655 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(e)(4)

**Type** Standard

**Regulation Definition**

(e) Slide examination and reporting. The laboratory must establish and follow written policies and procedures that ensure the following:

(e)(4) Unsatisfactory specimens or slide preparations are identified and reported as unsatisfactory.

**Interpretive Guideline**

Interpretative Guidelines §493.1274(e)(4)

The report should clearly specify when the slide is unsatisfactory for evaluation. Unsatisfactory slide preparations should not be reported as negative or normal. Use D5805.

Probes §493.1274(e)(4)

What criteria have been developed for categorizing a slide preparation as unsatisfactory (e.g., scant cellularity,

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obscuring blood, obscuring inflammation, or lack of endocervical component)?

**FED - D5657 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(e)(5)

**Type** Standard

**Regulation Definition**

(e) The laboratory must establish and follow written policies and procedures that ensure the following:

(e)(5) The report contains narrative descriptive nomenclature for all results.

**Interpretive Guideline**

Interpretative Guidelines §493.1274(e)(5)

In cytology, great variation exists among the systems and terms a laboratory may use to report patient results on cytology reports. The laboratory must specify the descriptive nomenclature used for reporting patient results. This nomenclature must define the criteria used to classify patient results in a particular category in a clear and concise manner to ensure that all employees report patient results in a uniform, consistent manner. Use of the Papanicolaou numerical system without narrative description is not acceptable.

The Bethesda System is an example of a recognized system of narrative descriptive nomenclature for gynecologic cytology.

Probes §493.1274(e)(5)

When cytology evaluations are recorded on worksheets in "code" how does the laboratory ensure that the correct interpretation is used in reporting the results? Use D5801.

**FED - D5659 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(e)(6)

**Type** Standard

**Regulation Definition**

(e) The laboratory must establish and follow written policies and procedures that ensure the following:

(e)(6) Corrected reports issued by the laboratory indicate the basis for correction.

**Interpretive Guideline**

Interpretative Guidelines §493.1274(e)(6)

Corrected reports, either hard copy or electronic, must clearly indicate both the corrected results(s), and the fact that the report is a corrected report. The corrected reports should be promptly sent to the authorized person and to all

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known recipients of the original incorrect report.

Probes §493.1274(e)(6)

How does the laboratory indicate that the report is a corrected report (to avoid confusion with the initial report)? Use D5821.

How does the laboratory include the cause or reason for the correction in the report?

**FED - D5660 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274 (f)(1)

**Type** Standard

**Regulation Definition**

(f) Record and slide retention  
(f)(1) The laboratory must retain all records and slide preparations as specified in §493.1105.

**Interpretive Guideline**

**FED - D5661 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(f)(2)(3)

**Type** Standard

**Regulation Definition**

(f) Record and slide retention.  
(f)(2) Slides may be loaned to proficiency testing programs in lieu of maintaining them for the required time period, provided the laboratory receives written acknowledgment of the receipt of slides by the proficiency testing program and maintains the acknowledgment to document the loan of these slides. (f)(3) Documentation of slides loaned or referred for

**Interpretive Guideline**

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purposes other than proficiency testing must be maintained.

**FED - D5663 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(f)(4)

**Type** Standard

**Regulation Definition**

(f) Record and slide retention.  
(f)(4) All slides must be retrievable upon request.

**Interpretive Guideline**

Probes §493.1274(f)(4)  
If the laboratory loans slides, what protocol has been established to ensure prompt return of slides, when necessary?

**FED - D5665 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(g)

**Type** Standard

**Regulation Definition**

Automated and semi-automated screening devices. When performing evaluations using automated and semi-automated screening devices, the laboratory must follow manufacturer's instructions for preanalytic, analytic, and postanalytic phases of testing, as applicable, and meet the applicable requirements of this subpart K.

**Interpretive Guideline**

Interpretative Guidelines §493.1274(g)  
Some automated devices, such as instruments where only a portion of the slide is reviewed, may have a higher workload limit than 100 slides. This must be stated in the manufacturer's product insert to be applicable. However, the maximum workload limit for those slides which require 100% manual review (as a result of automated or semi-automated analysis OR in the routine workload) remains 100 slides.

Probes §493.1274(g)  
When technology (automated/semi-automated devices) is introduced into the laboratory, how does the laboratory ensure its operation is within the specifications of previous methods used by the laboratory?

Some automated devices remove a percentage of the slides from the workload. How does the laboratory ensure that the correct slides are archived?

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**FED - D5667 - CYTOLOGY**

**Title** CYTOLOGY

**CFR** 493.1274(h)

**Type** Standard

**Regulation Definition**

Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1274(h)

QC records should include lot numbers, date prepared/opened, expiration dates, the actual measurements, reactions, and/or observations and demonstrate that controls were tested when shipments of reagents, stains, or kits were opened or when the laboratory prepared these materials.

The actual measurements(s) taken, reactions and/or observations must be recorded. However, do not dictate the acceptable format for documentation.

The laboratory must maintain documentation to demonstrate that ten percent of the negative cases were rescreened.

All QC records must be maintained for two years, for example: five year retrospective review, 10 percent rescreens, cytology/histology correlations, cytotechnologist's performance evaluations, individual's and laboratory's statistics (use D3031). Use D3043 for retention of glass slides and D3041 for retention of patient test reports.

The laboratory must document the evaluation of quality control data and ensure that corrective actions are effective. Use D5793.

NOTE: Please refer to D2064 and D6116 for laboratories performing Human Papillomavirus (HPV) testing.

Probes §493.1274(h)

What information is documented on the quality control records?

What records does the laboratory maintain to document that stains are filtered or changed when necessary?

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**FED - D5681 - CLINICAL CYTOGENETICS**

**Title** CLINICAL CYTOGENETICS

**CFR** 493.1276(a)(e)

**Type** Standard

**Regulation Definition**

- (a) The laboratory must have policies and procedures for ensuring accurate and reliable patient specimen identification during the process of accessioning, cell preparation, photographing or other image reproduction technique, photographic printing, and reporting and storage of results, karyotypes, and photographs.
- (e) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1276(a)

When condition level deficiencies in Clinical Cytogenetics are in any or all phases of testing, use D5034.

Determine which of the following services may be provided:

- o Tissue Cultures (e.g., skin, lung, product of conception);
- o Bone Marrow Cultures;
- o Solid Tumors;
- o Lymph Nodes;
- o Chorionic Villus Samples (CVS);
- o Peripheral Lymphocyte Cultures;
- o Amniotic Fluid Cultures;
- o High resolution chromosome analysis;
- o Special techniques (e.g., Fragile "X" Studies, Chromosome Breakage analysis);
- o Karyotype analysis (photographic and/or computer methods);
- o Transplant studies;
- o Chromosome staining (banding techniques) such as:
  - Quinacrine fluorescence (Q Banding);
  - Giesma/trypsin (G Banding);
  - Sodium phosphate/acridine or giesma/heat (R Banding);
  - Barium hydroxide/heat (C Banding);
  - Nuclear Organizing Region - Silver Stain (NOR);
  - Distamycin A/4-6-diamidino-2-phenylindole (DA/DAPI); or
  - Giemsa 11 (pH 11.0 for heterochromatin) (G 11).

NOTE: The above listing is not intended to be all-inclusive.

Review a sample of patient case files to determine if it is possible to go from the accession number to the patient's file with karyotypes, report and observation records, the microscope slide, photographs or requisition forms.

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Probes §493.1276(a)

When photographs are taken, are the coordinates of the microscope noted for each cell selected? If not, how does the laboratory identify the cell for future reference?

What system does the laboratory use to ensure that records reflect accurate patient identification when:

- o Photographing chromosome spreads;
- o Using computer systems to assist in karyotyping; or
- o Storing photographic images of chromosomes and chromosomes spreads?

Probes §493.1276(e)

Each day of use, does the laboratory test the positive and negative reactivity of staining materials to ensure predictable staining characteristics? Use D5473.

Does the laboratory, concurrent with the initial use, check each batch of media for pH (amniotic cell cultures should be kept between pH 6.8 and 7.8), sterility, and ability to support growth? Use D5477.

Does the laboratory employ an alternative procedure for the immediate assessment and monitoring of all testing over time? For example: Control materials are not routinely available to demonstrate chromosome abnormalities for linkage, breakage or translocation, but the laboratory must demonstrate an alternative mechanism for detecting chromosome abnormalities to be analyzed. Use 5485.

An alternative procedure might include spit sample with another laboratory, repeat patient specimen, special stains, FISH assays, and/or molecular assays.

**FED - D5683 - CLINICAL CYTOGENETICS**

**Title** CLINICAL CYTOGENETICS

**CFR** 493.1276(b)(e)

**Type** Standard

**Regulation Definition**

(b) The laboratory must have records that document the following:

(b)(1) The media used, reactions observed, number of cells counted, number of cells karyotyped, number of chromosomes

**Interpretive Guideline**

Interpretative Guidelines §493.1276(b)(1)-(b)(3)

Culture Type	Minimum Number of Spreads Counted per	Minimum Number of Cells Analyzed per Patient
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counted for each metaphase spread, and the quality of the banding

(b)(2) The resolution is appropriate for the type of tissue or specimen and the type of study required based on the clinical information provided to the laboratory.

(b)(3) An adequate number of karyotypes are prepared for each patient.

(e) The laboratory must document all control procedures performed, as specified in this section.

	Patient		
	Amniotic Fluid		
	Flasks	15 cells from at least 2 independent primary cultures	5 cells from at least 2 independent primary cultures
	in situ	15 cells from at least 10 colonies from 2 independent primary cultures	5 cells from different colonies and split between different primary cultures

Many laboratories use a combination of the flask and in situ culture methods or use the flask method as a backup for the in situ method.

	Chorionic Villus		
	Direct	15 cells	5 cells
	Culture	as in amniotic fluid, flask technique	

	Peripheral Blood		
	Constitutional	20 cells	5 cells
	Possible sex chromosome abnormality	30 cells (total count)	5 cells

Culture Type	Minimum Number of Spreads Analyzed per Patient	Minimum Number of Cells	Counted per Patient
Blood (cancer)	20 cells	20 cells	
Bone Marrow (cancer)	20 cells	20 cells	
Tissue Fibroblasts	15 cells from 2 independent cultures	5 cells split between 2 independent cell cultures	

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For confirmation of chromosomally abnormal amniotic fluid results, or familial chromosome abnormality, examination of fewer cells is permitted.

A number of factors may influence the quality of the metaphase spreading (e.g., humidity, air flow, cell concentration, and cell storage conditions).

An analysis of at least 50 cells is recommended when:

- o Single trisomic cells are found during a study;
- o Mosaicism is suspected on the basis of a phenotype not correlating with the karyotype during the study; or
- o Sex chromosome abnormalities are suspected.

Additionally, when mosaicism is suspected, ensure that adequate number of cells or nuclei are scored.

- o Follow manufacturer's instructions for the use of the probe in accordance with the FDA requirements for "Analyte Specific Reagents (ASR)."
- o Establish or verify test system performance using each new probe and each new lot of probe in accordance with D5421 or D5423; thereafter the laboratory must ensure test methodology performance in accordance with D5411.
- o Establish criteria for scoring the number of probe signals and the number of cells to be examined. Use D5425.

For fragile X analysis:

- o Males - at least 50-100 cells should be scored for negative analysis.
- o Females - at least 100-150 cells should be scored for negative analysis.

The presence of the Xq27.3 fragile site should be confirmed with chromosome banding.

Fragile X studies require low folate medium and media which includes treatment with an antimetabolite such as fluorodeoxyuridine (FUdR), methotrexate, excess thymidine, fluorodeoxycytidine (FdC) or other proven induction systems.

General guidance:

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Examine the karyotypes and a slide from among the laboratory cases and determine if the quality of banding and resolution was sufficient to render the reported interpretation. Examination of the long arm on the 18th chromosome should demonstrate at least two distinct dark staining G-bands at the 400 band level.

Verify that the laboratory's policy establishes a specific band level of resolution that would be dependent upon the study requested.

High resolution chromosome analysis should refer to studies done above the 550 band stage. (Above 650 band stage for an unfocused study. A focused study should be done at a level of resolution at which the band in question is clearly separated from surrounding bands in one member of the homologous pair in question.) Use D5683.

Probes ?493.1276(b)(1)-(b)(3)

For fragile X analysis, if a folate deficient medium is not used as described above, how does the laboratory assure the validity of the test system and the accuracy of results? Use D5411 or D5413, as applicable.

How many photographic and/or computerized karyotypes are prepared from each cell line? (A minimum of 2 is recommended.)

What band level of resolution is used by the laboratory to rule out structural defects (i.e., routine or 400-500 band stage, or high resolution or 650-850 band stage)?

Probes ?493.1276(e)

Each day of use, does the laboratory test the positive and negative reactivity of staining materials to ensure predictable staining characteristics? Use D5473.

Does the laboratory, concurrent with the initial use, check each batch of media for pH (amniotic cell cultures should be kept between pH 6.8 and 7.8), sterility, and ability to support growth? Use D5477.

Does the laboratory employ an alternative procedure for the immediate assessment and monitoring of all testing over time? For example: Control materials are not routinely available to demonstrate chromosome abnormalities for linkage, breakage or translocation, but the laboratory must demonstrate an alternative mechanism for detecting chromosome abnormalities to be analyzed. Use 5485.

An alternative procedure might include spit sample with another laboratory, repeat patient specimen, special stains, FISH assays, and/or molecular assays.

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**FED - D5685 - CLINICAL CYTOGENETICS**

**Title** CLINICAL CYTOGENETICS

**CFR** 493.1276(c)(e)

**Type** Standard

**Regulation Definition**

- (c) Determination of sex must be performed by full chromosome analysis.
- (e) The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Probes §493.1276(e)

Each day of use, does the laboratory test the positive and negative reactivity of staining materials to ensure predictable staining characteristics? Use D5473.

Does the laboratory, concurrent with the initial use, check each batch of media for pH (amniotic cell cultures should be kept between pH 6.8 and 7.8), sterility, and ability to support growth? Use D5477.

Does the laboratory employ an alternative procedure for the immediate assessment and monitoring of all testing over time? For example: Control materials are not routinely available to demonstrate chromosome abnormalities for linkage, breakage or translocation, but the laboratory must demonstrate an alternative mechanism for detecting chromosome abnormalities to be analyzed. Use 5485.

An alternative procedure might include spit sample with another laboratory, repeat patient specimen, special stains, FISH assays, and/or molecular assays.

**FED - D5687 - CLINICAL CYTOGENETICS**

**Title** CLINICAL CYTOGENETICS

**CFR** 493.1276(d)

**Type** Standard

**Regulation Definition**

The laboratory report must include a summary and interpretation of the observations, number of cells counted and analyzed, and use the International System for Human Cytogenetic Nomenclature.

**Interpretive Guideline**

Probes §493.1276(d)

Does the laboratory report include:

- o Type of banding method used, if applicable;
- o Stage of cell mitosis when banded;

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- o Number of cells counted and analyzed microscopically;
- o Number of cells from which photographic or computerized karyotypes were prepared; and
- o Estimate of the banding resolution achieved?

Does the laboratory, where appropriate, ensure that FISH clinical interpretations are made in conjunction with standard cytogenetic analyses and evaluated against patient medical history and other diagnostic test results?

Preliminary reports of karyotypes based on less than full analysis are acceptable if the diagnosis is clear.

For what types of cultures are preliminary reports issued? These may include, but are not limited to, the following:

- o Bone marrow analysis (within 14 days);
- o Unstimulated blood cultures (within 14 days); and
- o Lymphocytes from newborns (within 7 days).

What is the average length of time for reporting (use D5801 or D5815, as appropriate):

- o Amniotic fluid cell cultures (90% of prenatal diagnosis cases should be signed out in 21 days);
- o Routine lymphocyte cultures (approximately 4-5 weeks); and
- o Fibroblast cultures (approximately 2-3 months)?

Do records document:

- o Observations made concurrently with the performance of each step in the examination of specimens/cultures (use D5683); and
- o The number of cases reviewed, signed out and/or the frequency of failed or sub-optimal cultures?

**FED - D5729 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(a)(1)(g)

**Type** Standard

**Regulation Definition**

(a) General. The laboratory must meet the following requirements.

(a)(1) An audible alarm system must be used to monitor the storage temperature of specimens (donor and recipient) and reagents. The laboratory must have an emergency plan for

**Interpretive Guideline**

Interpretative Guidelines §493.1278(a):

When condition-level deficiencies in Histocompatibility are identified in any or all phases of testing, cite D5042.

Interpretative Guidelines §493.1278(a)(1)-(a)(2):

Ultra low (-80oC) freezers and liquid nitrogen (LN2) reservoirs are common in these laboratories. LN2 reservoirs

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alternate storage.

(g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

should be monitored to ensure adequate supply of LN2 at all times.

Verify that the laboratory has an audible alarm system for freezers and refrigerators where critical patient specimens and test reagents are stored. The laboratory should have established the temperature at which the audible alarm will activate. Determine if the laboratory has an emergency power source for this alarm system in the event of an electrical failure. If emergency power is not available, the laboratory should have policies/procedures on how to ensure a prompt response to an activated alarm, 24 hours a day, 7 days a week, including holidays.

An emergency plan for alternate storage of historic patient serum specimens necessary for pre-transplant crossmatching is critical. Verify that the laboratory has an emergency plan for alternate storage appropriate for its operational needs.

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

**FED - D5731 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(a)(2)

**Type** Standard

**Regulation Definition**

(a) General. The laboratory must meet the following requirements.

(a)(2) All patient specimens must be easily retrievable.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(a):

When condition-level deficiencies in Histocompatibility are identified in any or all phases of testing, cite D5042.

Interpretative Guidelines §493.1278(2):

Patient specimens needed for pre-transplant testing should be stored on-site.

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**FED - D5733 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(a)(3)(g)

**Type** Standard

**Regulation Definition**

- (a) General. The laboratory must meet the following requirements.
- (a)(3) Reagent typing sera inventory prepared in-house must indicate source, bleeding date and identification number, reagent specificity, and volume remaining.
- (g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

- Interpretative Guidelines §493.1278(a):  
When condition-level deficiencies in Histocompatibility are identified in any or all phases of testing, cite D5042.
- Interpretative Guidelines §493.1278(g):  
All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

**FED - D5735 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(a)(4)(g)

**Type** Standard

**Regulation Definition**

- (a) General. The laboratory must meet the following requirements.
- (a)(4) If the laboratory uses immunologic reagents (for example, antibodies, antibody-coated particles, or complement) to facilitate or enhance the isolation of lymphocytes, or lymphocyte subsets, the efficacy of the methods must be monitored with appropriate quality control procedures.
- (g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

- Interpretative Guidelines §493.1278(a):  
When condition-level deficiencies in Histocompatibility are identified in any or all phases of testing, cite D5042.
- Interpretative Guidelines 493.1278(a)(4):  
Lymphocytes can be isolated from peripheral blood, lymph nodes and spleen. These cells can be further separated into subsets such as T cells and B cells. Examples of commonly used commercial immunologic reagents include immunomagnetic beads and monoclonal reagents. The laboratory should determine the quality (cell viability), the quantity (final yield), subset specificity (T cell, B cell, etc.), and purity (contaminating cells removed) of the final cell preparation. The laboratory should have policies and/or procedures for assessment of the efficacy of these reagents to include criteria for acceptability. For deficiencies related to the procedure, use D5403; for control material acceptability, use D5469.

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The subset specificity of each lot of immunomagnetic beads should be verified with antiserum specific for each cell type (e.g., T cell beads with anti-T-lymphocyte serum).

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

**FED - D5737 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(a)(5)(g)

**Type** Standard

**Regulation Definition**

(a) General. The laboratory must meet the following requirements.

(a)(5) Participate in at least one national or regional cell exchange program, if available, or develop an exchange system with another laboratory in order to validate interlaboratory reproducibility.

(g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(a):

When condition-level deficiencies in Histocompatibility are identified in any or all phases of testing, cite D5042.

Interpretative Guidelines §493.1278(a)(5):

Programs offered by proficiency testing companies and cell exchanges for histocompatibility laboratories are readily available. An example of a regional exchange program is the Southeastern Organ Procurement Foundation (SEOPF). UCLA provides an international monthly exchange program with sera, cells and DNA. The College of American Pathologists (CAP) and the American Society for Histocompatibility and Immunogenetics (ASHI) each offer programs that assess the primary areas of testing in histocompatibility laboratories by test techniques (i.e., antibody screening and identification, HLA typing for Class I (HLA-A, B, C) and Class II (HLA-DR, DQ), lymphocyte crossmatching (T cell and B cell)).

Laboratories participating in a local exchange should record information concerning the frequency of exchange and the grading system.

Cite a deficiency if the laboratory is not enrolled in a cell exchange program or is enrolled in a program, but fails to return the results. A laboratory's performance in a regional or national exchange program should be evaluated against a peer group performing the same technique.

Interpretative Guidelines §493.1278(g):

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All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

**FED - D5739 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(b)(1)(2)(3)(4)(g)

**Type** Standard

**Regulation Definition**

(b) HLA typing. The laboratory must do the following:

(b)(1) Use a technique(s) that is established to optimally define, as applicable, HLA Class I and II specificities.

(b)(2) HLA type all potential transplant recipients at a level appropriate to support clinical transplant protocol and donor selection.

(b)(3) HLA type cells from organ donors referred to the laboratory.

(b)(4) Use HLA antigen terminology that conforms to the latest report of the World Health Organization (W.H.O.) Committee on Nomenclature. Potential new antigens not yet approved by this committee must have a designation that cannot be confused with W.H.O. terminology.

(g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(b):

HLA (Human Leukocyte Antigens) typing is the identification of histocompatibility antigens and/or alleles. HLA typing is performed by serologic or molecular methods.

Serologic typing is usually performed by incubating viable lymphocytes with antisera of known HLA specificities. Antibodies will bind cells with the corresponding HLA antigen(s) on their surface. When complement is added to an immune complex, it binds to the complex causing cell death. The surface of the lymphocyte becomes permeable to stains and this positivity is determined microscopically.

HLA typing using nucleic acid (DNA) and primers and/or probes involves using the polymerase chain reaction (PCR) to amplify HLA sequences of interest which are detected by gel electrophoresis, ELISA or by fluorescence detection using flow cytometry.

Interpretative Guidelines §493.1278(b)(1):

HLA CLASS I specificities include HLA-A, B, Cw.

HLA CLASS II specificities include HLA-DR, DQ, and DP.

Verify that the laboratory has validated the reagents and methods it uses. For deficiencies related to verification of methods, use D5421; for establishment of methods, use D5423.

Interpretative Guidelines §493.1278(b)(2):

The laboratory should be an active participant of the transplant center's clinical program. It should provide the technical assistance and pertinent data necessary to help establish transplant protocols for solid organ, tissue and cellular transplants and transfusions. Each protocol should specify what HLA specificities should be identified and at

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what level this testing needs to be performed. HLA Class I and Class II typing must be performed in accordance with the protocol.

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

**FED - D5741 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(b)(5)(i)(g)

**Type** Standard

**Regulation Definition**

(b) HLA Typing. The laboratory must do the following:

(b)(5) Have available and follow written criteria for the following:

(b)(5)(i) The preparation of cells or cellular extracts (for example, solubilized antigens and nucleic acids), as applicable to the HLA typing technique(s) performed.

(g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(b)

HLA (Human Leukocyte Antigens) typing is the identification of histocompatibility antigens and/or alleles. HLA typing is performed by serologic or molecular methods.

Serologic typing is usually performed by incubating viable lymphocytes with antisera of known HLA specificities. Antibodies will bind cells with the corresponding HLA antigen(s) on their surface. When complement is added to an immune complex, it binds to the complex causing cell death. The surface of the lymphocyte becomes permeable to stains and this positivity is determined microscopically.

HLA typing using nucleic acid (DNA) and primers and/or probes involves using the polymerase chain reaction (PCR) to amplify HLA sequences of interest which are detected by gel electrophoresis, ELISA or by fluorescence detection using flow cytometry.

Interpretative Guidelines §493.1278(b)(5)(i):

The laboratory's procedure manual should contain cell and /or DNA isolation procedures for each type of specimen it uses (e.g., peripheral blood, lymph nodes and spleen, cell cultures, filter paper blood spots, buccal swabs).

Laboratories should assess pretest viability of cells prior to dotting on typing trays. They may use trypan blue stain, wet preps, etc. Verify that the laboratory maintains records of this activity. For most techniques, viability should exceed 80%.

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Determine if the laboratory has verified their extraction method. Use D5421.

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

**FED - D5743 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(b)(5)(ii)(g)

**Type** Standard

**Regulation Definition**

- (b) HLA Typing. The laboratory must do the following:
- (b)(5) Have available and follow written criteria for the following:
- (b)(5)(ii) Selecting typing reagents, whether prepared in-house or commercially.
- (g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(b)

HLA (Human Leukocyte Antigens) typing is the identification of histocompatibility antigens and/or alleles. HLA typing is performed by serologic or molecular methods.

Serologic typing is usually performed by incubating viable lymphocytes with antisera of known HLA specificities. Antibodies will bind cells with the corresponding HLA antigen(s) on their surface. When complement is added to an immune complex, it binds to the complex causing cell death. The surface of the lymphocyte becomes permeable to stains and this positivity is determined microscopically.

HLA typing using nucleic acid (DNA) and primers and/or probes involves using the polymerase chain reaction (PCR) to amplify HLA sequences of interest which are detected by gel electrophoresis, ELISA or by fluorescence detection using flow cytometry.

Interpretative Guidelines §493.1278(b)(5)(ii)

For HLA complement dependent lymphocytotoxicity typing, each batch of complement must be tested to determine that it mediates cytotoxicity (cell death) in the presence of a specific HLA antibody, but is not cytotoxic in the absence of a specific antibody. The test should ensure that it is maximally active at least one dilution beyond that intended for use. The test should be carried out with at least two antibodies known to react with at least two different cells (positive control), and at least one cell which should not react (negative control). A strong and a weak antibody should be selected for the test. Serial dilutions of a single serum may also be used. Verify that the laboratory has performed complement quality control and that an optimum dilution has been selected and documented. Complement is temperature sensitive (labile) and should be retitered periodically to ensure its activity. Determine if the laboratory

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has complement retitering policies/procedures.

The results of each batch/lot of reagents (typing trays) whether commercially made or prepared in-house must be reviewed to determine which sera failed to react as expected (false negative reactions) and which sera had unexpected reactions (false positive reactions). Future tray preparation and interpretation of commercially purchased trays should be evaluated and revised based on the results of these reviews.

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

Probes §493.1278(b)(5)(ii):

What criteria were used to determine the acceptability of each batch of complement for HLA serologic assays?

How does the laboratory select the typing trays it uses for each patient?

**FED - D5745 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(b)(5)(iii)(g)

**Type** Standard

**Regulation Definition**

(b) HLA Typing. The laboratory must do the following:  
(b)(5) Have available and follow written criteria for the following:  
(b)(5)(iii) Ensuring that reagents used for typing are adequate to define all HLA-A, B and DR specificities that are officially recognized by the most recent W.H.O. Committee on Nomenclature and for which reagents are readily available.  
(g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(b)

HLA (Human Leukocyte Antigens) typing is the identification of histocompatibility antigens and/or alleles. HLA typing is performed by serologic or molecular methods.

Serologic typing is usually performed by incubating viable lymphocytes with antisera of known HLA specificities. Antibodies will bind cells with the corresponding HLA antigen(s) on their surface. When complement is added to an immune complex, it binds to the complex causing cell death. The surface of the lymphocyte becomes permeable to stains and this positivity is determined microscopically.

HLA typing using nucleic acid (DNA) and primers and/or probes involves using the polymerase chain reaction (PCR) to amplify HLA sequences of interest which are detected by gel electrophoresis, ELISA or by fluorescence detection using flow cytometry.

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Interpretative Guidelines §493.1278(b)(5)(iii):

Antisera for less frequent and rare specificities may be unavailable to laboratories. It is good laboratory practice for each HLA antigen to be defined by at least two operationally monospecific sera. Typing for (HLA) class I or class II antigens must employ a sufficient number of antisera or monoclonal antibodies to clearly define all the antigens for which the laboratory tests. For example: If multispecific sera must be used, at least three partially non-overlapping sera should be used to define each HLA-antigen. For each HLA-DR and HLA-DQ antigen to be defined, at least 3 operationally monospecific sera should be used. If multispecific sera must be used, at least 5 partially non-overlapping sera should be used.

The laboratory should demonstrate that typing sera reactions are recorded, reviewed and used to modify locally prepared typing trays and interpret commercial tray specificities.

Primer and/or probe sequence, specificity and sensitivity should be defined with reference material (previously typed DNA). For typing methods using probe technology, verify whether optimum hybridization temperatures have been verified or established for each probe.

The laboratory should demonstrate that reference material testing is recorded regularly, reviewed and used to modify locally prepared reagents, as well as interpret commercial primer and/or probe specificities.

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

Probes §493.1278(b)(5)(iii):

How are the specificities of new typing sera, primers and probes (whether local or commercial) verified, e.g., by parallel testing with known cells or DNA?

How does the laboratory report HLA typings performed by serology and DNA (i.e., follow the W.H.O. nomenclature list)? Are antigens and alleles reported appropriately?

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**FED - D5747 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(b)(5)(iv)(g)

**Type** Standard

**Regulation Definition**

- (b) HLA Typing. The laboratory must do the following:
- (b)(5) Have available and follow written criteria for the following:
  - (b)(5)(iv) The assignment of HLA antigens.
  - (g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(b)

HLA (Human Leukocyte Antigens) typing is the identification of histocompatibility antigens and/or alleles. HLA typing is performed by serologic or molecular methods.

Serologic typing is usually performed by incubating viable lymphocytes with antisera of known HLA specificities. Antibodies will bind cells with the corresponding HLA antigen(s) on their surface. When complement is added to an immune complex, it binds to the complex causing cell death. The surface of the lymphocyte becomes permeable to stains and this positivity is determined microscopically.

HLA typing using nucleic acid (DNA) and primers and/or probes involves using the polymerase chain reaction (PCR) to amplify HLA sequences of interest which are detected by gel electrophoresis, ELISA or by fluorescence detection using flow cytometry.

Interpretative Guidelines §493.1278(b)(5)(iv):

Criteria for antigen and/or allele assignment must take into account basic principles of genetic inheritance.

Examples:

1. No more than 2 antigens or alleles per HLA-A, B, and DR locus can be assigned to any patient; e.g., antigens HLA-A2, A24; B46, B61; DR8, 14; alleles HLA-A\*02XX, 24XX; B\*4002, 4601; DRB1\*0803, 1401. Public specificities may be observed; i.e. for HLA-B, additional specificities of Bw4 and/or Bw6 are reported, for Class II antigens, additional gene products of DR51, DR52 and/or DR53 are reported.
2. When family studies are performed, typing interpretations should be in accordance with genetic relationships (i.e., haplotype assignments, determination of homozygosity at a particular locus).

Verify that the laboratory has established acceptability criteria for assignment of HLA antigens and/or alleles. Examples for alleles include signal intensity, band clarity and migration, specificity, and procedures to resolve ambiguous alternative combinations.

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Determine if testing personnel follow the scoring and reporting system defined in the procedure manual. Two independent interpretations are recommended for each DNA analysis. Determine if the laboratory has validated computer software for the analysis of antigens and/or alleles.

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

**FED - D5749 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(b)(5)(v)(g)

**Type** Standard

**Regulation Definition**

(b) HLA Typing. The laboratory must do the following:  
(b)(5) Have available and follow written criteria for the following:  
(b)(5)(v) When antigen redefinition and retyping are required.  
(g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(b)

HLA (Human Leukocyte Antigens) typing is the identification of histocompatibility antigens and/or alleles. HLA typing is performed by serologic or molecular methods.

Serologic typing is usually performed by incubating viable lymphocytes with antisera of known HLA specificities. Antibodies will bind cells with the corresponding HLA antigen(s) on their surface. When complement is added to an immune complex, it binds to the complex causing cell death. The surface of the lymphocyte becomes permeable to stains and this positivity is determined microscopically.

HLA typing using nucleic acid (DNA) and primers and/or probes involves using the polymerase chain reaction (PCR) to amplify HLA sequences of interest which are detected by gel electrophoresis, ELISA or by fluorescence detection using flow cytometry.

Interpretative Guidelines §493.1278(b)(5)(v):

Verify that the laboratory has policies and procedures for antigen and/or allele redefinition and retyping. Records should indicate that results from redefinition and retyping are evaluated and that patient typings are updated accordingly. Discrepancies identified as the result of this activity should be documented and resolved. Use D5775.

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data,

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manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

**FED - D5751 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(b)(6)(g)

**Type** Standard

**Regulation Definition**

(b) HLA Typing. The laboratory must do the following:

(b)(6) Check each HLA typing by testing, at a minimum the following:

(b)(6)(i) A positive control material.

(b)(6)(ii) A negative control material in which, if applicable to the technique performed, cell viability at the end of incubation is sufficient to permit accurate interpretation of results. In assays in which cell viability is not required, the negative control result must be sufficiently different from the positive control result to permit accurate interpretation of results.

(b)(6)(iii) Positive control materials for specific cell types when applicable (that is, T cells, B cells, and monocytes).

(g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(b)

HLA (Human Leukocyte Antigens) typing is the identification of histocompatibility antigens and/or alleles. HLA typing is performed by serologic or molecular methods.

Serologic typing is usually performed by incubating viable lymphocytes with antisera of known HLA specificities. Antibodies will bind cells with the corresponding HLA antigen(s) on their surface. When complement is added to an immune complex, it binds to the complex causing cell death. The surface of the lymphocyte becomes permeable to stains and this positivity is determined microscopically.

HLA typing using nucleic acid (DNA) and primers and/or probes involves using the polymerase chain reaction (PCR) to amplify HLA sequences of interest which are detected by gel electrophoresis, ELISA or by fluorescence detection using flow cytometry.

Interpretative Guidelines §493.1278(b)(6):

Each HLA-A, B, C or supplemental Class I typing tray must include at least one positive control serum, previously shown to react with all lymphocytes, and one negative control serum which has been demonstrated to be non-cytotoxic. HLA-DR and DQ typing trays must include a positive control serum, previously shown to react with only B cells, and one negative control serum which has been demonstrated to be non-cytotoxic.

Cell controls must be tested with each batch/lot/shipment of typing trays. Typing results are invalid if controls fail to react as expected. The negative control should either be one previously shown to lack antibody or should be from a healthy male with no history of blood transfusion. Cell viability in the negative control well at the end of the incubation must be sufficient to permit accurate interpretation of results. For most techniques, viability should exceed 80%. However, when less than optimal specimens, such as cadaver and mailed specimens, this threshold may not be met.

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For DNA typing, negative control wells or wells with no DNA should not give a positive result (the presence of a band), however, internal controls should give a positive result. DNA reference material must be tested with each lot of typing reagents. Primers and/or probes must be tested for allele specificity with reference material.

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

**FED - D5753 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(c)(g)

**Type** Standard

**Regulation Definition**

(c) Disease-associated studies. The laboratory must check each typing for disease-associated HLA antigens using control materials to monitor the test components and each phase of the test system to ensure acceptable performance.

(g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(c):

Disease association studies are single or limited antigen typings usually performed by serologic typing methods and more rarely performed by flow cytometric methods.

Positive and negative controls must be run with each test.

Control cells must be tested with each lot and shipment of reagents. Use D5753.

For serologic typings, the control cells should include at least two cells known to express the specified antigen and two cells known to express cross-reacting antigens that might be confused with the specific antigen. Control cells should also include at least two cells lacking the specific and cross-reacting antigen.

For typing sera acceptability, use D5745.

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

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**FED - D5755 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(d)(1)(2)(3)(g)

**Type** Standard

**Regulation Definition**

(d) Antibody Screening. The laboratory must do the following:

(d)(1) Use a technique(s) that detects HLA-specific antibody with a specificity equivalent or superior to that of the basic complement-dependent microlymphocytotoxicity assay.

(d)(2) Use a method that distinguishes antibodies to HLA Class II antigens from antibodies to Class I antigens to detect antibodies to HLA Class II antigens.

(d)(3) Use a panel that contains all the major HLA specificities and common splits. If the laboratory does not use commercial panels, it must maintain a list of individuals for fresh panel bleeding.

(g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(d)(1)-(d)(3):

An antibody screen is performed to identify whether a patient's serum contains antibodies to one or more HLA antigens. This is accomplished by screening the serum against target antigens from a suitable panel appropriate for the population served, i.e., a variety of ethnic groups. Results are expressed as percent reactive antibodies (PRA).

The panel of antigens used must include all of the HLA antigens to which the most common HLA antibodies are formed. Cell panels of known HLA type must be available to prove the specificity of new antibodies. The serum cell panel should be consistent from month to month and from lot to lot. Verify that the frequency of each antigen represented does not vary significantly.

An example of PRA differences from panel to panel:

If a patient demonstrates a HLA-A2 antibody and the cell panel contains 15 A2 positive cells out of 100, the patient's PRA on this tray will be 15%. If the same patient is tested against a panel where there are 37 A2 positive cells out of 100, the patient's PRA will increase to 37%. The number of A2 positive cells on this laboratory's cell panel should reflect the frequency observed in the population it serves; e.g., 15-20% of the local population possess the HLA-A2 antigen.

If the laboratory tests for antibodies to Class II antigens, the laboratory should have a procedure for removing Class I antibodies or should use purified Class II antigens. Class II antigens (HLA-DR, DQ) are found only on the B cell subset of lymphocytes. B cells also have a high density of Class I antigens (HLA-A, B, C), which are found on all nucleated cells. If a patient has a significant titer of Class I antibodies, it may result in a false positive Class II antibody test result. Platelet absorption is one method of removing the Class I antibodies.

Verify that the laboratory's antibody screening technique is as sensitive as the crossmatch method it uses to ensure optimum compatibility.

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data,

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manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

**FED - D5757 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(d)(4)(5)(g)

**Type** Standard

**Regulation Definition**

- (d) Antibody Screening. The laboratory must do the following:
- (d)(4) Make a reasonable attempt to have available monthly serum specimens for all potential transplant recipients for periodic antibody screening and crossmatch.
  - (d)(5) Have available and follow a written policy consistent with clinical transplant protocols for the frequency of screening potential transplant recipient sera for preformed HLA-specific antibodies.
  - (g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(d)(4)-(d)(5):

A recipient's antibody profile should be evaluated when the individual is entered on the transplant waiting list. Determine whether the laboratory obtains specimens at initial typing for antibody screening and for pre-transplantation auto crossmatches.

The laboratory should have clearly defined and appropriate screening protocols for potentially sensitizing events such as transfusion, transplant loss, pregnancy or infection. Verify that the laboratory obtains and tests patient specimens to determine if there have been changes in the antibody profiles as defined by the transplant center's protocols. Determine when the laboratory verifies that the antibodies in the serum have been characterized against HLA antigens.

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

Probes §493.1278(d)(4)-(d)(5)

What policies and procedures has the laboratory implemented in an effort to procure monthly serum specimens for potential transplant recipients?

What is the laboratory's frequency for screening potential transplant recipient sera for preformed HLA-specific antibodies?

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**FED - D5759 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(d)(6)(g)

**Type** Standard

**Regulation Definition**

- (d) Antibody Screening. The laboratory must do the following:
- (d)(6) Check each antibody screening by testing, at a minimum the following:
- (d)(6)(i) A positive control material containing antibodies of the appropriate isotype for the assay.
- (d)(6)(ii) A negative control material.
- (g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(d)(6):

For serologic antibody screening, each tray must include at least one positive control serum previously shown to react with all lymphocytes and one negative control serum which has been demonstrated to be non-cytotoxic or lack antibody. Results are invalid if controls fail to react as expected. Cell viability in the negative control well at the time of reading must be sufficient to permit accurate interpretation of results. Viability should exceed 80%. The positive control must contain antibodies of the appropriate isotype (e.g., IgG and/or IgM). If the frozen cell tray is specific for Class II (HLA-DR or DQ) antibody testing, the laboratory must ensure B cells are being tested and have a mechanism to distinguish Class II antibodies from antibodies to Class I antigens that are also found on B cells.

Laboratories using ELISA and/or flow cytometric techniques must include one positive control serum and one negative control serum. Reagent controls for non-specific binding of antibody should be included with all ELISA testing. The negative control for flow cytometers should demonstrate non-reactivity and the positive control should be specific for HLA antigens. Again, the positive control for both techniques must contain antibodies of the appropriate isotype (i.e., IgG and/or IgM).

Verify that the laboratory uses a negative control and the appropriate isotype for its positive control.

Verify that the laboratory has established acceptability criteria for each control and for each method it uses.

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

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**FED - D5761 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(d)(7)(g)

**Type** Standard

**Regulation Definition**

(d) Antibody Screening. The laboratory must do the following:  
(d)(7) As applicable, have available and follow written criteria and procedures for antibody identification to the level appropriate to support clinical transplant protocol.  
(g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

Probe §493.1278(d)(7):

Do the laboratory's policies specify when antibody reactivity (positive antibody screen) will be further characterized, (i.e., identification of antibody directed against specific HLA antigens) and the procedures to be used for antibody identification?

**FED - D5763 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(e)(1)(g)

**Type** Standard

**Regulation Definition**

(e) Crossmatching. The laboratory must do the following:  
(e)(1) Use a technique(s) documented to have increased sensitivity in comparison with the basic complement-dependent microlymphocytotoxicity assay.  
(g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(e)(1):

The minimum technique for crossmatching for transplantation must be more sensitive than the basic lymphocytotoxicity test (standard complement dependent or NIH procedure). A technique that enhances sensitivity must be used (e.g., increased incubation time, additional wash steps, antihumanglobulin (AHG) augmentation, ELISA testing, flow cytometry testing).

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays,

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primers and/or probes. Do not dictate the acceptable format for documentation.

**FED - D5765 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(e)(2)(g)

**Type** Standard

**Regulation Definition**

- (e) Crossmatching. The laboratory must do the following:
- (e)(2) Have available and follow written criteria for the following:
- (e)(2)(i) Selecting appropriate patient serum samples for crossmatching.
- (e)(2)(ii) The preparation of donor cells or cellular extracts (for example, solubilized antigens and nucleic acids), as applicable to the crossmatch technique(s) performed.
- (g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(e)(2)(i):

The laboratory must have clearly defined protocols for selection of serum for crossmatch testing. There are numerous acceptable protocols for the selection of crossmatch samples which vary from transplant center to center. However, every effort should be made to procure a specimen at the time of transplant or unless the laboratory can clearly establish that the patient did not receive a blood transfusion or other alloimmunizing event between the times of specimen collection and transplant date.

Review patient transplant records for lymphocyte crossmatch results. Verify serum selected for crossmatching against antibody screening/identification records. Verify if the serum is tested at an optimal dilution. Crossmatches are performed with donor T cells (T lymphocytes) or unseparated lymphocytes. Crossmatches with donor B cells (B lymphocytes ) may be performed.

Interpretative Guidelines §493.1278(e)(2)(ii):

There are various techniques for the isolation of donor cells for use in crossmatching e.g., immunomagnetic beads, monoclonal antibody preparations, density gradient (ficoll hypaque). Crossmatching techniques utilizing cellular extracts (solubilized antigens and nucleic acid) are not well documented in the clinical setting.

Determine if the laboratory follows manufacturer's product insert procedures. Use D5479.

Verify that the laboratory has established procedures and criteria for cell preparation viability, purity and quantity (i.e. peripheral blood, lymph node, spleen).

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

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Probes §493.1278(e)(2)(i):

Does the laboratory's policies and procedures specify which patient serum samples are to be used for crossmatching (e.g., renal, pancreas, heart, lung, small intestine or liver transplants)?

**FED - D5767 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(e)(3)(g)

**Type** Standard

**Regulation Definition**

- (e) Crossmatching. The laboratory must do the following:
- (e)(3) Check each crossmatch and compatibility test for HLA Class II antigenic differences using control materials to monitor the test components and each phase of the test system to ensure acceptable performance.
- (g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(e)(3):

The mixed leukocyte (lymphocyte) culture (MLC) is used by a small number of laboratories and it may be used in conjunction with other cellular assays such as cell mediated lympholysis (CML), primed lymphocyte typing (PLT) or homozygous typing cell (HTC) to determine donor recipient pair compatibility in renal or tissue transplants.

The MLC method may vary from micro, macro, one way or both one way, and two way. Data expressed in counts per minute of tritiated thymidine (H3) are used to calculate the stimulation index (SI) or the relative response (RR). Controls include: a negative control (responder cells stimulated with autologous cells), positive controls (responder cells stimulated with cells from unrelated individuals with known Class II antigen differences or fresh or frozen cell pool). If the laboratory performs MLCs, review their criteria for accepting or rejecting a run and a narrative report on donor recipient compatibility. Confirm that all combinations of any given stimulator is tested against any given responder.

Verify that the laboratory has established criteria for defining positive and negative crossmatches.

Example 1:

Basic crossmatch technique: (includes increased incubation time testing or wash(es))

- 1) Each crossmatch tray must include one positive control serum previously shown to react with all cells and one negative control serum which demonstrates non-cytotoxic activity. Additional controls may include antisera against specific cell lines and reagent controls.
- 2) Each serum is tested undiluted and at one or more dilutions.

Example 2:

Anti-human globulin augmentation:

- 1) Each crossmatch tray must include one positive control serum previously shown to react with all cells and one

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negative control serum which demonstrates non-cytotoxic activity. Additional controls may include antisera against specific cell lines and reagent controls.

- 2) Each serum is tested undiluted and at one or more dilutions.
- 3) Verify that AHG has been titered for optimum test performance.

Example 3:

Flow cytometry:

- 1) Each crossmatch must include one positive control serum and one negative control serum. The positive control should be human serum of the appropriate isotype and specific for HLA antigens shown to react with all cells. The negative control should demonstrate non-reactivity against lymphocytes.
- 2) Verify that the laboratory has established a threshold for determining a positive reaction (e.g., mean channel shifts, quantitative fluorescence measurements).
- 3) The laboratory should be running an optical standard (lens focusing and alignment) and fluorescent standard (adequate signal amplification) with each use of the instrument.
- 4) Verify that the laboratory has established an optimum serum/cell ratio (standard number of cells to a fixed volume of serum).
- 5) A multi color technique should be used to ensure the purity of the cell population being tested.

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

Probes §493.1278(e)(3):

What is the laboratory's control acceptance criteria for MLC testing?

**FED - D5769 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(f)(1)(g)

**Type** Standard

**Regulation Definition**

(f) Transplantation. Laboratories performing histocompatibility testing for transfusion and transplantation purposes must do the following:

**Interpretive Guideline**

Interpretative Guidelines §493.1278(f):

In conjunction with the transplantation center the laboratory establishes written policies on the testing protocols it performs in support of the clinical transplant program. Policies should address when HLA testing and final

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(f)(1) Have available and follow written policies and protocols specifying the histocompatibility testing (that is, HLA typing, antibody screening, compatibility testing and crossmatching) to be performed for each type of cell, tissue or organ to be transfused or transplanted. The laboratory's policies must include, as applicable--

(f)(1)(i) Testing protocols for cadaver donor, living, living-related, and combined organ and tissue transplants;

(f)(1)(ii) Testing protocols for patients at high risk for allograft rejection; and

(f)(1)(iii) The level of testing required to support clinical transplant protocols (for example, antigen or allele level).

(g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

crossmatches are required for patients that have demonstrated presensitization. For organs such as liver and heart (non-renal), it is not uncommon for laboratories to perform retrospective crossmatches if the patient demonstrates the absence of preformed antibodies by prior screening. Failure to perform a crossmatch prior to transplant is not a deficiency provided emergency transplant circumstances are documented.

For solid organ transplants (renal, heart, liver, lung, small intestine):

1. Determine what tests are performed for potential kidney and pancreas recipients.
2. Determine what tests are performed on living-related or unrelated donors and cadaver donors referred to the laboratory.
3. Determine if the laboratory performs HLA typing using complement dependent lymphocytotoxicity testing (antigen level) and/or DNA testing (allele level);
4. Compare policies for pre-sensitized patients with laboratory antibody screening and identification protocols for consistency;
5. Verify that the laboratory is using a crossmatch technique with increased sensitivity; and
6. Deviations from the established protocols should be documented by the laboratory, indicating the reason for the deviation, e.g., transplant physician request, emergency transplant.

For transfusions (platelet support of refractory patients):

1. Determine what tests are performed on recipients and donors. Recipients are usually HLA-A and HLA-B typed, e.g., platelets do not have Class II (HLA-DR, DQ) antigens on their surface. Donors may be typed by the laboratory, a blood center or a donor program laboratory. HLA typing may be performed using complement dependent lymphocytotoxicity testing (antigen level) and/or DNA testing (allele level).
2. Determine if the laboratory performs antibody screening/identification on the recipient. Compare with the laboratory protocol for antibody screening and identification.
3. Determine if the laboratory performs Class I crossmatch testing.

For tissue transplant (bone marrow/stem cells, etc.)

1. Determine what level of HLA typing is performed on recipients and donors. For bone marrow/stem cell transplantation, recipients are at a minimum HLA-A and HLA-B typed by complement dependent lymphocytotoxicity and/or DNA testing. Recipients should be HLA-DR typed by high resolution DNA typing (allele level). Donors may be typed by the laboratory or a donor program laboratory.
2. Determine if the laboratory performs crossmatch testing, when a selected potential donor has an HLA mismatch. Determine if the laboratory performs Class II compatibility to evaluate Class II identity by either MLC testing, high resolution DNA typing, or a family study.

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Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

§493.1278(f) Probes:

What is the laboratory's policy/protocol on referring patient specimens for testing at another laboratory?

What is the laboratory's policy/protocol on accepting HLA typing results obtained at another laboratory (i.e., does the laboratory reconfirm (repeat) testing)?

**FED - D5771 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(f)(2)(g)

**Type** Standard

**Regulation Definition**

(f) Transplantation. Laboratories performing histocompatibility testing for transfusion and transplantation purposes must do the following:  
(f)(2) For renal allotransplantation and combined organ and tissue transplants in which a kidney is to be transplanted, have available results of final crossmatches before the kidney is transplanted.  
(g) Documentation. The laboratory must document all control procedures performed, as specified in this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1278(g):

All QC records must be maintained for two years including instrument charts, graphs, printouts, transcribed data, manufacturer's assay information sheet for control and calibration materials and reagents to include typing trays, primers and/or probes. Do not dictate the acceptable format for documentation.

Probes §493.1278(f)(2):

If the laboratory performs cadaveric renal transplant testing, what are the staffing policies and how do they ensure 24-hour coverage of qualified testing personnel and supervision for technical review?

**FED - D5773 - HISTOCOMPATIBILITY**

**Title** HISTOCOMPATIBILITY

**CFR** 493.1278(f)(3)

**Type** Standard

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**Regulation Definition**

(f) Transplantation. Laboratories performing histocompatibility testing for transfusion and transplantation purposes must do the following:

(f)(3) For nonrenal transplantation, if HLA testing and final crossmatches were not performed prospectively because of an emergency situation, the laboratory must document the circumstances, if known, under which the emergency transplant was performed, and records of the transplant must reflect any information provided to the laboratory by the patient's physician.

**Interpretive Guideline**

**FED - D5775 - COMPARISON OF TEST RESULTS**

**Title** COMPARISON OF TEST RESULTS

**CFR** 493.1281(a)(c)

**Type** Standard

**Regulation Definition**

(a) If a laboratory performs the same test using different methodologies or instruments, or performs the same test at multiple testing sites, the laboratory must have a system that twice a year evaluates and defines the relationship between test results using the different methodologies, instruments, or testing sites.

(c) The laboratory must document all test result comparison activities.

**Interpretive Guideline**

Interpretative Guidelines §493.1281(a)-(c):

The laboratory must have a system to monitor and evaluate all testing it performs. Examples of materials that may be used to evaluate the same test performed by different methodologies, at multiple locations, and/or on multiple instruments in the same laboratory are proficiency testing samples, split samples or "blind" testing of materials with known values.

A laboratory that performs the same test at multiple locations or on more than one instrument must have written criteria for acceptable differences in test values (e.g., between different or identical models of an instrument from the same manufacturer, between instruments from different manufacturers).

If the laboratory performs calibration verification as specified in §493.1255(b), it may use the calibration verification to meet the requirements at §493.1281(a), provided the 3 levels of materials used for calibration verification meet the laboratory's criteria for acceptable differences in test values.

Interpretative Guidelines §493.1281(c):

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The actual measurement(s) of test results and comparison activities must be recorded. Acceptable formats for documentation may vary. Cite documentation deficiencies at §493.1281(a) or §493.1281(b). Use D5775 or D5777, as appropriate.

**FED - D5777 - COMPARISON OF TEST RESULTS**

**Title** COMPARISON OF TEST RESULTS

**CFR** 493.1281(b)(c)

**Type** Standard

**Regulation Definition**

(b) The laboratory must have a system to identify and assess patient test results that appear inconsistent with the following relevant criteria, when available:

- (b)(1) Patient age.
- (b)(2) Sex.
- (b)(3) Diagnosis or pertinent clinical data.
- (b)(4) Distribution of patient test results.
- (b)(5) Relationship with other test parameters.

(c) The laboratory must document all test result comparison activities.

**Interpretive Guideline**

Interpretative Guidelines §493.1281(b):

Verify that the laboratory has a system in place to monitor and evaluate test results for inconsistencies with patient information, and for correlation between test results. For example, a laboratory could multiply the hemoglobin result by a factor of 3, to see if the result is equal to the hematocrit. If the laboratory has auto-validation in its Laboratory Information System (LIS), verify that the laboratory is taking steps to reduce the likelihood of sample-switching errors, for example, when the creatinine result is significantly different from the patient's previous creatinine test results, or if the MCV is significantly different from the patient's previous test results and the patient did not receive a blood transfusion.

For automated laboratories, inconsistent patient results may be evaluated through the use of verified LIS supported logic, patient distribution test results, verified automated test comparison logic programs and individual test repeat criteria.

Interpretative Guidelines §493.1281(c):

The actual measurement(s) of test results and comparison activities must be recorded. Acceptable formats for documentation may vary. Cite documentation deficiencies at §493.1281(a) or §493.1281(b). Use D5775 or D5777, as appropriate.

Probes §493.1281(b):

How does the laboratory obtain sufficient information to enable an evaluation of test results with clinically relevant patient information?

Does the laboratory have procedures to assess and evaluate patient test results for inconsistencies?

For example:

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- o Hemoglobin and Hematocrit (MCHC value exceeds reference range);
- o BUN and Creatinine comparison;
- o Albumin and Total Protein;
- o Correlation of urine culture with urine microscopic; and
- o Alkaline phosphatase with orthopedic surgical patients and/or pediatric patients; and
  - . Correlation of microscopic sediment findings with macroscopic results, such as, the presence of protein with casts, positive occult blood with red cells, and positive leukocyte esterase with white cells.

**FED - D5779 - CORRECTIVE ACTIONS**

**Title** CORRECTIVE ACTIONS

**CFR** 493.1282(a)

**Type** Standard

**Regulation Definition**

Corrective action policies and procedures must be available and followed as necessary to maintain the laboratory's operation for testing patient specimens in a manner that ensures accurate and reliable patient test results and reports.

**Interpretive Guideline**

Interpretative Guidelines §493.1282(a):

Corrective action must be taken when unacceptable differences in test values occur with testing performed using different methodologies or instruments or with the same test performed at multiple testing sites.

Probes §493.1282(a):

When test results do not correlate with patient information (e.g., age, sex, submitted diagnosis) what actions are taken by the laboratory to confirm test results or patient information?

**FED - D5781 - CORRECTIVE ACTIONS**

**Title** CORRECTIVE ACTIONS

**CFR** 493.1282(b)(1)

**Type** Standard

**Regulation Definition**

(b) The laboratory must document all corrective actions taken, including actions taken when any of the following occur:  
(b)(1) Test systems do not meet the laboratory's verified or established performance specifications, as determined in

**Interpretive Guideline**

Interpretative Guidelines §493.1282(b)(1)

The laboratory's corrective action records should contain sufficient information to resolve the problem and prevent reoccurrence.

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§493.1253(b), which include but are not limited to--

(b)(1)(i) Equipment or methodologies that perform outside of established operating parameters or performance specifications;

(b)(1)(ii) Patient test values that are outside of the laboratory's reportable range of test results for the test system; and

(b)(1)(iii) When the laboratory determines that the reference intervals (normal values) for a test procedure are inappropriate for the laboratory's patient population.

Probes §493.1282(b)(1)

When equipment malfunctions or a test method problem exists, how does the laboratory identify and solve the problem?

What corrective actions are taken if patient test results fall outside of the laboratory's reportable range of patient test results?

If a dilution procedure is used when patient results exceed the test system's reportable range, how does the laboratory ensure the appropriate diluent is used for each type of specimen? Use D5401.

How does the laboratory verify and document the accuracy of the results for diluted specimens? Use D5421 or D5423 as appropriate.

**FED - D5783 - CORRECTIVE ACTIONS**

**Title** CORRECTIVE ACTIONS

**CFR** 493.1282(b)(2)

**Type** Standard

**Regulation Definition**

(b) The laboratory must document all corrective actions taken, including actions taken when any of the following occur:

(b)(2) Results of control or calibration materials, or both, fail to meet the laboratory's established criteria for acceptability.

All patient test results obtained in the unacceptable test run and since the last acceptable test run must be evaluated to determine if patient test results have been adversely affected. The laboratory must take the corrective action necessary to ensure the reporting of accurate and reliable patient test results.

**Interpretive Guideline**

Interpretative Guidelines §493.1282(b)(2):

When an internal control fails to fall within the defined limits of acceptability, the laboratory must identify the reason for the failure and correct the problem before resuming testing of patients. The laboratory must evaluate all patients test results since the last acceptable external control.

Probes §493.1282(b)(2):

When suboptimal staining or improper coverslipping are identified through quality control procedures, what corrective actions does the laboratory take?

What actions does the laboratory take when controls reflect an unusual trend or are outside of the acceptable limits and other means of assessing and correcting unacceptable control values have failed to identify and correct the problem?

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**FED - D5785 - CORRECTIVE ACTIONS**

**Title** CORRECTIVE ACTIONS

**CFR** 493.1282(b)(3)

**Type** Standard

**Regulation Definition**

(b) The laboratory must document all corrective actions taken, including actions taken when any of the following occur:  
(b)(3) The criteria for proper storage of reagents and specimens, as specified under §493.1252(b), are not met.

**Interpretive Guideline**

Probes §493.1282(b)(3):  
What action does the laboratory take if the storage temperature for a test system's reagents falls outside the acceptable limits?

**FED - D5787 - TEST RECORDS**

**Title** TEST RECORDS

**CFR** 493.1283(a)

**Type** Standard

**Regulation Definition**

The laboratory must maintain an information or record system that includes the following:  
(a)(1) The positive identification of the specimen.  
(a)(2) The date and time of specimen receipt into the laboratory.  
(a)(3) The condition and disposition of specimens that do not meet the laboratory's criteria for specimen acceptability.  
(a)(4) The records and dates of all specimen testing, including the identity of the personnel who performed the test(s).

**Interpretive Guideline**

Interpretative Guidelines §493.1283(a):  
The regulations provide laboratories the flexibility to establish a system that ensures positive patient identification through specimen accessioning and storage, testing and reporting of test results. This may include a system that involves labeling the specimen container and request slip or the patient's medical record or chart with a unique patient identification number, but does not preclude the use of other mechanisms to assist in patient identification and tracking of specimens throughout the testing and reporting processes. The patient's name may be used as part of the identification system.

Ensure that work records reflect all the tests and dates of performance of in-house patient testing. For example, in bacteriology, each step from media inoculation to organism isolation and identification must be documented on worksheet records either manually or in a computer system.

Corrections of laboratory results include the corrected result, incorrect result (noted as such), the date of the correction, and the initials of the person making the correction. Laboratory records should not be documented in

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pencil and the use of whiteout is not acceptable for making corrections.

Probes §493.1283(a):

Do the records reflect all patient testing and the dates of their performance?

If handwritten values were reported, can the laboratory demonstrate the analytic source of those results?

If the laboratory has not retained the appropriate test records, cite D3031, D3033, or D3035.

**FED - D5789 - TEST RECORDS**

**Title** TEST RECORDS

**CFR** 493.1283(b)

**Type** Standard

**Regulation Definition**

Records of patient testing including, if applicable, instrument printouts, must be retained.

**Interpretive Guideline**

Interpretative Guidelines §493.1283(b):

The regulations do not require that instrument printouts be posted directly in the patient's medical record or chart. However, these printouts must be maintained as part of the laboratory's record retention requirements specified throughout the regulations.

Probes §493.1283(b):

Are the original analytic work records complete (e.g., in a randomly chosen sample, is there an instrument printout for every day of the month on which testing was performed)? Are the original, as opposed to transcribed and/or edited work records, being retained? If the laboratory fails to retain the records for the appropriate amount of time, use D3031.

**FED - D5791 - ANALYTIC SYSTEMS QUALITY ASSESSMENT**

**Title** ANALYTIC SYSTEMS QUALITY ASSESSMENT

**CFR** 493.1289(a)(c)

**Type** Standard

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**Regulation Definition**

(a) The laboratory must establish and follow written policies and procedures for an ongoing mechanism to monitor, assess, and when indicated, correct problems identified in the analytic systems specified in §§493.1251 through 493.1283.

(c) The laboratory must document all analytic systems assessment activities.

**Interpretive Guideline**

Interpretative Guidelines §493.1289(a)-(c):

Quality Assessment (QA) is an ongoing review process that encompasses all facets of the laboratory's technical and non-technical functions at all location/sites where testing is performed. QA also extends to the laboratory's interactions with and responsibilities to patients, physicians, and other laboratories ordering tests, and the non-laboratory areas or the facility of which it is a part.

When the laboratory discovers an error or identifies a potential problem, actions must be taken to correct the situation. This correction process involves identification and resolution of the problem, and development of policies that will prevent recurrence. Policies for preventing problems that have been identified must be written as well as communicated to the laboratory personnel and other staff, clients, etc., as appropriate. Over time, the laboratory must monitor the corrective action(s) to ensure the action(s) taken have prevented recurrence of the original problem.

All pertinent laboratory staff must be involved in the assessment process through discussions or active participation.

QA of the Analytic System includes assessing:

- o Test procedures;
- o Accurate and reliable test systems, equipment, instruments, reagents, materials, and supplies;
- o Specimen and reagent storage condition;
- o Equipment/instrument/test/system maintenance and function checks;
- o Establishment and verification of method performance specifications;
- o Calibration and calibration verification;
- o Control procedures;
- o Comparison of test results;
- o Corrective actions; and
- o Test records.

For Clinical Cytogenetics, cases, the laboratory should identify increases in or excessive culture failure rates, determine the contributing factors, document efforts to reduce or eliminate these factors and assess the effectiveness of actions taken. (i.e., a decrease in the culture failure rate).

Review assessment policies, procedures and reports to verify that the laboratory has a system in place to ensure continuous improvement. Corrective action reports are one indication that the laboratory is monitoring and evaluating laboratory performance and the quality of services.

Select a sample of abnormal cytology patient reports and determine that, when available, the histopathology and

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cytology comparison was performed and the cytology 5-year retrospective review was performed. Ensure the laboratory documents any discrepancies and performs corrective action.

Review quality control records to determine if the laboratory's monitoring efforts are detecting control failures, shifts, and trends. If the surveyor identifies previously undetected quality control failures or omission, then the laboratory's system for monitoring and evaluating quality control may not be adequate.

Interpretative Guidelines §493.1289(c)

The steps taken by the laboratory to identify and correct problems and prevent their recurrence must be documented. All laboratory policies amended due to its QA activities must also be noted.

Probes §493.1289(a):

For clinical cytogenetics cases, does the laboratory monitor the frequency of culture failures and sub-optimal analyses?

Does the laboratory add additional maintenance procedures and/or function checks, when needed, to ensure accurate and reliable test results?

What is the laboratory's system for monitoring and evaluating test results for inconsistencies with patient information?

**FED - D5793 - ANALYTIC SYSTEMS QUALITY ASSESSMENT**

**Title** ANALYTIC SYSTEMS QUALITY ASSESSMENT

**CFR** 493.1289(b)(c)

**Type** Standard

**Regulation Definition**

- (b) The analytic systems quality assessment must include a review of the effectiveness of corrective actions taken to resolve problems, revision of policies and procedures necessary to prevent recurrence of problems, and discussion of analytic systems quality assessment reviews with appropriate staff.
- (c) The laboratory must document all analytic systems assessment activities.

**Interpretive Guideline**

Interpretative Guidelines §493.1289(b):

Verify that the laboratory has a system in place to monitor and evaluate test results for inconsistencies with patient information, and for correlation between test results. For example, a laboratory could multiply the hemoglobin result by a factor of 3, to see if the result is equal to the hematocrit. If the laboratory has auto-validation in its Laboratory Information System (LIS), verify that the laboratory is taking steps to reduce the likelihood of sample-switching errors, for example, when the creatinine result is significantly different from the patient's previous creatinine test results, or if the MCV is significantly different from the patient's previous test results and the patient did not receive a blood transfusion.

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Interpretative Guidelines §493.1289(c):

The steps taken by the laboratory to identify and correct problems and prevent their recurrence must be documented. All laboratory policies amended due to its QA activities must also be noted.

Probes §493.1289(b):

How does the laboratory address multiple failed or sub-optimal cultures that have been submitted from one client?

How does the laboratory use the review of all normal or negative gynecologic specimens received within the previous 5 years to assess the analytic system and communicate findings to the staff?

**FED - D5800 - POSTANALYTIC SYSTEMS**

**Title** POSTANALYTIC SYSTEMS

**CFR** 493.1290

**Type** Condition

**Regulation Definition**

Each laboratory that performs nonwaived testing must meet the applicable postanalytic systems requirements in §493.1291 unless HHS approves a procedure, specified in Appendix C of the State Operations Manual (CMS Pub. 7) that provides equivalent quality testing. The laboratory must monitor and evaluate the overall quality of the postanalytic systems and correct identified problems as specified in §493.1299 for each specialty and subspecialty of testing performed.

**Interpretive Guideline**

Interpretative Guidelines §493.1290:

Significant deficiencies cited under this condition may indicate deficiencies under personnel responsibilities. Use D5800 when deficiencies are identified that are: significant and have the potential to, or adversely affect, patient testing, are systemic and pervasive throughout the laboratory, and are not limited to any one specialty or subspecialty.

**FED - D5801 - TEST REPORT**

**Title** TEST REPORT

**CFR** 493.1291(a)

**Type** Standard

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**Regulation Definition**

The laboratory must have an adequate manual or electronic system(s) in place to ensure test results and other patient-specific data are accurately and reliably sent from the point of data entry (whether interfaced or entered manually) to final report destination, in a timely manner. This includes the following:

- (a)(1) Results reported from calculated data.
- (a)(2) Results and patient-specific data electronically reported to network or interfaced systems.
- (a)(3) Manually transcribed or electronically transmitted results and patient-specific information reported directly or upon receipt from outside referral laboratories, satellite or point-of-care testing locations.

**Interpretive Guideline**

Interpretative Guidelines §493.1291(a)

The regulations apply to manual as well as automated record systems (e.g., a laboratory information system or LIS). Regardless of the means used to transmit laboratory results, routine checks should be conducted to verify that transmissions are being accurately and reliably conveyed to the final report destination.

For CLIA purposes, the final report destination for test results is considered to be the authorized person and/or their designated personal representative (a personal representative is generally a person authorized under applicable law to make health care decisions for the individual. See 45 CFR §164.502(g). Additional individuals or entity(s) who are responsible for using the test results may also receive test results from the laboratory if they are designated by the authorized person on the test requisition. As of April 7, 2014 a new CLIA regulation was added at §493.1291(l) in order to provide patients with more access to laboratory test report(s). In accordance with amendments to the HIPAA Privacy Rule, the new regulation states: "Upon request by a patient (or the patient's personal representative), the laboratory may provide patients, their personal representatives, and those persons specified under 45 CFR 164.524(c) (3)(ii), as applicable, with access to completed test reports that, using the laboratory's authentication process, can be identified as belonging to that patient. The HIPAA Privacy Rule preempts contrary state laws on patient access to laboratory test report(s), but where a HIPAA-covered laboratory can continue to comply with both the HIPAA Privacy Rule and state law, it must frame its policies and procedures in a way that complies with both laws. Further, the HIPAA Privacy Rule does not preempt more stringent state laws, even if contrary to the Privacy Rule. CLIA laboratories that are not subject to HIPAA will have discretion to provide patients with direct access to their laboratory test reports, subject to any applicable state laws that may constrain access.

To ensure the accurate, timely, confidential, and easily understood reporting of patient test results to the authorized person, their personal representative (if applicable) and others who are identified as responsible for using the test results on the requisition, a laboratory may contract with another entity to assist in the delivery of patient reports in a manner that complies with all applicable laws, including the CLIA regulatory and statutory requirements. Please note that if the laboratory is subject to HIPAA and the entity with which it contracts meets the HIPAA definition of a business associate, see 45 CFR §160.103 (definition of "business associate"), the laboratory's contract or other written arrangement with its business associate must contain the elements specified at 45 CFR §164.504(e).

Note: An example of an electronic system that a laboratory or health care provider can contract with is Direct, which provides secure, authenticated, encrypted transport of laboratory test results to an authorized person, their personal representative, and others responsible for using the test results. Laboratories utilizing Direct, in addition to fully supporting the Direct Implementation Guide for Delivery Notification, and meeting all other relevant CLIA requirements, would meet the CLIA regulations for an adequate electronic system for sending test results to the final

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report destination (§493.1291(a)).  
Probes §493.1291(a)

How does the laboratory ensure that transmitted reports are legible and the information received at the final destination was the same data sent by the laboratory?

If the laboratory uses a LIS or facsimile, what security measures have been instituted to ensure that transmitted reports go directly from the device sending reports to the authorized person, their personal representative (if applicable), and others who are identified as responsible for using the test results on the requisition?

Interpretative Guidelines §493.1291(a)(3)

Manually transcribed or electronically transmitted results from an outside referral laboratory or from within the laboratory system (e.g., satellite or point-of-care testing locations) must be periodically verified for accuracy and timely reporting.

**FED - D5803 - TEST REPORT**

**Title** TEST REPORT

**CFR** 493.1291(b)

**Type** Standard

**Regulation Definition**

Test report information maintained as part of the patient's chart or medical record must be readily available to the laboratory and to CMS or a CMS agent upon request.

**Interpretive Guideline**

Interpretative Guidelines §493.1291(b):  
The test report information should be legible, understandable, and complete.

**FED - D5805 - TEST REPORT**

**Title** TEST REPORT

**CFR** 493.1291(c)

**Type** Standard

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**Regulation Definition**

The test report must indicate the following:

- (c)(1) For positive patient identification, either the patient's name and identification number, or a unique patient identifier and identification number.
- (c)(2) The name and address of the laboratory location where the test was performed.
- (c)(3) The test report date.
- (c)(4) The test performed.
- (c)(5) Specimen source, when appropriate.
- (c)(6) The test result and, if applicable, the units of measurement or interpretation, or both.
- (c)(7) Any information regarding the condition and disposition of specimens that do not meet the laboratory's criteria for acceptability.

**Interpretive Guideline**

Interpretative Guidelines §493.1291(c)(1)- (c)(6):

Use D5203 for deficiencies related to specimen identification problems.

When used on the test report, the patient's name must be accompanied by an identification or accession number. When for confidentiality purposes a patient's name is not used or when the identity of the person is not known, a unique patient identifier and identification or accession number must be used on the report.

Interpretative Guidelines §493.1291(c)(2)

Laboratories having a single certificate for multiple sites/locations must have a system in place to identify which tests were performed at each site. When testing is performed in more than one location in a hospital, the specific location in the hospital must be stated on the laboratory report (for example, ER, NICU, etc.)

A code to identify the name and address of the laboratory performing testing is acceptable as long as the code is clearly annotated on the patient test report. This may be accomplished by using abbreviated indicators (e.g., asterisks) as long as they are identified and apparent to the individual receiving the report. This or a similar system may be seen on cumulative reports. The name and address of the reference laboratory may also be defined on a subsequent page or on the back of the report. Laboratories have latitude to develop other formats to meet this requirement.

Interpretative Guidelines §493.1291(c)(3):

The date of the test report is the date results were generated as a final report and must not change on copies generated at a later date.

Interpretative Guidelines §493.1291(c)(4)

For tests that have not been FDA-cleared or approved (including test systems not subject to FDA clearance or approval, methods developed in-house, standardized methods such as textbook procedures, and FDA-cleared or approved test systems modified by the laboratory), the test report must include the statement "The performance characteristics of this test were determined by (Laboratory Name). It has not been cleared or approved by the U.S. Food and Drug Administration".

The disclaimer for Analyte Specific Reagents (ASR) should state ["This test was developed and its performance characteristics determined by (Laboratory Name). It has not been cleared or approved by the U.S. Food and Drug Administration"]. The ASR disclaimer on the test report is required by the FDA under 21 CFR, Part 809.30(

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<<http://www.accessdata.fda.gov/scripts/cdrh/cfdocs/cfcfr/CFRSearch.cfm?fr=809.30>>e) "Restrictions on the sale, distribution and use of analyte specific reagents."

In either case, the laboratory must establish performance specifications in accordance with §493.1253(b)(2), and must make them available to clients in accordance with §493.1291(e).

Interpretative Guidelines §493.1291(c)(5):

Some examples of source of the specimen needed by the laboratory to accurately perform testing and report results would be: site of culture; type of body fluid; whether a submitted separated specimen is plasma, serum, urine, etc.

Interpretative Guidelines §493.1291(c)(6):

If the laboratory prints normal ranges on the patient test report, verify that "sex and/or age specific" normal ranges are printed by the LIS on the patient test report.

"Less than" is used for reporting test results (qualitative or quantitative) that are below the laboratory's detection limits for an analyte. (Detection limits must be established through method verification as described in §493.1253.) "Equivalent designation" is used to report test results for those methods that yield results below a clinically significant level (e.g., for a quantitative immunology test, patient results may be clinically negative at a 1:8 titer and test results may be reported as "1:8 negative". The normal range is 1:8 or less.) "Greater than" is used for reporting test results (qualitative or quantitative) that are above the laboratory's detection limits for an analyte. If patient test results exceed the laboratory's reportable range, the laboratory must report the result as greater than the highest detection limit, reassay a diluted patient specimen and report the calculated result, or send the specimen to a reference laboratory.

For flow cytometry, to interpret results, staff should have access to the complementary clinical picture of the patient. This may include such results as white cell count, cell differential, cell morphology, and cytogenetics.

Flow cytometry patient data files should include any gating analysis regions used to obtain reported test results.

For genetic tests, the laboratory should include the test method(s) employed and any mutations on the test report.

For DNA or nucleic acid based genetic tests, the laboratory should include the test method(s) employed and mutation(s) detected on the test report.

Interpretative Guidelines §493.1291(c)(7):

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If the laboratory functions as a reference laboratory, how does it notify the referring laboratory or client of unacceptable specimens in a timely manner? Use D5801 to cite timeliness deficiencies. Use D5805 to cite the referring laboratory's failure to notify the appropriate individual concerning the unacceptable specimen.

Probes §493.1291(c)(6):

When additional information is critical for the interpretation of test results (e.g., screening vs. confirmatory procedures), how does the laboratory convey this information to the individual ordering or using test results?

If the laboratory does not print normal ranges on the test report, how does the laboratory notify the client that reported results are abnormal for the patient due to their particular sex and/or age?

**FED - D5807 - TEST REPORT**

**Title** TEST REPORT

**CFR** 493.1291(d)

**Type** Standard

**Regulation Definition**

Pertinent "reference intervals" or "normal" values, as determined by the laboratory performing the tests, must be available to the authorized person who ordered the tests and, if applicable, the individual responsible for using the test results.

**Interpretive Guideline**

Interpretative Guidelines §493.1291(d):

The laboratory must ensure the "reference intervals" or "normal" values it provides to its clients are accurate, include appropriate units of measurement, and reflect the method performed and the patient population (if applicable).

**FED - D5809 - TEST REPORT**

**Title** TEST REPORT

**CFR** 493.1291(e)

**Type** Standard

**Regulation Definition**

The laboratory must, upon request, make available to clients a list of test methods employed by the laboratory and, as applicable, the performance specifications established or verified as specified in §493.1253. In addition, information

**Interpretive Guideline**

Interpretative Guidelines §493.1291(e)

When the laboratory changes methods, establishes a new procedure or refers tests to another laboratory, the laboratory must make the updated information concerning parameters such as patient preparation, preservation of

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that may affect the interpretation of test results, for example test interferences, must be provided upon request. Pertinent updates on testing information must be provided to clients whenever changes occur that affect the test results or interpretation of test results.

specimens, specimen collection, or new "normal" ranges or units of measure available to its clients.

§493.1291(e) Probes

How does the laboratory keep its clients informed about tests offered, methods used, and specimen requirements?

What means does the laboratory use to provide interpretation of results to its clients?

**FED - D5811 - TEST REPORT**

**Title** TEST REPORT

**CFR** 493.1291(f)

**Type** Standard

**Regulation Definition**

(f) Except as provided in §493.1291(l), test results must be released only to authorized persons and, if applicable, the persons responsible for using the test results and the laboratory that initially requested the test.

**Interpretive Guideline**

Interpretative Guidance §493.1291(f)

Test results must be released to the authorized person and, if the authorized person is a patient, the patient's personal representatives and those persons specified under 45 CFR 164.524(c)(3)(ii), as applicable. If the authorized person is not a patient, test results must be released to the authorized person, and, if applicable, the persons responsible for using the test results and the laboratory that initially requested the test. Test results must also be released to any additional individuals/entities designated on the test requisition. These entities are understood to be "responsible for using" the test results.

When the authorized person, and, if applicable, the individual responsible for using the test results receives the results, the laboratory's CLIA responsibility ends. When a reference laboratory receives a specimen from another referring laboratory, the referring laboratory is responsible for getting the results back to the authorized person and, if applicable, any individuals responsible for using the results.

See D5301 for the definition of an "authorized person".

Probes §493.1291(f)

What security measures have been instituted to ensure that reports go directly from the device sending reports (e.g., LIS, facsimile) to the authorized person and: (i) if the authorized person is a patient, the patient's personal representatives and those persons specified under 45 CFR 164.524(c)(3)(ii), as applicable; and (ii) if the authorized person is not a patient, the persons who are identified as responsible for using the test results and the laboratory that initially requested the test, as applicable?

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**FED - D5813 - TEST REPORT**

**Title** TEST REPORT

**CFR** 493.1291(g)

**Type** Standard

**Regulation Definition**

The laboratory must immediately alert the individual or entity requesting the test and, if applicable, the individual responsible for using the test results when any test result indicates an imminently life-threatening condition, or panic or alert values.

**Interpretive Guideline**

Interpretative Guidelines §493.1291(g)

The laboratory records should document the date, time, test results, and person to whom the test results were reported.

See D5301 for the definition of an "authorized person".

Probes §493.1291(g)

What means does the laboratory use to ensure the authorized person is alerted in a timely manner to critical, alert, or panic test results?

**FED - D5815 - TEST REPORT**

**Title** TEST REPORT

**CFR** 493.1291(h)

**Type** Standard

**Regulation Definition**

When the laboratory cannot report patient test results within its established time frames, the laboratory must determine, based on the urgency of the patient test(s) requested, the need to notify the appropriate individual(s) of the delayed testing.

**Interpretive Guideline**

Interpretative Guidelines §493.1291(h):

If a delay in reporting patient test results may negatively impact patient care, the laboratory should have an alternative method for reporting patient results when the LIS or test system is down.

Cite deficiencies only when the laboratory has failed to notify its client(s) when delays in testing patient specimens have the potential for or are adversely affecting patient care.

Probes §493.1291(h):

What criteria has the laboratory established for notifying the appropriate individual of the delay in testing? Use D5403.

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How will the laboratory report patient test results if the LIS or test system is down?

**FED - D5817 - TEST REPORT**

**Title** TEST REPORT

**CFR** 493.1291(i)

**Type** Standard

**Regulation Definition**

If a laboratory refers patient specimens for testing--

(i)(1) The referring laboratory must not revise results or information directly related to the interpretation of results provided by the testing laboratory;

(i)(2) The referring laboratory may permit each testing laboratory to send the test result directly to the authorized person who initially requested the test. The referring laboratory must retain or be able to produce an exact duplicate of each testing laboratory's report; and

(i)(3) The authorized person who orders a test must be notified by the referring laboratory of the name and address of each laboratory location where the test was performed.

**Interpretive Guideline**

Interpretative Guidelines §493.1291(i)(1)

If the laboratory transcribes results from the reference laboratory report, the test results, interpretation and information directly related to the interpretation must be copied exactly as reported by the reference laboratory. The report must adhere to the requirements in §§493.1291(c)(1)-(c)(7) and 493.1291(d).

Interpretative Guidelines §493.1291(i)(2)

An "exact duplicate" is an exact copy of the information sent to the individual requesting the test or using the test result(s), and includes the name and address of the laboratory performing the test. The exact copy need not be paper, it may be retrieved from a computer system, microfilm or microfiche record, as long as it contains the exact information as sent to the individual ordering the test or utilizing the test results. The duplicate laboratory report must contain information positioned such that it is clear and includes all original interpretative information. For tests requiring an authorized signature or containing personnel identifiers (e.g., Pathology), the exact duplicate must include the signatures or identifiers. "Pathology" includes all of its subspecialties (i.e., Histopathology, Oral pathology, Cytology).

A "preliminary report" means a test result that has been reported to the authorized person or laboratory that initially requested the test before the final test result is completed. Frequently, a preliminary report will contain significant, but not definitive information (e.g., a urine culture preliminary report of >100,000 Gram-negative bacilli after 24 hours incubation or a beta subunit preliminary report of > 200 miu/ml). It should be noted on the report when the result is a preliminary result and that a final report will follow.

A "partial report" means multiple tests are ordered on the same specimen or patient. If partial reports are issued for only those tests that have been completed, then the report date will be the date when all tests have been completed. However, the laboratory should be able to identify the date that each new test is appended to the report.

The laboratory must have a system for retaining copies of all reports including original, preliminary, corrected, and

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final reports. This includes computer-generated reports.

Interpretative Guidelines §493.1291(i)(3)

Test report forms may include codes to identify the name and address of the laboratory that performed the test, provided the interpretations of the codes are available to the authorized person using the test results.

**FED - D5819 - TEST REPORT**

**Title** TEST REPORT

**CFR** 493.1291(j)

**Type** Standard

**Regulation Definition**

All test reports or records of the information on the test reports must be maintained by the laboratory in a manner that permits ready identification and timely accessibility.

**Interpretive Guideline**

Interpretative Guidelines §493.1291(j)

The regulations do not specify the mechanism or frequency for which a laboratory should evaluate its record storage and retrieval system.

**FED - D5821 - TEST REPORT**

**Title** TEST REPORT

**CFR** 493.1291(k)

**Type** Standard

**Regulation Definition**

When errors in the reported patient test results are detected, the laboratory must do the following:

(k)(1) Promptly notify the authorized person ordering the test and, if applicable, the individual using the test results of reporting errors.

(k)(2) Issue corrected reports promptly to the authorized person ordering the test and, if applicable, the individual using the test results.

(k)(3) Maintain duplicates of the original report, as well as the corrected report.

**Interpretive Guideline**

Interpretative Guidelines §493.1291(k)

Errors in test results may include incorrect patient identification, test results, reference or normal ranges, interpretative information, or other significant information. See D5625 for specific guidance regarding certain amended cytology reports.

Interpretative Guidelines §493.1291(k)(1)

When determining whether the laboratory gave prompt notification of test and/or reporting errors to the authorized person(s), their agent (if applicable), and others who are identified as responsible for using the test results on the requisition, consider whether contact information was provided to the laboratory, when the error was identified, when the authorized person was notified, and the extent of the error (e.g., clinically significant results reported on the

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wrong patient).

Interpretative Guidelines §493.1291(k)(2)

Corrected reports, either hard copy or electronic, must clearly indicate both the corrected results(s) and the fact that the report is a corrected report. The corrected reports should be promptly sent to the authorized person, their agent (if applicable) and others who are identified as responsible for using the test results on the requisition.

For corrected reports in Cytology, use D5659.

Interpretative Guidelines §493.1291(k)(3)

The laboratory must have a system for maintaining copies of the original and corrected reports. Computer-generated reports or electronically stored copies are acceptable.

Copies of all reports, including corrected reports, provided by the referral laboratory must be maintained by both the referral and referring laboratories for the required time periods.

Probes §493.1291(k)(1)

What mechanism(s) does the laboratory use for notifying the authorized person(s) of the corrected values?

Probes §493.1291(k)(2)

How does the laboratory ensure that incorrect original results are not reissued verbally, in writing or electronically?

Probes §493.1291(k)(3)

For laboratories that maintain the patient's medical record as the test report, what is the mechanism for differentiating between the incorrect original report and the corrected report?

**FED - D5823 - TEST REPORTS**

**Title** TEST REPORTS

**CFR** 493.1291(l)

**Type** Standard

**Regulation Definition**

Upon request by a patient (or the patient's personal representative), the laboratory may provide patients, their personal representatives, and those persons specified under 45

**Interpretive Guideline**

Interpretative Guidance §493.1291(l)

The laboratory must have and follow a written policy that is available to the laboratory staff and details how it handles patient requests for access to their completed laboratory reports. Test reports are considered to be complete

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CFR 164.524(c)(3)(ii), as applicable, with access to completed test reports that, using the laboratory's authentication process, can be identified as belonging to that patient.

when all results associated with the ordered tests are finalized and ready for release.

**FED - D5891 - POSTANALYTIC SYSTEMS QUALITY ASSESSMENT**

**Title** POSTANALYTIC SYSTEMS QUALITY ASSESSMENT

**CFR** 493.1299(a)

**Type** Standard

**Regulation Definition**

The laboratory must establish and follow written policies and procedures for an ongoing mechanism to monitor, assess and, when indicated, correct problems identified in the postanalytic systems specified in §493.1291.

**Interpretive Guideline**

Interpretative Guidelines §493.1299(a)-(c)

Quality Assessment (QA) is an ongoing review process that encompasses all facets of the laboratory's technical and non-technical functions and all locations/sites where testing is performed. QA also extends to the laboratory's interactions with and responsibilities to patients, physicians, and other laboratories ordering tests, and non-laboratory areas or departments of the facility of which it is a part.

When the laboratory discovers an error or identifies a potential problem, actions must be taken to correct the situation. This correction process involves investigation, identification and resolution of the problem, and development of policies that will prevent recurrence. Policies for preventing problems that have been identified must be written as well as communicated to the laboratory personnel and other staff, clients, etc., as appropriate. Over time, the laboratory must monitor the corrective action(s) to ensure the action(s) taken has prevented recurrence of the original problem.

All pertinent laboratory staff must be involved in the assessment process through discussions or active participation.

QA of the Postanalytic System includes assessing practices/issues related to test reports. Examples include monitoring and evaluating the accuracy and completeness of the laboratory's test reports (i.e., patient information, test results, normal ranges, and the disposition of unacceptable specimens), and the laboratory's turn-around times and procedures for notification of test results e.g., routine tests, STATS, abnormal or panic values.

Review a cross-section of patient test reports for accuracy of patient information, test results and normal ranges to verify that the laboratory is effectively monitoring and evaluating the quality and accuracy of the information supplied to its clients.

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Verify that the laboratory has a system in place to monitor and evaluate its established reporting time frames and procedures for notification of test results, routine tests, STATS, abnormal or panic values.

If the laboratory uses an LIS, the laboratory must have a mechanism to periodically verify the accuracy of:

- o Its calculated data;
- o Its results sent to interfaced systems; and
- o Patient specific data.

In the event that the laboratory becomes aware of information that reasonably suggests that an in vitro diagnostic device may have caused or contributed to a patient death or serious injury, verify that the laboratory has reported such instances to the FDA. Reports must be submitted on FDA Form 3500A (<<http://www.fda.gov/medwatch/getforms.htm>>) or an electronic equivalent as soon as practical, but no later than 10 days from the time personnel become aware of the event. For more information on reporting requirements, contact the FDA: Office of In Vitro Diagnostic Device Evaluation and Safety, Center for Devices and Radiological Health, Food and Drug Administration, HFZ-440, 2098 Gaither Road, Rockville, MD 20850, Phone: 240-276-0450, Fax: 240-276-0652.

**FED - D5893 - POSTANALYTIC SYSTEMS QUALITY ASSESSMENT**

**Title** POSTANALYTIC SYSTEMS QUALITY ASSESSMENT

**CFR** 493.1299(b)(c)

**Type** Standard

**Regulation Definition**

(b) The postanalytic systems quality assessment must include a review of the effectiveness of corrective actions taken to resolve problems, revision of policies and procedures necessary to prevent recurrence of problems, and discussion of postanalytic systems quality assessment reviews with appropriate staff.

(c) The laboratory must document all postanalytic systems quality assessment activities.

**Interpretive Guideline**

Interpretative Guidelines §493.1299(a)-(c)

Quality Assessment (QA) is an ongoing review process that encompasses all facets of the laboratory's technical and non-technical functions and all locations/sites where testing is performed. QA also extends to the laboratory's interactions with and responsibilities to patients, physicians, and other laboratories ordering tests, and non-laboratory areas or departments of the facility of which it is a part.

When the laboratory discovers an error or identifies a potential problem, actions must be taken to correct the situation. This correction process involves investigation, identification and resolution of the problem, and

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development of policies that will prevent recurrence. Policies for preventing problems that have been identified must be written as well as communicated to the laboratory personnel and other staff, clients, etc., as appropriate. Over time, the laboratory must monitor the corrective action(s) to ensure the action(s) taken has prevented recurrence of the original problem.

All pertinent laboratory staff must be involved in the assessment process through discussions or active participation.

QA of the Postanalytic System includes assessing practices/issues related to test reports. Examples include monitoring and evaluating the accuracy and completeness of the laboratory's test reports (i.e., patient information, test results, normal ranges, and the disposition of unacceptable specimens), and the laboratory's turn-around times and procedures for notification of test results e.g., routine tests, STATS, abnormal or panic values.

Review a cross-section of patient test reports for accuracy of patient information, test results and normal ranges to verify that the laboratory is effectively monitoring and evaluating the quality and accuracy of the information supplied to its clients.

Verify that the laboratory has a system in place to monitor and evaluate its established reporting time frames and procedures for notification of test results, routine tests, STATS, abnormal or panic values.

If the laboratory uses an LIS, the laboratory must have a mechanism to periodically verify the accuracy of:

- o Its calculated data;
- o Its results sent to interfaced systems; and
- o Patient specific data.

In the event that the laboratory becomes aware of information that reasonably suggests that an in vitro diagnostic device may have caused or contributed to a patient death or serious injury, verify that the laboratory has reported such instances to the FDA. Reports must be submitted on FDA Form 3500A (<<http://www.fda.gov/medwatch/getforms.htm>>) or an electronic equivalent as soon as practical, but no later than 10 days from the time personnel become aware of the event. For more information on reporting requirements, contact the FDA: Office of In Vitro Diagnostic Device Evaluation and Safety, Center for Devices and Radiological Health, Food and Drug Administration, HFZ-440, 2098 Gaither Road, Rockville, MD 20850, Phone: 240-276-0450, Fax: 240-276-0652.

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Interpretative Guidelines §493.1299(b):

Review assessment policies, procedures and reports to verify that the laboratory has a system in place to ensure continuous improvement. Corrective action reports are one indication that the laboratory is monitoring and evaluating laboratory performance and the quality of services.

Interpretative Guidelines §493.1299(c):

The steps taken by the laboratory to identify and correct problems, and prevent their recurrence must be documented. All laboratory policies amended due to its QA activities must be noted.

Probes §493.1299(a)-(c):

What mechanism does the laboratory use to update and correlate the information to clients (e.g., client reference manuals), procedure manuals, reporting systems (e.g., LIS) when the laboratory introduces a new test system with different normal/reference range?

**FED - D5980 - PPM LABORATORY DIRECTOR**

**Title** PPM LABORATORY DIRECTOR

**CFR** 493.1355

**Type** Condition

**Regulation Definition**

The laboratory must have a director who meets the qualification requirements of §493.1357 and provides overall management and direction in accordance with §493.1359.

**Interpretive Guideline**

**FED - D5981 - PPM LABORATORY DIRECTOR QUALIFICATIONS**

**Title** PPM LABORATORY DIRECTOR QUALIFICATIONS

**CFR** 493.1357

**Type** Standard

**Regulation Definition**

The laboratory director must be qualified to manage and direct

**Interpretive Guideline**

Interpretative Guidelines §493.1357(b)(2)

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the laboratory personnel and the performance of PPM procedures as specified in §493.19(c) and must be eligible to be an operator of a laboratory within the requirements of subpart R of this part.

(a) The laboratory director must possess a current license as a laboratory director issued by the State in which the laboratory is located, if the licensing is required.

(b) The laboratory director must meet one of the following requirements:

(b)(1) Be a physician, as defined in §493.2.

(b)(2) Be a midlevel practitioner, as defined in §493.2, authorized by a State to practice independently in the State in which the laboratory is located.

(b)(3) Be a dentist, as defined in §493.2.

Midlevel practitioner means a nurse midwife, nurse practitioner, or physician's assistant licensed by the State within which the individual practices, if such licensing is required in the State in which the laboratory is located.

**FED - D5983 - PPM LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** PPM LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1359

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the prompt, accurate, and proficient reporting of test results.

**Interpretive Guideline**

**FED - D5985 - PPM LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** PPM LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1359(a)

**Type** Standard

**Regulation Definition**

The laboratory director must--

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(a) Direct no more than five laboratories.

**FED - D5987 - PPM LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** PPM LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1359(b)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

The laboratory director must--

(b) Ensure that any procedure listed under §493.19(c)--

(b)(1) is personally performed by an individual who meets the qualification requirements in §493.1363; and

(b)(2) Is performed in accordance with applicable requirements in subparts H, J, K, and M of this part.

**FED - D5990 - PPM TESTING PERSONNEL**

**Title** PPM TESTING PERSONNEL

**CFR** 493.1361

**Type** Condition

**Regulation Definition**

**Interpretive Guideline**

The laboratory must have a sufficient number of individuals

who meet the qualification requirements of

§493.1363 to perform the functions specified in §493.1365 for the volume and complexity of testing performed.

**FED - D5991 - PPM TESTING PERSONNEL QUALIFICATIONS**

**Title** PPM TESTING PERSONNEL QUALIFICATIONS

**CFR** 493.1363

**Type** Standard

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**Regulation Definition**

Each individual performing PPM procedures must--

- (a) Possess a current license issued by the State in which the laboratory is located if the licensing is required; and
- (b) Meet one of the following requirements:
  - (b)(1) Be a physician, as defined in §493.2.
  - (b)(2) Be a midlevel practitioner, as defined in §493.2, under the supervision of a physician or in independent practice if authorized by the State in which the laboratory is located.
  - (b)(3) Be a dentist as defined in §493.2 of this part.

**Interpretive Guideline**

**FED - D5993 - PPM TESTING PERSONNEL RESPONSIBILITIES**

**Title** PPM TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1365(a)

**Type** Standard

**Regulation Definition**

The testing personnel are responsible for specimen processing, test performance, and for reporting test results. Any PPM procedure must be--

- (a) Personally performed by one of the following practitioners:
  - (a)(1) A physician during the patient's visit on a specimen obtained from his or her own patient or from a patient of a group medical practice of which the physician is a member or employee.
  - (a)(2) A midlevel practitioner, under the supervision of a physician or in independent practice if authorized by the State in which the laboratory is located, during the patient's visit on a specimen obtained from his or her own patient or from the patient of a clinic, group medical practice, or other health care provider, in which the midlevel practitioner is a member or an employee.
  - (a)(3) A dentist during the patient's visit on a specimen

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obtained from his or her own patient or from a patient of a group dental practice of which the dentist is a member or an employee

**FED - D5995 - PPM TESTING PERSONNEL RESPONSIBILITIES**

**Title** PPM TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1365(b)

**Type** Standard

**Regulation Definition**

The testing personnel are responsible for specimen processing, test performance, and for reporting test results. Any PPM procedure must be--  
(b) Performed using a microscope limited to a brightfield or a phase/contrast microscope.

**Interpretive Guideline**

**FED - D6000 - MODERATE COMPLEXITY LABORATORY DIRECTOR**

**Title** MODERATE COMPLEXITY LABORATORY DIRECTOR

**CFR** 493.1403

**Type** Condition

**Regulation Definition**

The laboratory must have a director who meets the qualification requirements of §493.1405 of this subpart and provides overall management and direction in accordance with §493.1407 of this subpart.

**Interpretive Guideline**

Interpretative Guidelines §493.1403:

The Condition: laboratory director is not met when the laboratory director:

- o position is not filled;
- o is not qualified; or
- o does not fulfill the laboratory director's responsibilities.

An individual qualified as laboratory director may not qualify as a technical consultant in a particular specialty or subspecialty unless he or she has the required testing experience.

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**FED - D6003 - LABORATORY DIRECTOR QUALIFICATIONS**

**Title** LABORATORY DIRECTOR QUALIFICATIONS

**CFR** 493.1405 AND 493.1406

**Type** Standard

**Regulation Definition**

The laboratory director must be qualified to manage and direct the laboratory personnel and the performance of moderate complexity tests and must be eligible to be an operator of a laboratory within the requirements of subpart R of this part.

(a) The laboratory director must possess a current license as a laboratory director issued by the State in which the laboratory is located, if such licensing is required; and

(b) The laboratory director must--

(b)(1)(i) Be a doctor of medicine or doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located; and

(b)(1)(ii) Be certified in anatomic or clinical pathology, or both, by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification; or

(b)(2)(i) Be a doctor of medicine, doctor of osteopathy, or doctor of podiatric medicine licensed to practice medicine, osteopathy, or podiatry in the State in which the Laboratory is located; and

(b)(2)(ii) Have had laboratory training or experience consisting of:

(b)(2)(ii)(A) At least one year directing or supervising non-waived laboratory testing; or

(b)(2)(ii)(B) Beginning September 1, 1993, have at least 20 continuing medical education credit hours in laboratory practice commensurate with the director responsibilities defined in §493.1407; or

(b)(2)(ii)(C) Laboratory training equivalent to paragraph (b)

**Interpretive Guideline**

Interpretative Guidelines §493.1405

When qualifying a Laboratory Director, please refer to section 353(i)(3) of the PHS Act as amended by the TEST Act, which now states, "No person who has owned or operated a laboratory which has had its certificate revoked may, within 2 years of the revocation of the certificate, own or operate a laboratory for which a certificate has been issued under this section (see §493.1840), except that if the revocation occurs pursuant to paragraph (4) the Secretary may substitute intermediate sanctions under subsection (h) instead of the 2-year prohibition against ownership or operation which would otherwise apply under this paragraph. The certificate of a laboratory which has been excluded from participation under the Medicare program under title XVIII of the Social Security Act [42 U.S.C.A. S 1395 et seq.] because of actions relating to the quality of the laboratory shall be suspended for the period the laboratory is so excluded."

Interpretative Guidelines §493.1405(a)

The term "State" as used in this provision, includes the District of Columbia, the Commonwealth of Puerto Rico, the Commonwealth of Northern Mariana Islands, the Virgin Islands, Guam and American Samoa.

Interpretative Guidelines §493.1405(b)(1)(ii)

Board certified means the individual has completed all the designated board's requirements, including the examination. If the director is named in a current edition of "The Official American Board of Medical Specialties (ABMS) Directory of Board Certified Medical Specialists (published by ABMS by Elsevier, 11830 Westline Industrial Drive, St. Louis, Missouri 63146, 1-866-856-8075) as appropriately board certified, this may be accepted as evidence of certification without needing further documentation. You may make a notation of this in the laboratory's file.

Qualifications that are equivalent for certification include board eligibility (i.e., the individual meets all education, training or experience requirements to take the examination, but has not actually taken and successfully completed the examination.) An individual who wishes to qualify as a director must supply evidence of this eligibility status. The designated boards, upon request, send a letter to the individual confirming his/her eligibility status. Note that some boards set time restrictions for taking the examination. For purposes of the regulations, the individual must meet the

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(2)(ii)(B) of this section obtained during medical residency. (For example, physicians certified either in hematology or hematology and medical oncology by the American Board of Internal Medicine); or

(b)(3) Hold an earned doctoral degree in a chemical, physical, biological, or clinical laboratory science from an accredited institution; and

(b)(3)(i) Be certified by the American Board of Medical Microbiology, the American Board of Clinical Chemistry, the American Board of Bioanalysis, or the American Board of Medical Laboratory Immunology; or

(b)(3)(ii) Have had at least one year experience directing or supervising non-waived laboratory testing;

(b)(4)(i) Have earned a master's degree in a chemical, physical, biological or clinical laboratory science or medical technology from an accredited institution;

(b)(4)(ii) Have at least one year of laboratory training or experience, or both in non-waived testing; and

(b)(4)(iii) In addition, have at least one year of supervisory laboratory experience in non-waived testing; or

(b)(5)(i) Have earned a bachelor's degree in a chemical, physical, or biological science or medical technology from an accredited institution;

(b)(5)(ii) Have at least 2 years of laboratory training or experience, or both in non-waived testing; and

(b)(5)(iii) In addition, have at least 2 years of supervisory laboratory experience in non-waived testing;

(b)(6) Be serving as a laboratory director and must have previously qualified or could have qualified as a laboratory director under §493.1406; or

(b)(7) On or before February 28, 1992, qualified under State law to direct a laboratory in the State in which the laboratory is located.

Laboratory director qualifications on or before February 28,

education, training or experience required by the board to be eligible to take the examination and must have confirmation of eligibility status.

*Interpretative Guidelines §493.1405(b)(2)(i)*

Individuals who have earned a Doctor of Optometry are qualified to serve as a laboratory director of certain moderate complexity tests under CLIA, but only for test procedures performed in their specialty area. [Ref: S&C-05-44] Optometrists may perform tests that are FDA-approved or cleared, of waived or moderate test complexity with the specimen source of tears such as lactoferrin, adenovirus, IgE, and osmolality.

*Interpretative Guidelines §493.1405(b)(2)(ii)*

The type of experience required under this regulation is clinical in nature. This means directing or supervising personnel who examine and perform tests on human specimens for the purpose of providing information that is used in diagnosing, treating, and monitoring a patient's condition. This experience may include the laboratory director personally examining and performing tests on patient specimens. Patient or medically oriented experience, which is defined as the ordering of tests and interpreting and applying the results of these tests in diagnosing and treating a patient's illness, is unacceptable to meet the requirement for laboratory training or experience.

The laboratory director should have documentation, e.g., signed procedure manuals, test reports, worksheets and workcards, that indicates the director assumes the responsibilities in §493.1407.

Teaching experience directly related to a medical technology program, clinical laboratory sciences program, or a clinical laboratory section of a residency program is considered acceptable experience. Research experience is also acceptable experience if it is obtained while performing tests on human specimens.

Ophthalmologists with a doctor of medicine (MD) degree are qualified to direct moderate complexity laboratories, provided they have had at least one year of experience directing or supervising moderate complexity laboratories, or have obtained at least 20 CMEs in laboratory practice commensurate with the laboratory director's responsibilities in §493.1407. [Ref: S&C-05-44]

*Interpretative Guidelines §493.1405(b)(2)(ii)(B)*

The 20 CMEs must be obtained prior to qualifying as a laboratory director. The CME courses must encompass preanalytic, analytic, and postanalytic phases of testing, and be of such quality as to provide the physician with education equivalent to the experience described in §493.1405(b)(2)(ii)(A). Courses related to laboratory payment and CPT coding would not fulfill this requirement.

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The laboratory director must be qualified to manage and direct the laboratory personnel and test performance.

(a) The laboratory director must possess a current license as a laboratory director issued by the State, if such licensing exists; and

(b) The laboratory director must:

(b)(1) Be a physician certified in anatomical or clinical pathology (or both) by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification;

(b)(2) Be a physician who:

(b)(2)(i) Is certified by the American Board of Pathology or the American Osteopathic Board of Pathology in at least one of the laboratory specialties; or

(b)(2)(ii) Is certified by the American Board of Medical Microbiology, the American Board of Clinical Chemistry, the American Board of Bioanalysis, or other national accrediting board in one of the laboratory specialties; or

(b)(2)(iii) Is certified by the American Society of Cytology to practice cytopathology or possesses qualifications that are equivalent to those required for such certification; or

(b)(2)(iv) Subsequent to graduation, has had 4 or more years of full-time general laboratory training and experience of which at least 2 years were spent acquiring proficiency in one of the laboratory specialties;

(b)(3) For the subspecialty of oral pathology only, be certified by the American Board of Oral Pathology, American Board of Pathology or the American Osteopathic Board of Pathology or possesses qualifications that are equivalent to those required for certification;

(b)(4) Hold an earned doctoral degree from an accredited institution with a chemical, physical, or biological science as a major subject and

For a list of some CME providers, please see the CLIA web page at [www.cms.hhs.gov/clia](http://www.cms.hhs.gov/clia) <<http://www.cms.hhs.gov/clia>>. The list of courses on the CLIA web page is not all inclusive. Other courses may meet the criteria, but all courses must be accredited.

In evaluating the 20 CMEs, verify they include the laboratory director responsibilities detailed in §493.1407.

Interpretative Guidelines §493.1405(b)(2)(ii)(C)

The residency program should provide the director the knowledge in principles and theories of laboratory practice including: quality control and quality assessment, proficiency testing, the phases of the total process (i.e., preanalytic, analytic and postanalytic), as well as, general laboratory systems, facility administration, and development and implementation of personnel policy and procedure manuals. This training should also include hands-on laboratory testing.

Interpretative Guidelines §493.1405(b)(3)

See §493.2 for the definition of an accredited institution.

Interpretative Guidelines §493.1405(b)(6)

For tests of moderate complexity, individuals qualify as laboratory directors, if on February 28, 1992, they previously qualified, or could have qualified under the Federal regulations, published on March 14, 1990, as a laboratory director. After February 28, 1992, individuals must meet the requirements at §§493.1405(b)(1)-(5) to qualify as a laboratory director, unless the individual can demonstrate compliance with §493.1405(b)(6), (that is, on February 28, 1992, he or she could have qualified as a laboratory director under Federal regulations published on March 14, 1990).

§493.1406 Standard; Laboratory director qualifications on or before February 28, 1992

There are no Interpretative Guidelines for §493.1406.

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- (b)(4)(i) Is certified by the American Board of Medical Microbiology, the American Board of Clinical Chemistry, the American Board of Bioanalysis, or other national accrediting board acceptable to HHS in one of the laboratory specialties; or
- (b)(4)(ii) Subsequent to graduation, has had 4 or more years of full-time general laboratory training and experience of which at least 2 years were spent acquiring proficiency in one of the laboratory specialties;
- (b)(5) With respect to individuals first qualifying before July 1, 1971, have been responsible for the direction of a laboratory for 12 months between July 1, 1961, and January 1, 1968, and, in addition, either:
- (b)(5)(i) Was a physician and subsequent to graduation had at least 4 years of pertinent full-time laboratory experience;
- (b)(5)(ii) Held a master's degree from an accredited institution with a chemical, physical, or biological science as a major subject and subsequent to graduation had at least 4 years of pertinent full-time laboratory experience;
- (b)(5)(iii) Held a bachelor's degree from an accredited institution with a chemical, physical, or biological science as a major subject and subsequent to graduation had at least 6 years of pertinent full-time laboratory experience; or
- (b)(5)(iv) Achieved a satisfactory grade through an examination conducted by or under the sponsorship of the U.S. Public Health Service on or before July 1, 1970; or
- (b)(6) Qualify under State law to direct the laboratory in the State in which the laboratory is located.

Note: The January 1, 1968 date for meeting the 12 months' laboratory direction requirement in paragraph (b)(5) of this section may be extended 1 year for each year of full-time laboratory experience obtained before January 1, 1958 required by State law for a laboratory director license. An exception to the July 1, 1971 qualifying date in paragraph (b)

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(5) of this section was made provided that the individual requested qualification approval by October 21, 1975 and had been employed in a laboratory for at least 3 years of the 5 years preceding the date of submission of his qualifications.

**FED - D6004 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(a)(b)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

- (a) The laboratory director, if qualified, may perform the duties of the technical consultant, clinical consultant, and testing personnel, or delegate these responsibilities to personnel meeting the qualifications of 493.1409, 493.1415, and 493.1421, respectively.
- (b) If the laboratory director reapportions performance of his or her responsibilities, he or she remains responsible for ensuring that all duties are properly performed.

**Interpretive Guideline**

Interpretative Guidelines §493.1407

If the laboratory has more than one person qualifying as director, the laboratory is required to designate one individual who has ultimate responsibility for overall operation and administration of the laboratory.

The requirement that a laboratory must be under the direction of a qualified person is not automatically met simply because the director meets the education and experience requirements. It must be demonstrated that the individual is, in fact, providing effective direction over the operation of the laboratory.

In determining whether the director responsibilities are met, consider deficiencies found in other conditions, e.g., facility administration, general laboratory systems, preanalytic systems, analytic systems, postanalytic systems, and proficiency testing.

Interpretative Guidelines §493.1407(a)

If the laboratory director is not qualified as a technical consultant or clinical consultant, he or she must employ individuals meeting the appropriate qualifications.

**FED - D6005 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(c)

**Type** Standard

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**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(c) The laboratory director must be accessible to the laboratory to provide onsite, telephone or electronic consultation as needed.

**Interpretive Guideline**

Interpretative Guidelines §493.1407(c)

If the director cannot practically provide personal, on-site supervision it must be demonstrated that the director:

- o Provides direction and consultation by telephone or electronic means (e.g. email, text message or fax), as necessary; or
- o Delegates to qualified personnel specific responsibilities as provided in the regulations.

The laboratory director may delegate to a technical consultant, in writing, the responsibilities in: §§493.1407(e)(3), (4), (5), (6), (7), (11), (12), and (13).

The laboratory director may delegate to a clinical consultant, in writing, the responsibilities in: §§493.1407(e)(8) and (9).

**FED - D6006 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(d)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(d) Each individual may direct no more than five laboratories.

**Interpretive Guideline**

Interpretative Guidelines §493.1407(d)

An individual may serve as a director of 5 nonwaived certified laboratories. An individual may serve as a technical consultant or clinical consultant for any number of laboratories.

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**FED - D6007 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(1)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(E) The laboratory director must--

(E)(1) Ensure that testing systems developed and used for each of the tests performed in the laboratory provide quality laboratory services for all aspects of test performance, which includes the preanalytic, analytic, and postanalytic phases of testing;

**Interpretive Guideline**

**FED - D6010 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(2)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

**Interpretive Guideline**

Interpretative Guidelines §493.1407(e)(2)

OSHA/EPA issues cannot be cited using these requirements. If immediate jeopardy exists, the director should be informed immediately.

If you observe or obtain information regarding potential safety violations not applicable under CLIA, notify the appropriate State or local authority. Consult with the Regional Office (RO) for notification to other Federal agencies

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(e) The laboratory director must--  
(e)(2) Ensure that the physical plant and environmental conditions of the laboratory are appropriate for the testing performed.

such as the Occupational Safety and Health Administration (OSHA) [www.osha.gov], Environmental Protection Agency (EPA) [www.epa.gov], or Nuclear Regulatory Commission (NRC). The appropriate Federal, State or local authority, if warranted, will investigate and, if necessary, conduct an on-site visit.

**FED - D6011 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(2)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--  
(e)(2) and provide a safe environment in which employees are protected from physical, chemical, and biological hazards.

**Interpretive Guideline**

**FED - D6012 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(3)(i)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with

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the applicable regulations.

(e) The laboratory director must--

(e)(3) Ensure that--

(e)(3)(ii) The test methodologies selected have the capability of providing the quality of results required for patient care;

**FED - D6013 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(3)(ii)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(3) Ensure that--

(e)(3)(ii) Verification procedures used are adequate to determine the accuracy, precision, and other pertinent performance characteristics of the method;

**Interpretive Guideline**

**FED - D6014 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(3)(iii)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the

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employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(3) Ensure that--

(e)(3)(iii) Laboratory personnel are performing the test methods as required for accurate and reliable results.

**FED - D6015 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(4)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(4) Ensure that the laboratory is enrolled in an HHS approved proficiency testing program for the testing performed.

**Interpretive Guideline**

**FED - D6016 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(4)(i)

**Type** Standard

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**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(4)(i) Ensure that the proficiency testing samples are tested as required under Subpart H of this part;

**Interpretive Guideline**

**FED - D6017 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(4)(ii)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(4)(ii) Ensure that results are returned within the timeframes established by the proficiency testing program.

**Interpretive Guideline**

**FED - D6018 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(4)(iii)

**Type** Standard

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**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(4)(iii) Ensure that all proficiency testing reports received are reviewed by the appropriate staff to evaluate the laboratory's performance and to identify any problems that require corrective action;

**Interpretive Guideline**

**FED - D6019 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(4)(iv)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(4)(iv) Ensure that an approved corrective action plan is followed when any proficiency testing results are found to be unacceptable or unsatisfactory.

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**FED - D6020 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(5)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(5) Ensure that the quality control program is established and maintained to assure the quality of laboratory services provided.

**Interpretive Guideline**

**FED - D6021 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(5)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(5) Ensure that quality assessment programs are established

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and maintained to assure the quality of laboratory services provided.

**FED - D6022 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(5)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(5) Ensure that the quality control and quality assessment programs are established and maintained to identify failures in quality as they occur.

**Interpretive Guideline**

**FED - D6023 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(6)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

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- (e) The laboratory director must--
- (e)(6) Ensure the establishment and maintenance of acceptable levels of analytical performance for each test system;

**FED - D6024 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(7)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

- (e) The laboratory director must--
- (e)(7) Ensure that all necessary remedial actions are taken and documented whenever significant deviations from the laboratory's established performance specifications are identified,

**Interpretive Guideline**

**FED - D6025 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(7)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly,

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accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(7) Ensure that patient test results are reported only when the system is functioning properly.

**FED - D6026 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(8)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(8) Ensure that reports of test results include pertinent information required for interpretation.

**Interpretive Guideline**

**FED - D6027 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(9)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly,

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accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(9) Ensure that consultation is available to the laboratory's clients on matters relating to the quality of the test results reported and their interpretation concerning specific patient conditions;

**FED - D6028 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(10)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(10) Employ a sufficient number of laboratory personnel with the appropriate education and either experience or training to provide appropriate consultation, properly supervise and accurately perform tests and report test results in accordance with the personnel responsibilities described in this subpart;

**Interpretive Guideline**

**FED - D6029 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(11)

**Type** Standard

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**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(11) Ensure that prior to testing patients' specimens, all personnel have the appropriate education and experience, receive the appropriate training for the type and complexity of the services offered, and have demonstrated that they can perform all testing operations reliably to provide and report accurate results.

**Interpretive Guideline**

**FED - D6030 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(12)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(12) Ensure that policies and procedures are established for monitoring individuals who conduct preanalytical, analytical, and postanalytical phases of testing to assure that they are competent and maintain their competency to process specimens, perform test procedures and report test results

**Interpretive Guideline**

Interpretative Guidelines §493.1407(e)(12)

Personnel performing only preanalytic and postanalytic activities are not required to be listed on Form 209. Surveyors do not normally check for documented competency evaluation on these individuals. However, if you discover problems in the laboratory and you find that a factor in these problems is poor performance of incompetent staff, cite D6030 or D6103 (laboratory director).

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promptly and proficiently, and whenever necessary, identify needs for remedial training or continuing education to improve skills;

**FED - D6031 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(13)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with the applicable regulations.

(e) The laboratory director must--

(e)(13) Ensure that an approved procedure manual is available to all personnel responsible for any aspect of the testing process;

**Interpretive Guideline**

Interpretative Guidelines §493.1497(e)(13)

The laboratory director can delegate to the technical supervisor the responsibility of making the procedure manual available, but cannot delegate the responsibility for signing new and revised procedures.

**FED - D6032 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1407(e)(14)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, and record and report test results promptly, accurate, and proficiently and for assuring compliance with

**Interpretive Guideline**

Interpretative Guidelines §493.1407(e)(14)

The director must assign, in writing, the duties/responsibilities to each person involved in all phases of the testing process. The list of assigned duties must be current.

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the applicable regulations.

(e) The laboratory director must--

(e)(14) Specify, in writing, the responsibilities and duties of each consultant and each person, engaged in the performance of the preanalytic, analytic, and postanalytic phases of testing, that identifies which examinations and procedures each individual is authorized to perform, whether supervision is required for specimen processing, test performance or results reporting, and whether consultant or director review is required prior to reporting patient test results.

**FED - D6033 - TECHNICAL CONSULTANT-MODERATE COMPLEXITY**

**Title** TECHNICAL CONSULTANT-MODERATE COMPLEXITY

**CFR** 493.1409

**Type** Condition

**Regulation Definition**

The laboratory must have a technical consultant who meets the qualification requirements of §493.1411 of this subpart and provides technical oversight in accordance with §493.1413 of this subpart.

**Interpretive Guideline**

The Condition of technical consultant is not met when the technical consultant:

- o Position is not filled;
- o Is not qualified; or
- o Does not fulfill the technical consultant's responsibilities.

**FED - D6034 - TECHNICAL CONSULTANT QUALIFICATIONS**

**Title** TECHNICAL CONSULTANT QUALIFICATIONS

**CFR** 493.1411

**Type** Standard

**Regulation Definition**

The laboratory must employ one or more individuals who are qualified by education and either training or experience to provide technical consultation for each of the specialties and subspecialties of service in which the laboratory performs

**Interpretive Guideline**

Interpretative Guidelines §493.1411

The type of experience required under this regulation is clinical in nature. This means, examination and test performance on human specimens for purposes of obtaining information for the diagnosis, treatment, and monitoring of patients, or for providing information to others who will do the diagnosing and treating of the patient's condition.

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moderate complexity tests or procedures. The director of a laboratory performing moderate complexity testing may function as the technical consultant provided he or she meets the qualifications specified in this section.

Patient or medically-oriented experience, which is defined as the ordering of tests and interpreting and applying the results of these tests in diagnosing and treating a patient's illness is unacceptable to meet the requirement for laboratory training or experience.

The term "laboratory training or experience" means that the individual qualifying has the training and experience in the specialties and subspecialties in which the individual is providing technical consultation.

Technical consultants should have documentation of hands-on testing experience. This documentation may consist of, but is not limited to, the individual's initials on worksheets or work cards, attestation of the laboratory director to the experience the individual has, or formal laboratory rotation through a medical residency program or laboratory internship program.

Teaching experience directly related to a medical technology program, clinical laboratory sciences program, or a clinical laboratory section of a residency program is considered acceptable experience. Research experience is also acceptable experience if it is obtained while performing tests on human specimens.

**FED - D6035 - TECHNICAL CONSULTANT QUALIFICATIONS**

**Title** TECHNICAL CONSULTANT QUALIFICATIONS

**CFR** 493.1411

**Type** Standard

**Regulation Definition**

- (a) The technical consultant must be qualified and must possess a current license issued by the State in which the laboratory is located, if such licensing is required.
- (b) The technical consultant must--
  - (b)(1)(i) Be a doctor of medicine or doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located; and
  - (b)(1)(ii) Be certified in anatomic or clinical pathology, or both, by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification; or

**Interpretive Guideline**

Interpretative Guidelines §493.1411(b)(1)(ii)  
Qualifications that are equivalent for certification include board eligibility, i.e., the individual meets all education, training, or experience requirements to take the examination, but has not actually taken and successfully completed the examination. An individual who wishes to qualify as a technical consultant must supply evidence of this eligibility status. The designated boards, upon request, will send a letter to the individual confirming his/her eligibility status. Note that some boards set time restrictions for taking the examination. For purposes of the regulations, the individual must meet the education, training or experience required by the board to be eligible to take the examination and must have confirmation of eligibility status.

Interpretative Guidelines §493.1411(b)(3)-(b)(4)  
See §493.2 for the definition of an accredited institution.

Some examples of how the one-year requirement for training or experience can be met are:

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(b)(2)(i) Be a doctor of medicine, doctor of osteopathy, or doctor of podiatric medicine licensed to practice medicine, osteopathy, or podiatry in the State in which the laboratory is located; and

- o Medical technology internship;
- o 1 year experience performing nonwaived tests in a particular specialty(ies) or subspecialty(ies); or
- o Performance of nonwaived testing in a particular specialty(ies) or subspecialty(ies) on a part-time basis, equivalent to 2080 hours.

(b)(2)(ii) Have at least one year of laboratory training or experience, or both in non-waived testing, in the designated specialty or subspecialty areas of service for which the technical consultant is responsible (for example, physicians certified either in hematology or hematology and medical oncology by the American Board of Internal Medicine are qualified to serve as the technical consultant in hematology); or

NOTE: §493.1411(b)(4) requires 2 years of laboratory training or experience and can be met by any combination equivalent to 2 years of laboratory training or experience.

(b)(3)(i) Hold an earned doctoral or master's degree in a chemical, physical, biological or clinical laboratory science or medical technology from an accredited institution; and

(b)(3)(ii) Have at least one year of laboratory training or experience, or both in non-waived testing, in the designated specialty or subspecialty areas of service for which the technical consultant is responsible; or

(b)(4)(i) Have earned a bachelor's degree in a chemical, physical or biological science or medical technology from an accredited institution; and

(b)(4)(ii) Have at least 2 years of laboratory training or experience, or both in non-waived testing, in the designated specialty or subspecialty areas of service for which the technical consultant is responsible.

Note: The technical consultant requirements for "laboratory training or experience, or both" in each specialty or subspecialty may be acquired concurrently in more than one of

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the specialties or subspecialties of service, excluding waived tests. For example, an individual who has a bachelor's degree in biology and additionally has documentation of 2 years of work experience performing tests of moderate complexity in all specialties and subspecialties of service, would be qualified as a technical consultant in a laboratory performing moderate complexity testing in all specialties and subspecialties of service.

**FED - D6036 - TECHNICAL CONSULTANT RESPONSIBILITIES**

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413

**Type** Standard

**Regulation Definition**

The technical consultant is responsible for the technical and scientific oversight of the laboratory.

**Interpretive Guideline**

Interpretative Guidelines §493.1413

In a specialty in which neither the director nor testing personnel can qualify to provide technical consultation, the laboratory may engage the services of a qualified person either on a part-time or full-time basis for this service. Under these circumstances, the qualified person is not required to be on the premises full-time or at all times tests are being performed in his/her specialty(ies). However, the technical consultant must be available to provide consultation and should spend time in the laboratory sufficient to supervise the technical performance of the staff in his/her specialty(ies).

**FED - D6037 - TECHNICAL CONSULTANT RESPONSIBILITIES**

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413

**Type** Standard

**Regulation Definition**

The technical consultant is not required to be onsite at all times testing is performed; however, he or she must be available to the laboratory on an as needed basis to provide

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consultation, as specified in paragraph (a) of this section.

**FED - D6038 - TECHNICAL CONSULTANT RESPONSIBILITIES**

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(a)

**Type** Standard

**Regulation Definition**

The technical consultant must be accessible to the laboratory to provide on-site, telephone, or electronic consultation.

**Interpretive Guideline**

Interpretative Guidelines §493.1413(a)

Since the testing personnel usually will not have experience and training in all specialties, technical consultation is essential in identifying training needs and assuring that each individual performing testing receives regular in-service training and education. There should be documentation, such as a log book or training/discussion reports, to indicate the services provided or activities performed by the technical consultant. These activities should correlate with the responsibilities delegated to the technical consultant by the laboratory director. The technical consultant is responsible for evaluating the capabilities of the technical personnel and advising the director on proper test performance in the specialty.

**FED - D6039 - TECHNICAL CONSULTANT RESPONSIBILITIES**

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(1)

**Type** Standard

**Regulation Definition**

The technical consultant is responsible for--  
(b)(1) Selection of test methodology appropriate for the clinical use of the test results;

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FED - D6040 - TECHNICAL CONSULTANT RESPONSIBILITIES

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(2)

**Type** Standard

**Regulation Definition**

The technical consultant is responsible for--  
(b)(2) Verification of the test procedures performed and the establishment of the laboratory's test performance characteristics, including the precision and accuracy of each test and test system.

**Interpretive Guideline**

FED - D6041 - TECHNICAL CONSULTANT RESPONSIBILITIES

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(3)

**Type** Standard

**Regulation Definition**

(b) The technical consultant is responsible for--  
(b)(3) Enrollment and participation in an HHS approved proficiency testing program commensurate with the services offered;

**Interpretive Guideline**

FED - D6042 - TECHNICAL CONSULTANT RESPONSIBILITIES

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(4)

**Type** Standard

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**Regulation Definition**

**Interpretive Guideline**

(b) The technical consultant is responsible for--  
(b)(4) Establishing a quality control program appropriate for the testing performed and establishing the parameters for acceptable levels of analytic performance and ensuring that these levels are maintained throughout the entire testing process from the initial receipt of the specimen, through sample analysis and reporting of test results;

FED - D6043 - TECHNICAL CONSULTANT RESPONSIBILITIES

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(5)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

(b) The technical consultant is responsible for--  
(b)(5) Resolving technical problems and ensuring that remedial actions are taken whenever test systems deviate from the laboratory's established performance specifications;

FED - D6044 - TECHNICAL CONSULTANT RESPONSIBILITIES

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(6)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

(b) The technical consultant is responsible for--  
(b)(6) Ensuring that patient test results are not reported until all corrective actions have been taken and the test system is functioning properly;

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**FED - D6045 - TECHNICAL CONSULTANT RESPONSIBILITIES**

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(7)

**Type** Standard

**Regulation Definition**

(b) The technical consultant is responsible for--  
(b)(7) Identifying training needs and assuring that each individual performing tests receives regular in-service training and education appropriate for the type and complexity of the laboratory services performed;

**Interpretive Guideline**

Interpretative Guidelines §493.1413(b)(7)

In some instances, in-service training may be specifically related to an instrument or test, or may be very general in nature. The laboratory may establish its own format, content, and schedule or provide training on an as-needed basis. This is acceptable provided the laboratory does not have deficiencies related to test performance.

**FED - D6046 - TECHNICAL CONSULTANT RESPONSIBILITIES**

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(8)

**Type** Standard

**Regulation Definition**

(b) The technical consultant is responsible for--  
(b)(8) Evaluating the competency of all testing personnel and assuring that the staff maintain their competency to perform test procedures and report test results promptly, accurately and proficiently.

**Interpretive Guideline**

Interpretative Guidelines §493.1413(b)(8)

All testing personnel must be listed on the CMS Form 209 and must undergo documented competency assessment. The technical consultant/supervisor is responsible for assessing the competency of the testing personnel, and the 6 competency assessment criteria are found under the technical consultant/supervisor responsibilities. Depending on the situation, non-compliance can be cited at General Laboratory Systems (D5209/§493.1235), laboratory director (D6030/§493.1407 or D6103/§493.1445, or technical consultant/supervisor (D6046-D6055/§493.1413(b)(8)-§493.1413(b)(9)).

Probes §493.1413(b)(8)

What mechanism is used to ensure that testing personnel are following the laboratory's policies and procedures?

Evaluations of technical and clinical consultants' performance is located at §493.1235 - Personnel Competency Assessment Policies and §§493.1239(a)-(b) - General Laboratory Systems Assessment.

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FED - D6047 - TECHNICAL CONSULTANT RESPONSIBILITIES

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(8)(i)

**Type** Standard

**Regulation Definition**

The procedures for evaluation of the competency of the staff must include, but are not limited to direct observations of routine patient test performance, including patient preparation, if applicable, specimen handling, processing and testing.

**Interpretive Guideline**

FED - D6048 - TECHNICAL CONSULTANT RESPONSIBILITIES

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(8)(ii)

**Type** Standard

**Regulation Definition**

The procedures for evaluation of the competency of the staff must include, but are not limited to monitoring the recording and reporting of test results.

**Interpretive Guideline**

FED - D6049 - TECHNICAL CONSULTANT RESPONSIBILITIES

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(8)(iii)

**Type** Standard

**Regulation Definition**

The procedures for evaluation of the competency of the staff

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must include, but are not limited to review of intermediate test results or worksheets, quality control records, proficiency testing results, and preventive maintenance records.

**FED - D6050 - TECHNICAL CONSULTANT RESPONSIBILITIES**

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(8)(iv)

**Type** Standard

**Regulation Definition**

The procedures for evaluation of the competency of the staff must include, but are not limited to direct observation of performance of instrument maintenance and function checks.

**Interpretive Guideline**

**FED - D6051 - TECHNICAL CONSULTANT RESPONSIBILITIES**

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(8)(v)

**Type** Standard

**Regulation Definition**

The procedures for evaluation of the competency of the staff must include, but are not limited to assessment of test performance through testing previously analyzed specimens, internal blind testing samples or external proficiency testing samples.

**Interpretive Guideline**

**FED - D6052 - TECHNICAL CONSULTANT RESPONSIBILITIES**

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(8)(vi)

**Type** Standard

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**Regulation Definition**

The procedures for evaluation of the competency of the staff must include, but are not limited to assessment of problem solving skills.

**Interpretive Guideline**

**FED - D6053 - TECHNICAL CONSULTANT RESPONSIBILITIES**

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(9)

**Type** Standard

**Regulation Definition**

The technical consultant is responsible for evaluating and documenting the performance of individuals responsible for moderate complexity testing at least semiannually during the first year the individual tests patient specimens.

**Interpretive Guideline**

**FED - D6054 - TECHNICAL CONSULTANT RESPONSIBILITIES**

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(9)

**Type** Standard

**Regulation Definition**

The technical consultant is responsible for evaluating and documenting the performance of individuals responsible for moderate complexity testing at least annually, after the first year.

**Interpretive Guideline**

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**FED - D6055 - TECHNICAL CONSULTANT RESPONSIBILITIES**

**Title** TECHNICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1413(b)(9)

**Type** Standard

**Regulation Definition**

The technical consultant is responsible for evaluating and documenting the performance of individuals responsible for moderate complexity testing whenever test methodology or instrumentation changes. The individual's performance must be reevaluated to include the use of the new test methodology or instrumentation prior to reporting patient test results.

**Interpretive Guideline**

**FED - D6056 - CLINICAL CONSULTANT**

**Title** CLINICAL CONSULTANT

**CFR** 493.1415

**Type** Condition

**Regulation Definition**

The laboratory must have a clinical consultant who meets the qualification requirements of §493.1417 of this part and provides clinical consultation in accordance with §493.1419 of this part.

**Interpretive Guideline**

Interpretative Guidelines §493.1415

The Condition of clinical consultant is not met when the clinical consultant:

- o Position is not filled;
- o Is not qualified; or
- o Does not fulfill the clinical consultant's responsibilities.

**FED - D6057 - CLINICAL CONSULTANT QUALIFICATIONS**

**Title** CLINICAL CONSULTANT QUALIFICATIONS

**CFR** 493.1417

**Type** Standard

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**Regulation Definition**

The clinical consultant must be qualified to consult with and render opinions to the laboratory's clients concerning the diagnosis, treatment and management of patient care. The clinical consultant must--

(a) Be qualified as a laboratory director under §493.1405(b) (1), (2), or (3)(i); or

(b) Be a doctor of medicine, doctor of osteopathy or doctor of podiatric medicine and possess a license to practice medicine, osteopathy or podiatry in the State in which the laboratory is located.

**Interpretive Guideline**

**FED - D6058 - CLINICAL CONSULTANT RESPONSIBILITIES**

**Title** CLINICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1419

**Type** Standard

**Regulation Definition**

The clinical consultant provides consultation regarding the appropriateness of the testing ordered and interpretation of test results.

**Interpretive Guideline**

**FED - D6059 - CLINICAL CONSULTANT RESPONSIBILITIES**

**Title** CLINICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1419(a)

**Type** Standard

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**Regulation Definition**

The clinical consultant must be available to provide clinical consultation to the laboratory's clients.

**Interpretive Guideline**

**FED - D6060 - CLINICAL CONSULTANT RESPONSIBILITIES**

**Title** CLINICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1419(b)

**Type** Standard

**Regulation Definition**

The clinical consultant must be available to assist the laboratory's clients in ensuring that appropriate tests are ordered to meet the clinical expectations.

**Interpretive Guideline**

**FED - D6061 - CLINICAL CONSULTANT RESPONSIBILITIES**

**Title** CLINICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1419(c)

**Type** Standard

**Regulation Definition**

The clinical consultant must ensure that reports of test results include pertinent information required for specific patient interpretation.

Probes §493.1419(c)

Has the clinical consultant reviewed the reports to ensure that test results include patient information required for specific patient interpretations?

**Interpretive Guideline**

**FED - D6062 - CLINICAL CONSULTANT RESPONSIBILITIES**

**Title** CLINICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1419(d)

**Type** Standard

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**Regulation Definition**

The clinical consultant must ensure that consultation is available and communicated to the laboratory's clients on matters related to the quality of the test results reported and their interpretation concerning specific patient conditions.

**Interpretive Guideline**

**FED - D6063 - LABORATORY TESTING PERSONNEL**

**Title** LABORATORY TESTING PERSONNEL

**CFR** 493.1421

**Type** Condition

**Regulation Definition**

The laboratory must have a sufficient number of individuals who meet the qualification requirements of §493.1423, to perform the functions specified in §493.1425 for the volume and complexity of tests performed.

**Interpretive Guideline**

Interpretative Guidelines §493.1421

The Condition of testing personnel is not met when the testing personnel:

- o Is not qualified ;or
- o Does not fulfill the testing personnel responsibilities.

The criteria used to determine the adequacy of the testing personnel involves evaluating testing personnel responsibilities, and ensuring that these responsibilities are specified in writing by the director, and that the responsibilities are appropriate to ensure compliance with the requirements concerning reporting and recordkeeping, quality control monitoring, quality assurance activities and proficiency testing participation. Cite this deficiency only when compliance problems are found in these areas that can be directly related to insufficient numbers of testing personnel. (Use D6028, which relates the finding of insufficient personnel to director responsibilities.)

**FED - D6064 - TESTING PERSONNEL QUALIFICATIONS**

**Title** TESTING PERSONNEL QUALIFICATIONS

**CFR** 493.1423(a)

**Type** Standard

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**Regulation Definition**

Each individual performing moderate complexity testing must possess a current license issued by the State in which the laboratory is located, if such licensing is required.

**Interpretive Guideline**

Interpretative Guidelines §493.1423

The laboratory director is responsible for ensuring the testing personnel have the appropriate education and experience, and receive the appropriate training for the type and complexity of testing performed. The experience required is clinical in nature. This means, examination of and test performance on human specimens for purposes of obtaining information for the diagnosis, treatment, and monitoring of patients, or for providing information to others who will do the diagnosing and treating of the patient's condition. (Use D6029).

Each individual must have documentation of training applicable to the types and complexity of testing performed. This training should be such that the individual can demonstrate that he/she has the skills required for proper performance of preanalytic, analytic, and postanalytic phases of testing. For example, if the individual performs a rapid Strep test, he/she should be able to demonstrate the skills for:

- o Proper specimen handling prior to testing, e.g., assuring the specimen is properly labeled and received and tested within appropriate timeframes, the swab is received at the proper temperature, and the ampule on the swab containing transport media is broken;
- o Proper test performance according to the laboratory's policies and manufacturer's instructions, e.g., using reagents that are not outdated, are at the proper temperature, and of the same lot number, accurate timing of all steps in the procedure, proper performance of quality control procedures; and
- o Proper reporting of patient test results in accordance with the laboratory's policies, e.g., notifying the person authorized to receive test results of a positive result, not reporting the test result if quality control fails.

Training may include, but is not limited to, attendance at:

- o Seminars given by experts in the field, e.g., a lecture about antibiotic resistance given by the infection control officer of a local hospital;
- o On-site or off-site instrument trainings given by a manufacturer, e.g., a week-long training course given at the manufacturer's headquarters, or training by a manufacturer's technical representative on an instrument purchased by a laboratory;
- o Technical training sessions, workshops, or conferences given by a professional laboratory organization, e.g., CAP, ASMT, AACC, and ASCT;
- o Technical education classes or specialty courses that include hands-on test performance, e.g., parasitology, bacteriology, cytology, given by CDC, a State Health Department, or professional laboratory organizations;
- o A formal laboratory training program; or
- o Inservices offered by a local hospital laboratory staff, pathologist, or medical technologist to a physician's office personnel.

Documentation may consist of, but is not limited to, letters from training programs or employers, attestation

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statements by the laboratory director, a log sheet initialed by the attendees indicating attendance at a training session/in-service, certificates from organizations providing the training session, workshop, conference, specialty course.

**FED - D6065 - TESTING PERSONNEL QUALIFICATIONS**

**Title** TESTING PERSONNEL QUALIFICATIONS

**CFR** 493.1423(b)(1)(2)(3)(4)(i)

**Type** Standard

**Regulation Definition**

(b) Meet one of the following requirements:  
(b)(1) Be a doctor of medicine or doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located or have earned a doctoral, master's, or bachelor's degree in a chemical, physical, biological or clinical laboratory science, or medical technology from an accredited institution; or  
(b)(2) Have earned an associate degree in a chemical, physical or biological science or medical laboratory technology from an accredited institution; or  
(b)(3) Be a high school graduate or equivalent and have successfully completed an official military medical laboratory procedures course of at least 50 weeks duration and have held the military enlisted occupational specialty of Medical Laboratory Specialist (Laboratory Technician); or  
(b)(4)(i) Have earned a high school diploma or equivalent; and

**Interpretive Guideline**

Interpretative Guidelines §493.1423(b)(1)  
See §493.2 for the definition of an accredited institution.

Interpretative Guidelines §493.1423(b)(3)  
Equate similar military courses with different titles. Evaluate the course length and content to ensure that it provides effective training for testing personnel. Refer to "A Guide to the Evaluation of Educational Experience in the Armed Services," American Council on Education, Washington, D.C.

Interpretative Guidelines §493.1423(b)(4)  
Personnel qualifying under this requirement must have a high school diploma or GED.

Probes §493.1423(b)(4)  
How does the laboratory ensure that personnel receiving orientation and training have the necessary skills for properly performing assigned responsibilities?

**FED - D6066 - TESTING PERSONNEL QUALIFICATIONS**

**Title** TESTING PERSONNEL QUALIFICATIONS

**CFR** 493.1423(b)(4)(ii)

**Type** Standard

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**Regulation Definition**

Have documentation of training appropriate for the testing performed prior to analyzing patient specimens.

**Interpretive Guideline**

**FED - D6067 - TESTING PERSONNEL QUALIFICATIONS**

**Title** TESTING PERSONNEL QUALIFICATIONS

**CFR** 493.1423(b)(4)(ii)

**Type** Standard

**Regulation Definition**

Each individual performing moderate complexity testing must have training to ensure that the individual has--

- (A) the skills required for proper specimen collection, including patient preparation, if applicable, labeling, handling, preservation or fixation, processing or preparation, transportation and storage of specimens;
- (B) the skills required for implementing all standard laboratory procedures;
- (C) the skills required for performing each test method and for proper instrument use;
- (D) the skills required for performing preventive maintenance, troubleshooting and calibration procedures related to each test performed;
- (E) a working knowledge of reagent stability and storage;
- (F) the skills required to implement the quality control policies and procedures of the laboratory;
- (G) an awareness of the factors that influence test results; and
- (H) the skills required to assess and verify the validity of patient test results through the evaluation of quality control sample values prior to reporting patient test results.

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**FED - D6068 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1425

**Type** Standard

**Regulation Definition**

The testing personnel are responsible for specimen processing, test performance, and for reporting test results.

**Interpretive Guideline**

**FED - D6069 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1425(a)

**Type** Standard

**Regulation Definition**

Each individual performs only those moderate complexity tests that are authorized by the laboratory director and require a degree of skill commensurate with the individual's education, training or experience, and technical abilities.

**Interpretive Guideline**

**FED - D6070 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1425(b)(1)

**Type** Standard

**Regulation Definition**

Each individual performing moderate complexity testing must follow the laboratory's procedures for specimen handling and

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processing, test analyses, reporting and maintaining records of patient test results.

**FED - D6071 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1425(b)(2)

**Type** Standard

**Regulation Definition**

Each individual performing moderate complexity testing must maintain records that demonstrate that proficiency testing samples are tested in the same manner as patient samples.

**Interpretive Guideline**

**FED - D6072 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1425(b)(3)

**Type** Standard

**Regulation Definition**

Each individual performing moderate complexity testing must adhere to the laboratory's quality control policies, document all quality control activities, instrument and procedural calibrations and maintenance performed.

**Interpretive Guideline**

**FED - D6073 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1425(b)(4)

**Type** Standard

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**Regulation Definition**

Each individual performing moderate complexity testing must follow the laboratory's established corrective action policies and procedures whenever test systems are not within the laboratory's established acceptable levels of performance.

**Interpretive Guideline**

**FED - D6074 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1425(b)(5)

**Type** Standard

**Regulation Definition**

Each individual performing moderate complexity testing must be capable of identifying problems that may adversely affect test performance or reporting of test results and either must correct the problems or immediately notify the technical consultant, clinical consultant or director.

**Interpretive Guideline**

Interpretative Guidelines §493.1425(b)(5)

If, during the survey, testing personnel demonstrate an inability to identify a problem that adversely affects a patient test result, cite D6029 under director responsibilities.

Some examples of problems that may adversely affect patient test results may include, but are not limited to:

- o A pleural fluid that is mislabeled and, therefore, is processed as a urine culture;
- o Performing a potassium on a hemolyzed sample; or
- o Tests are incubated at 37°C when the manufacturer's instructions require 25°C incubation.

**FED - D6075 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1425(b)(6)

**Type** Standard

**Regulation Definition**

Each individual performing moderate complexity testing must document all corrective actions taken when test systems deviate from the laboratory's established performance specifications.

**Interpretive Guideline**

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**FED - D6076 - LABORATORY DIRECTOR**

**Title** LABORATORY DIRECTOR

**CFR** 493.1441

**Type** Condition

**Regulation Definition**

The laboratory must have a director who meets the qualification requirements of §493.1443 of this subpart and provides overall management and direction in accordance with §493.1445 of this subpart.

**Interpretive Guideline**

Interpretative Guidelines §493.1441

The Condition of laboratory director is not met when the laboratory director:

- o Position is not filled;
- o Is not qualified; or
- o Does not fulfill the laboratory director responsibilities.

**FED - D6078 - LABORATORY DIRECTOR QUALIFICATIONS**

**Title** LABORATORY DIRECTOR QUALIFICATIONS

**CFR** 493.1443

**Type** Standard

**Regulation Definition**

The laboratory director must be qualified to manage and direct the laboratory personnel and performance of high complexity tests and must be eligible to be an operator of a laboratory within the requirements of subpart R.

(a) The laboratory director must possess a current license as a laboratory director issued by the State in which the laboratory is located, if such licensing is required; and

(b) The laboratory director must--

(b)(1)(i) Be a doctor of medicine or doctor of osteopathy licensed to practice medicine or osteopathy in the State in

**Interpretive Guideline**

Interpretative Guidelines §493.1443

When qualifying a Laboratory Director, please refer to section 353(i)(3) of the PHS Act states "No person who has owned or operated a laboratory which has had its certificate revoked may, within 2 years of the revocation of the certificate, own or operate a laboratory for which a certificate has been issued under this section."

Interpretative Guidelines §493.1443(a)

The term "State" as used in this provision, includes the District of Columbia, the Commonwealth of Puerto Rico, the Commonwealth of Northern Mariana Islands, the Virgin Islands, Guam and American Samoa.

Interpretative Guidelines §493.1443(b)(1)(ii)

Qualifications that are equivalent for certification include board eligibility, i.e., the individual meets all education, training, or experience requirements to take the examination, but has not actually taken and successfully completed

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which the laboratory is located; and

(b)(1)(ii) Be certified in anatomic or clinical pathology, or both, by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification; or

(b)(2) Be a doctor of medicine, a doctor of osteopathy or doctor of podiatric medicine licensed to practice medicine, osteopathy or podiatry in the State in which the laboratory is located; and

(b)(2)(i) Have at least one year of laboratory training during medical residency (for example, physicians certified either in hematology or hematology and medical oncology by the American Board of Internal Medicine); or

(b)(2)(ii) Have at least 2 years of experience directing or supervising high complexity testing; or

(b)(3) Hold an earned doctoral degree in a chemical, physical, biological or clinical laboratory science from an accredited institution and--

(b)(3)(i) Be certified and continue to be certified by a board approved by HHS; or

(b)(3)(ii) Before February 24, 2003, must have served or be serving as director of a laboratory performing high complexity testing and must have at least--

(b)(3)(ii)(A) Two years of laboratory training or experience, or both; and

(b)(3)(ii)(B) Two years of laboratory experience directing or

the examination. An individual who wishes to qualify as a director must supply evidence of this eligibility status. The designated boards, upon request, will send a letter to the individual confirming his/her eligibility status. Note that some boards set time restrictions for taking the examination. For purposes of the regulations, the individual must meet the education, training, or experience as required by the board to be eligible to take the examination and must have confirmation of eligibility status.

*Interpretative Guidelines §493.1443(b)(2)(i)*

The residency program should provide the director the knowledge in principles and theories of laboratory practice including: quality control and quality assessment, proficiency testing, the phase of the total process (i.e., preanalytic, analytic and postanalytic), as well as, general laboratory systems, facility administration, and development and implementation of personnel policy and procedure manuals. This training should also include hands-on laboratory testing.

*Interpretative Guidelines §493.1443(b)(2)(ii)*

The type of experience required under this regulation is clinical in nature. This means directing or supervising personnel who examine and perform tests on human specimens for the purpose of providing information that is used in diagnosing, treating, and monitoring a patient's condition. This experience may include the laboratory director personally examining and performing tests on patient specimens. Patient or medically-oriented experience, which is defined as the ordering of tests and interpreting and applying the results of these tests in diagnosing and treating a patient's illness is unacceptable to meet the requirement for laboratory training or experience.

The laboratory director should have documentation, e.g., signed procedure manuals, test reports, worksheets and workcards, that indicates the director assumes the responsibilities in §493.1445.

Teaching experience directly related to a medical technology program, clinical laboratory sciences program, or a clinical laboratory section of a residency program is considered acceptable experience. Research experience is also acceptable experience if it is obtained while performing tests on human specimens.

*Interpretative Guidelines §493.1443(b)(3)*

See §493.2 for the definition of and guidance for accredited institution.

To qualify as a laboratory director of high complexity testing on or after February 24, 2003, individuals possessing a Ph.D. or Dr.P.H. must be board certified by an approved board.

"Certified" means the individual has completed all the designated board's requirements, including the examination.

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supervising high complexity testing.

(b)(4) Be serving as a laboratory director and must have previously qualified or could have qualified as a laboratory director under regulations at 42 CFR 493.1415, published March 14, 1990 at 55 FR 9538, on or before February 28, 1992; or

(b)(5) On or before February 28, 1992, be qualified under State law to direct a laboratory in the State in which the laboratory is located; or

(b)(6) For the subspecialty of oral pathology, be certified by the American Board of Oral Pathology, American Board of Pathology, the American Osteopathic Board of Pathology, or possess qualifications that are equivalent to those required for certification.

Currently approved boards are:

ABB - American Board of Bioanalysis,

ABB public health microbiology certification,

ABCC - American Board of Clinical Chemistry,

ABCC 24 month-Commission on Accreditation in Clinical Chemistry (COMACC) accredited program

ABFT - American Board of Forensic Toxicology (limited to individuals with a doctoral degree)\*,

ABHI - American Board of Histocompatibility and Immunogenetics,

ABMGG - American Board of Medical Genetics and Genomics (formerly ABMG - American Board of Medical Genetics),

ABMLI - American Board of Medical Laboratory Immunology,

ABMM - American Board of Medical Microbiology,

NRCC - National Registry for Certified Chemists (limited to individuals with a doctoral degree)\*,

\*NOTE: ABFT and NRCC also certify non-doctoral individuals; however, the director of high-complexity testing must have a doctoral degree.

An acceptable doctoral degree is a Doctor of Philosophy - Ph.D., Doctor of Science - D.Sc. If acceptable to the board, a Doctor of Dental Surgery - D.D.S., Doctor of Veterinary Medicine - D.V.M., Doctor of Public Health - Dr.P.H.

Laboratory testing of non-human specimens is not acceptable experience, e.g., environmental, animal testing.

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##### Interpretative Guidelines §493.1443(b)(4)

An individual is qualified as a laboratory director if he or she was serving as a laboratory director on or before February 28, 1992. After February 28, 1992, individuals must meet the requirements at §493.1443(b)(1)-(3) to qualify as a laboratory director for high complexity.

In accordance with the regulations, the requirements listed below may be used only for individuals meeting these qualifications and functioning in the position as of February 28, 1992.

The requirements for a laboratory director under 42 CFR 493.1415, published March 14, 1990 (55 FR 9538) are as follows:

(a) The laboratory director must possess a current license as a laboratory director issued by the State, if such licensing exists; and

(b) The laboratory director must:

(b)(1) Be a physician certified in anatomical or clinical pathology (or both) by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification;

(b)(2) Be a physician who: (b)(2)(i) is certified by the American Board of Pathology or the American Osteopathic Board of Pathology in at least one of the laboratory specialties, or (b)(2)(ii) is certified by the American Board of Medical Microbiology, the American Board of Clinical Chemistry, the American Board of Bioanalysis, or other national accrediting board in one of the laboratory specialties, or (b)(2)(iii) is certified by the American Society of Cytology to practice cytopathology or possesses qualifications that are equivalent to those required for such certification, or (b)(2)(iv) subsequent to graduation, has had 4 or more years of full-time general laboratory training and experience of which at least 2 years were spent acquiring proficiency in one of the laboratory specialties;

(b)(3) For the subspecialty of oral pathology only, be certified by the American Board of Oral Pathology, American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for certification;

(b)(4) Hold an earned doctoral degree from an accredited institution with a chemical, physical, or biological science as a major subject and (b)(4)(i) is certified by the American Board of Medical Microbiology, the American Board of Clinical Chemistry, the American Board of Bioanalysis, or other national accrediting board acceptable to HHS in one of the laboratory specialties, or (b)(4)(ii) subsequent to graduation has had 4 or more years of full time general laboratory training and experience of which at least 2 years were spent acquiring proficiency in one of the laboratory specialties;

(b)(5) With respect to individuals first qualifying before July 1, 1971, have been responsible for the direction of a laboratory for 12 months between July 1, 1961, and January 1, 1968, and in addition, either:

(b)(5)(i) Was a physician and subsequent to graduation had at least 4 years of pertinent full-time laboratory

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experience;

- (b)(5)(ii) Held a master's degree from an accredited institution with a chemical, physical, or biological science as a major subject and subsequent to graduation had at least 4 years of pertinent full-time laboratory experience;
- (b)(5)(iii) Held a bachelor's degree from an accredited institution with a chemical, physical, or biological science as a major subject and subsequent to graduation had at least 6 years of pertinent full-time laboratory experience; or
- (b)(5)(iv) Achieved a satisfactory grade through an examination conducted by or under the sponsorship of the U.S. Public Health Service on or before July 1, 1970; or
- (b)(6) Qualify under State law to direct the laboratory in the State in which the laboratory is located.

Interpretative Guidelines §493.1443(b)(5)

Those individuals qualified after February 28, 1992, as directors solely under State law, will not meet this requirement.

**FED - D6079 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(a)(b)

**Type** Standard

**Regulation Definition**

The laboratory director is responsible for the overall operation and administration of the laboratory, including the employment of personnel who are competent to perform test procedures, record and report test results promptly, accurately and proficiently, and for assuring compliance with the applicable regulations.

(a) The laboratory director, if qualified, may perform the duties of the technical supervisor, clinical consultant, general supervisor, and testing personnel, or delegate these responsibilities to personnel meeting the qualifications under 493.1447, 493.1453, 493.1459, and 493.1487 respectively.

(b) If the laboratory director reappoints performance of his or her responsibilities, he or she remains responsible for ensuring that all duties are properly performed.

**Interpretive Guideline**

Interpretative Guidelines §493.1445

The requirement that a laboratory must be under the direction of a qualified person is not automatically met simply because the director meets the education and experience requirements. It must be demonstrated that the individual is, in fact, providing effective direction over the operation of the laboratory.

In determining whether the director responsibilities are met, consider deficiencies found in other conditions, e.g., facility administration, general laboratory systems, preanalytic systems, analytic systems, postanalytic systems, and proficiency testing.

If the laboratory has more than one person qualifying as a director, one individual must be designated as accepting ultimate responsibility for the overall operation and administration of the laboratory.

Interpretative Guidelines §493.1445(a)

An individual qualified as laboratory director under §493.1443 may not qualify as technical supervisor in a particular specialty or subspecialty unless he or she has the required training or experience. If the director of high complexity testing is not qualified to perform the duties of the technical supervisor or clinical consultant, he or she must employ

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individual(s) meeting the respective qualifications.

The laboratory director may reappportion to a technical supervisor, in writing, the responsibilities in: §§493.1445(e) (3), (4), (5), (6), (7), (12), (13), and (14).

The laboratory director may reappportion to a clinical consultant, in writing, the responsibilities in: §§493.1445(e)(8) and (9).

The only responsibilities that may be delegated to the general supervisor are listed at §§493.1463(b)(1)-(4).

**FED - D6080 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(c)

**Type** Standard

**Regulation Definition**

The laboratory director must be accessible to the laboratory to provide onsite, telephone or electronic consultation as needed.

**Interpretive Guideline**

Interpretative Guidelines §493.1445(c)

If the director cannot practically provide personal, on-site supervision, it must be demonstrated that the director:

- o Provides direction and consultation electronically (e.g., email, text message or fax) or by telephone, as necessary;  
or
- o Delegates to qualified personnel specific responsibilities as provided in the regulations.

**FED - D6081 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(d)

**Type** Standard

**Regulation Definition**

Each individual may direct no more than five laboratories.

**Interpretive Guideline**

Interpretative Guidelines §493.1445(d)

An individual may serve as a director of 5 nonwaived certified laboratories. However, an individual may serve as

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technical consultant, clinical consultant or technical supervisor for any number of laboratories.

**FED - D6082 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(1)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure that testing systems developed and used for each of the tests performed in the laboratory provide quality laboratory services for all aspects of test performance, which includes the preanalytic, analytic, and postanalytic phases of testing.

**Interpretive Guideline**

**FED - D6083 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(2)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure that the physical plant and environmental conditions of the laboratory are appropriate for the testing performed.

**Interpretive Guideline**

**FED - D6084 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(2)

**Type** Standard

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**Regulation Definition**

The laboratory director must ensure that the physical plant and environmental conditions provide a safe environment in which employees are protected from physical, chemical, and biological hazards.

**Interpretive Guideline**

Interpretative Guidelines §493.1445(e)(2)  
OSHA/EPA issues cannot be cited using these requirements. If immediate jeopardy exists, inform the director immediately.

If you observe or obtain information regarding potential safety violations not applicable under CLIA, notify the appropriate State or local authority. Consult with the Regional Office (RO) for notification to other Federal agencies such as the Occupational Safety and Health Administration (OSHA) [www.osha.gov], Environmental Protection Agency (EPA) [www.epa.gov], or Nuclear Regulatory Commission (NRC). The appropriate Federal, State or local authority, if warranted, will investigate and, if necessary, conduct an on-site visit.

**FED - D6085 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(3)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure that the test methodologies selected have the capability of providing the quality of results required for patient care.

**Interpretive Guideline**

**FED - D6086 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(3)(ii)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure that verification procedures used are adequate to determine the accuracy, precision, and other pertinent performance characteristics of the method.

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**FED - D6087 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(3)(iii)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure that laboratory personnel are performing the test methods as required for accurate and reliable results.

**Interpretive Guideline**

**FED - D6088 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(4)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure that the laboratory is enrolled in an HHS-approved proficiency testing program for the testing performed.

**Interpretive Guideline**

**FED - D6089 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(4)(i)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure the proficiency testing samples are tested as required under subpart H of this part.

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**FED - D6090 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(4)(ii)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure the results are returned within the timeframes established by the proficiency testing program.

**Interpretive Guideline**

**FED - D6091 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(4)(iii)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure all proficiency testing reports received are reviewed by the appropriate staff to evaluate the laboratory's performance and to identify any problems that require corrective action.

**Interpretive Guideline**

**FED - D6092 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(4)(iv)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure an approved corrective

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action plan is followed when any proficiency testing result is found to be unacceptable or unsatisfactory.

**FED - D6093 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(5)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure that the quality control programs are established and maintained to assure the quality of laboratory services provided and to identify failures in quality as they occur.

**Interpretive Guideline**

**FED - D6094 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(5)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure that the quality assessment programs are established and maintained to assure the quality of laboratory services provided and to identify failures in quality as they occur.

**Interpretive Guideline**

**FED - D6095 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(6)

**Type** Standard

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**Regulation Definition**

The laboratory director must ensure the establishment and maintenance of acceptable levels of analytical performance for each test system.

**Interpretive Guideline**

**FED - D6096 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(7)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure that all necessary remedial actions are taken and documented whenever significant deviations from the laboratory's established performance characteristics are identified.

**Interpretive Guideline**

**FED - D6097 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(7)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure that patient test results are reported only when the system is functioning properly.

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**FED - D6098 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(8)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure that reports of test results include pertinent information required for interpretation.

**Interpretive Guideline**

**FED - D6099 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(9)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure that consultation is available to the laboratory's clients on matters relating to the quality of the test results reported and their interpretation concerning specific patient conditions.

**Interpretive Guideline**

**FED - D6100 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(10)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure that a general supervisor provides on-site supervision of high complexity test

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performance by testing personnel qualified under §493.1489(b)(4).

**FED - D6101 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(11)

**Type** Standard

**Regulation Definition**

The laboratory director must employ a sufficient number of laboratory personnel with the appropriate education and either experience or training to provide appropriate consultation, properly supervise and accurately perform tests and report test results in accordance with the personnel responsibilities described in this subpart.

**Interpretive Guideline**

**FED - D6102 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(12)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure that prior to testing patients' specimens, all personnel have the appropriate education and experience, receive the appropriate training for the type and complexity of the services offered, and have demonstrated that they can perform all testing operations reliably to provide and report accurate results.

**Interpretive Guideline**

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**FED - D6103 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(13)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure that policies and procedures are established for monitoring individuals who conduct preanalytical, analytical, and postanalytical phases of testing to assure that they are competent and maintain their competency to process specimens, perform test procedures and report test results promptly and proficiently, and whenever necessary, identify needs for remedial training or continuing education to improve skills.

**Interpretive Guideline**

Interpretative Guidelines §493.1445(e)(13)

Personnel performing only preanalytic and postanalytic activities are not required to be listed on Form 209. Surveyors do not normally check for documented competency evaluation on these individuals. However, if you discover problems in the laboratory and you find that a factor in these problems is poor performance of incompetent staff, cite D6030 or D6103 (laboratory director).

**FED - D6106 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(14)

**Type** Standard

**Regulation Definition**

The laboratory director must ensure that an approved procedure manual is available to all personnel responsible for any aspect of the testing process.

**Interpretive Guideline**

Interpretative Guideline §493.1445(e)(14)

The laboratory director can delegate to the technical supervisor the responsibility of making the procedure manual available, but cannot delegate the responsibility for signing new and revised procedures.

**FED - D6107 - LABORATORY DIRECTOR RESPONSIBILITIES**

**Title** LABORATORY DIRECTOR RESPONSIBILITIES

**CFR** 493.1445(e)(15)

**Type** Standard

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**Regulation Definition**

The laboratory director must specify, in writing, the responsibilities and duties of each consultant and each supervisor, as well as each person engaged in the performance of the preanalytic, analytic, and postanalytic phases of testing, that identifies which examinations and procedures each individual is authorized to perform, whether supervision is required for specimen processing, test performance or result reporting and whether supervisory or director review is required prior to reporting patient test results.

**Interpretive Guideline**

Interpretative Guidelines §493.1445(e)(15)  
The director must assign, in writing, the duties/responsibilities to each person involved in all phases of the testing process. The list of assigned duties must be current.

**FED - D6108 - LABORATORY TECHNICAL SUPERVISOR**

**Title** LABORATORY TECHNICAL SUPERVISOR

**CFR** 493.1447

**Type** Condition

**Regulation Definition**

The laboratory must have a technical supervisor who meets the qualification requirements of §493.1449 of this subpart and provides technical supervision in accordance with §493.1451 of this subpart.

**Interpretive Guideline**

§493.1447 Guidelines:  
The Condition of technical supervisor is not met when the technical supervisor:

- o Position is not filled;
- o Is not qualified; or
- o Does not fulfill the technical supervisor responsibilities.

**FED - D6109 - TECHNICAL SUPERVISOR QUALIFICATIONS**

**Title** TECHNICAL SUPERVISOR QUALIFICATIONS

**CFR** 493.1449

**Type** Standard

**Regulation Definition**

The laboratory must employ one or more individuals who are qualified by education and either training or experience to

**Interpretive Guideline**

Interpretative Guidelines §493.1449  
The type of experience required under this regulation is clinical in nature. This means examination and test

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provide technical supervision for each of the specialties and subspecialties of service in which the laboratory performs high complexity tests or procedures. The director of a laboratory performing high complexity testing may function as the technical supervisor provided he or she meets the qualifications specified in this section.

performance on human specimens for purposes of obtaining information for the diagnosis, treatment, and monitoring of patients, or for providing information to others who will do the diagnosing and treating of the patient's condition. Patient or medically-oriented experience, which is defined as the ordering of tests and interpreting and applying the results of these tests in diagnosing and treating a patient's illness is unacceptable to meet the requirement for laboratory training or experience.

The term "laboratory training or experience" means that the individual qualifying has the training in and the experience with the specialties and subspecialties in which the individual is performing technical supervision. For technical supervisor, the requirement for training or experience can be met through any combination of training and/or experience in high complexity testing. This can be acquired subsequent to, concurrent with, or prior to obtaining academic requirements.

Be flexible in evaluating laboratory training and experience. The specified training or experience may be acquired simultaneously in more than one specialty/subspecialty. Although it is unreasonable in ?§493.1449(c)(5) and (j)(5) to expect four full-time years devoted only to high complexity microbiology testing and then four full-time years performing high complexity tests only in hematology, etc., to qualify under each specialty/subspecialty, it is necessary for the individual to have had continuous responsibilities in the specialty for the designated number of years and it would be more than simply performing an occasional test. Technical supervisors should have documentation of hands-on testing experience. This documentation may consist of, but is not limited to, the individual's initials on worksheets or work cards, attestation of the laboratory director to the experience the individual has, or formal laboratory rotation through a medical residency program or laboratory internship program.

Teaching experience directly related to a medical technology program, clinical laboratory sciences program, or a clinical laboratory section of a residency program is considered acceptable experience. Research experience is also acceptable experience if it is obtained while performing tests on human specimens.

A year of laboratory training or experience is equivalent to 2080 hours and could extend over more than one 12 calendar-month period.

**FED - D6111 - TECHNICAL SUPERVISOR QUALIFICATIONS**

**Title** TECHNICAL SUPERVISOR QUALIFICATIONS

**CFR** 493.1449

**Type** Standard

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**Regulation Definition**

- (a) The technical supervisor must possess a current license issued by the State in which the laboratory is located, if such licensing is required; and
- (b) The laboratory may perform anatomic and clinical laboratory procedures and tests in all specialties and subspecialties of services except histocompatibility and clinical cytogenetics services provided the individual functioning as the technical supervisor--
- (b)(1) Is a doctor of medicine or doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located; and
- (b)(2) Is certified in both anatomic and clinical pathology by the American Board of Pathology or the American Osteopathic Board of Pathology or Possesses qualifications that are equivalent to those required for such certification.
- (c) If the requirements of paragraph (b) of this section are not met and the laboratory performs tests in the subspecialty of bacteriology, the individual functioning as the technical supervisor must--
- (c)(1)(i) Be a doctor of medicine or doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located; and
- (c)(1)(ii) Be certified in clinical pathology by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification; or
- (c)(2)(i) Be a doctor of medicine, doctor of osteopathy, or doctor of podiatric medicine licensed to practice medicine, osteopathy, or podiatry in the State in which the laboratory is located; and
- (c)(2)(ii) Have at least one year of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of bacteriology; or

**Interpretive Guideline**

Interpretative Guidelines §493.1449(b)(2)

Qualifications that are equivalent for certification includes board eligibility, i.e., the individual meets all education, training, or experience requirements to take the examination, but has not actually taken and successfully completed the examination. An individual who wishes to qualify as a technical supervisor must supply evidence of this eligibility status. The designated boards, upon request, will send a letter to the individual confirming his/her eligibility status. Note that some boards set time restrictions for taking the examination. For purposes of the regulations, the individual must meet the education, training or experience required by the board to be eligible to take the examination and must have confirmation of eligibility status.

The tests in histopathology include gross examination (macro) and microscopic slide evaluation and interpretation with diagnostic reporting.

Interpretative Guidelines §493.1449(c)(1)(ii)

NOTE: See Interpretative Guidelines for §493.1449(b)(2)

Interpretative Guidelines §493.1449(c)(3)(i)

See §493.2 for the definition of an accredited institution.

Interpretative Guidelines §493.1449(d)(1)(ii)

NOTE: See Interpretative Guidelines for §493.1449(b)(2).

Interpretative Guidelines §493.1449(d)(3)(i)

See §493.2 for the definition of an accredited institution.

Interpretative Guidelines §493.1449(e)(1)(ii)

NOTE: See Interpretative Guidelines for §493.1449(b)(2)

Interpretative Guidelines §493.1449(e)(3)(i)

See §493.2 for the definition of an accredited institution.

Interpretative Guidelines §493.1449(f)(1)(ii)

NOTE: See Interpretative Guidelines for §493.1449(b)(2)

Interpretative Guidelines §493.1449(f)(3)(i)

See §493.2 for the definition of an accredited institution.

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(c)(3)(i) Have an earned doctoral degree in a chemical, physical, biological or clinical laboratory science from an accredited institution; and

(c)(3)(ii) Have at least 1 year of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of bacteriology; or

(c)(4)(i) Have earned a master's degree in a chemical, physical, biological or clinical laboratory science or medical technology from an accredited institution; and

(c)(4)(ii) Have at least 2 years of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of bacteriology; or

(c)(5)(i) Have earned a bachelor's degree in a chemical, physical, or biological science or medical technology from an accredited institution; and

(c)(5)(ii) Have at least 4 years of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of bacteriology.

(d) If the requirements of paragraph (b) of this section are not met and the laboratory performs tests in the subspecialty of mycobacteriology, the individual functioning as the technical supervisor must--

(d)(1)(i) Be a doctor of medicine or doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located; and

(d)(1)(ii) Be certified in clinical pathology by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification; or

Interpretative Guidelines §493.1449(g)(1)(ii)

NOTE: See Interpretative Guidelines for §493.1449(b)(2)

Interpretative Guidelines §493.1449(g)(3)(i)

See §493.2 for the definition of an accredited institution.

Interpretative Guidelines §493.1449(h)(1)(i)

NOTE: See Interpretative Guidelines for §493.1449(b)(2)

Interpretative Guidelines §493.1449(h)(3)(i)

See §493.2 for the definition of an accredited institution.

Interpretative Guidelines §493.1449 (i)(1)(ii)

NOTE: See Interpretative Guidelines for §493.1449(b)(2)

Interpretative Guidelines §493.1449(i)(3)(i)

See §493.2 for the definition of an accredited institution.

Interpretative Guidelines §493.1449 (j)(1)(ii)

NOTE: See Interpretative Guidelines for §493.1449(b)(2)

Interpretative Guidelines §493.1449(j)(3)(i)

See §493.2 for the definition of an accredited institution.

Interpretative Guidelines §493.1449(k)(1)(ii)(A) or (B)

NOTE: See Interpretative Guidelines for §493.1449(b)(2)

Interpretative Guidelines §493.1449(l)(1)(i)(B)

NOTE: See Interpretative Guidelines for §493.1449(b)(2)

Interpretative Guidelines §493.1449(l)(2)(i)(B)(1),(2), or (3)

NOTE: See Interpretative Guidelines for §493.1449(b)(2)

Interpretative Guidelines §493.1449(l)(3)(i)(B)(1) or (2)

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(d)(2)(i) Be a doctor of medicine, doctor of osteopathy, or doctor or podiatric medicine licensed to practice medicine, osteopathy, or podiatry in the State in which the laboratory is located; and

(d)(2)(ii) Have at least 1 year of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of mycobacteriology; or

(d)(3)(i) Have an earned doctoral degree in a chemical, physical, biological or clinical laboratory science from an accredited institution; and

(d)(3)(ii) Have at least 1 year of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of mycobacteriology; or

(d)(4)(i) Have earned a master's degree in a chemical, physical, biological or clinical laboratory science or medical technology from an accredited institution; and

(d)(4)(ii) Have at least 2 years of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of mycobacteriology; or

(d)(5)(i) Have earned a bachelor's degree in a chemical, physical or biological science or medical technology from an accredited institution; and

(d)(5)(ii) Have at least 4 years of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of mycobacteriology.

(e) If the requirements of paragraph (b) of this section are not met and the laboratory performs tests in the subspecialty of

NOTE: See Interpretative Guidelines for §493.1449(b)(2)

Interpretative Guidelines §493.1449(n)(1)(ii)

NOTE: See Interpretative Guidelines for §493.1449(b)(2)

Interpretative Guidelines §493.1449(n)(3)(i)

See §493.2 for the definition of an accredited institution.

Interpretative Guidelines §493.1449(o)(2)(i)

See §493.2 for the definition of an accredited institution.

Interpretative Guidelines §493.1449(p)(2)(i)

See §493.2 for the definition of an accredited institution.

Interpretative Guidelines §493.1449(q)(1)(ii)

NOTE: See Interpretative Guidelines for §493.1449(b)(2)

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mycology, the individual functioning as the technical supervisor must--

- (e)(1)(i) Be a doctor of medicine or doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located; and
- (e)(1)(ii) Be certified in clinical pathology by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification; or
- (e)(2)(i) Be a doctor of medicine, doctor of osteopathy, or doctor of podiatric medicine licensed to practice medicine, osteopathy, or podiatry in the State in which the laboratory is located; and
- (e)(2)(ii) Have at least 1 year of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of mycology; or
- (e)(3)(i) Have an earned doctoral degree in a chemical, physical, biological or clinical laboratory science from an accredited institution; and
- (e)(3)(ii) Have at least 1 year of laboratory training or experience, or both in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of mycology; or
- (e)(4)(i) Have earned a master's degree in a chemical, physical, biological or clinical laboratory science or medical technology from an accredited institution; and
- (e)(4)(ii) Have at least 2 years of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of mycology; or
- (e)(5)(i) Have earned a bachelor's degree in a chemical,

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physical or biological science or medical technology from an accredited institution; and

(e)(5)(ii) Have at least 4 years of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of mycology.

(f) If the requirements of paragraph (b) of this section are not met and the laboratory performs tests in the subspecialty of parasitology, the individual functioning as the technical supervisor must--

(f)(1)(i) Be a doctor of medicine or a doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located; and

(f)(1)(ii) Be certified in clinical pathology by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification; or

(f)(2)(i) Be a doctor of medicine, doctor of osteopathy, or doctor of podiatric medicine licensed to practice medicine, osteopathy, or podiatry in the State in which the laboratory is located; and

(f)(2)(ii) Have at least one year of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of parasitology;

(f)(3)(i) Have an earned doctoral degree in a chemical, physical, biological or clinical laboratory science from an accredited institution; and

(f)(3)(ii) Have at least 1 year of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of parasitology; or

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(f)(4)(i) Have earned a master's degree in a chemical, physical, biological or clinical laboratory science or medical technology from an accredited institution; and

(f)(4)(ii) Have at least 2 years of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of parasitology; or

(f)(5)(i) Have earned a bachelor's degree in a chemical, physical or biological science or medical technology from an accredited institution; and

(f)(5)(ii) Have at least 4 years of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of parasitology.

(g) If the requirements of paragraph (b) of this section are not met and the laboratory performs tests in the subspecialty of virology, the individual functioning as the technical supervisor must--

(g)(1)(i) Be a doctor of medicine or doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located; and

(g)(1)(ii) Be certified in clinical pathology by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification; or

(g)(2)(i) Be a doctor of medicine, doctor of osteopathy, or doctor of podiatric medicine licensed to practice medicine, osteopathy, or podiatry in the State in which the laboratory is located; and

(g)(2)(ii) Have at least 1 year of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty

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of virology; or

(g)(3)(i) Have an earned doctoral degree in a chemical, physical, biological or clinical laboratory science from an accredited institution; and

(g)(3)(ii) Have at least 1 year of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of virology; or

(g)(4)(i) Have earned a master's degree in a chemical, physical, biological or clinical laboratory science or medical technology from an accredited institution; and

(g)(4)(ii) Have at least 2 years of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of virology; or

(g)(5)(i) Have earned a bachelor's degree in a chemical, physical or biological science or medical technology from an accredited institution; and

(g)(5)(ii) Have at least 4 years of laboratory training or experience, or both, in high complexity testing within the specialty of microbiology with a minimum of 6 months experience in high complexity testing within the subspecialty of virology.

(h) If the requirements of paragraph (b) of this section are not met and the laboratory performs tests in the specialty of diagnostic immunology, the individual functioning as the technical supervisor must-

(h)(1)(i) Be a doctor of medicine or a doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located; and

(h)(1)(ii) Be certified in clinical pathology by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those

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required for such certification; or

(h)(2)(i) Be a doctor of medicine, doctor of osteopathy, or doctor of podiatric medicine licensed to practice medicine, osteopathy, or podiatry in the State in which the laboratory is located; and

(h)(2)(ii) Have at least 1 year of laboratory training or experience, or both, in high complexity testing for the specialty of diagnostic immunology; or

(h)(3)(i) Have an earned doctoral degree in a chemical, physical, biological or clinical laboratory science from an accredited institution; and

(h)(3)(ii) Have at least 1 year of laboratory training or experience, or both, in high complexity testing within the specialty of diagnostic immunology; or

(h)(4)(i) Have earned a master's degree in a chemical, physical, biological or clinical laboratory science or medical technology from an accredited institution; and

(h)(4)(ii) Have at least 2 years of laboratory training or experience, or both, in high complexity testing for the specialty of diagnostic immunology; or

(h)(5)(i) Have earned a bachelor's degree in a chemical, physical or biological science or medical technology from an accredited institution; and

(h)(5)(ii) Have at least 4 years of laboratory training or experience, or both, in high complexity testing for the specialty of diagnostic immunology.

(i) If the requirements of paragraph (b) of this section are not met and the laboratory performs tests in the specialty of chemistry, the individual functioning as the technical supervisor must--

(i)(1)(i) Be a doctor of medicine or doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located; and

(i)(1)(ii) Be certified in clinical pathology by the American Board of Pathology or the American Osteopathic Board of

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Pathology or possess qualifications that are equivalent to those required for such certification; or

(i)(2)(i) Be a doctor of medicine, doctor of osteopathy, or doctor of podiatric medicine licensed to practice medicine, osteopathy, or podiatry in the State in which the laboratory is located; and

(i)(2)(ii) Have at least 1 year of laboratory training or experience, or both, in high complexity testing for the specialty of chemistry; or

(i)(3)(i) Have an earned doctoral degree in a chemical, physical, biological or clinical laboratory science from an accredited institution; and

(i)(3)(ii) Have at least 1 year of laboratory training or experience, or both, in high complexity testing within the specialty of chemistry; or

(i)(4)(i) Have earned a master's degree in a chemical, physical, biological or clinical laboratory science or medical technology from an accredited institution; and

(i)(4)(ii) Have at least 2 years of laboratory training or experience, or both, in high complexity testing for the specialty of chemistry; or

(i)(5)(i) Have earned a bachelor's degree in a chemical, physical or biological science or medical technology from an accredited institution; and

(i)(5)(ii) Have at least 4 years of laboratory training or experience, or both, in high complexity testing for the specialty of chemistry.

(j) If the requirements of paragraph (b) of this section are not met and the laboratory performs tests in the specialty of hematology, the individual functioning as the technical supervisor must--

(j)(1)(i) Be a doctor of medicine or a doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located; and

(j)(1)(ii) Be certified in clinical pathology by the American

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Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification; or

(j)(2)(i) Be a doctor of medicine, doctor of osteopathy, or doctor of podiatric medicine licensed to practice medicine, osteopathy, or podiatry in the State in which the laboratory is located; and

(j)(2)(ii) Have at least one year of laboratory training or experience, or both, in high complexity testing for the specialty of hematology (for example, physicians certified either in hematology or hematology and medical oncology by the American Board of Internal Medicine); or

(j)(3)(i) Have an earned doctoral degree in a chemical, physical, biological or clinical laboratory science from an accredited institution; and

(j)(3)(ii) Have at least 1 year of laboratory training or experience, or both, in high complexity testing within the specialty of hematology; or

(j)(4)(i) Have earned a master's degree in a chemical, physical, biological or clinical laboratory science or medical technology from an accredited institution; and

(j)(4)(ii) Have at least 2 years of laboratory training or experience, or both, in high complexity testing for the specialty of hematology; or

(j)(5)(i) Have earned a bachelor's degree in a chemical, physical or biological science or medical technology from an accredited institution; and

(j)(5)(ii) Have at least 4 years of laboratory training or experience, or both, in high complexity testing for the specialty of hematology.

(k)(1) If the requirements of paragraph (b) of this section are not met and the laboratory performs tests in the subspecialty of cytology, the individual functioning as the technical supervisor must--

(k)(1)(i) Be a doctor of medicine or a doctor of osteopathy

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licensed to practice medicine or osteopathy in the State in which the laboratory is located; and

(k)(1)(ii) Meet one of the following requirements--

(k)(1)(ii)(A) Be certified in anatomic pathology by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification; or

(k)(1)(ii)(B) Be certified by the American Society of Cytology to practice cytopathology or possess qualifications that are equivalent to those required for such certification;

(l) If the requirements of paragraph (b) of this section are not met and the laboratory performs tests in the subspecialty of histopathology, the individual functioning as the technical supervisor must--

(l)(1) Meet one of the following requirements:

(l)(1)(i)(A) Be a doctor of medicine or a doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located; and

(l)(1)(i)(B) Be certified in anatomic pathology by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification;

(l)(1)(ii) An individual qualified under §493.1449(b) or paragraph (l)(1) of this section may delegate to an individual who is a resident in a training program leading to certification specified in paragraph (b) or (l)(1)(i)(B) of this section, the responsibility for examination and interpretation of histopathology specimens.

(l)(2) For tests in dermatopathology, meet one of the following requirements:

(l)(2)(i)(A) Be a doctor of medicine or doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located and--

(l)(2)(i)(B) Meet one of the following requirements:

(l)(2)(i)(B)(1) Be certified in anatomic pathology by the

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American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification; or (1)(2)(i)(B)(2) Be certified in dermatopathology by the American Board of Dermatology and the American Board of Pathology or possess qualifications that are equivalent to those required for such certification; or (1)(2)(i)(B)(3) Be certified in dermatology by the American Board of Dermatology or possess qualifications that are equivalent to those required for such certification; or (1)(2)(ii) An individual qualified under §493.1449(b) or paragraph (1)(2)(i) of this section may delegate to an individual who is a resident in a training program leading to certification specified in paragraphs (b) or (1)(2)(i)(B) of this section, the responsibility for examination and interpretation of dermatopathology specimens.

(1)(3) For tests in ophthalmic pathology, meet one of the following requirements:

(1)(3)(i)(A) Be a doctor of medicine or doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located and--

(1)(3)(i)(B) Must meet one of the following requirements:

(1)(3)(i)(B)(1) Be certified in anatomic pathology by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification; or

(1)(3)(i)(B)(2) Be certified by the American Board of Ophthalmology or possess qualifications that are equivalent to those required for such certification and have successfully completed at least 1 year of formal post-residency fellowship training in ophthalmic pathology; or

(1)(3)(ii) An individual qualified under §493.1449(b) or paragraph (1)(3)(i) of this section may delegate to an individual who is a resident in a training program leading to certification specified in paragraphs (b) or (1)(3)(i)(B) of this

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section, the responsibility for examination and interpretation of ophthalmic specimens; or

(m) If the requirements of paragraph (b) of this section are not met and the laboratory performs tests in the subspecialty of oral pathology, the individual functioning as the technical supervisor must meet one of the following requirements:

(m)(1)(i) Be a doctor of medicine or a doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located and--

(m)(1)(ii) Be certified in anatomic pathology by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification; or

(m)(2) Be certified in oral pathology by the American Board of Oral Pathology or possess qualifications for such certification; or

(m)(3) An individual qualified under §493.1449(b) or paragraph (m)(1) or (2) of this section may delegate to an individual who is a resident in a training program leading to certification specified in paragraphs (b) or (m)(1) or (2) of this section, the responsibility for examination and interpretation of oral pathology specimens.

(n) If the requirements of paragraph (b) of this section are not met and the laboratory performs tests in the specialty of radiobioassay, the individual functioning as the technical supervisor must--

(n)(1)(i) Be a doctor of medicine or a doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located; and

(n)(1)(ii) Be certified in clinical pathology by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification; or

(n)(2)(i) Be a doctor of medicine, doctor of osteopathy, or doctor of podiatric medicine licensed to practice medicine,

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osteopathy, or podiatry in the State in which the laboratory is located; and

(n)(2)(ii) Have at least 1 year of laboratory training or experience, or both, in high complexity testing for the specialty of radiobioassay; or

(n)(3)(i) Have an earned doctoral degree in a chemical, physical, biological or clinical laboratory science from an accredited institution; and

(n)(3)(ii) Have at least 1 year of laboratory training or experience, or both, in high complexity testing within the specialty of radiobioassay; or

(n)(4)(i) Have earned a master's degree in a chemical, physical, biological or clinical laboratory science or medical technology from an accredited institution; and

(n)(4)(ii) Have at least 2 years of laboratory training or experience, or both, in high complexity testing for the specialty of radiobioassay; or

(n)(5)(i) Have earned a bachelor's degree in a chemical, physical or biological science or medical technology from an accredited institution; and

(n)(5)(ii) Have at least 4 years of laboratory training or experience, or both, in high complexity testing for the specialty of radiobioassay.

(o) If the laboratory performs tests in the specialty of histocompatibility, the individual functioning as the technical supervisor must either--

(o)(1)(i) Be a doctor of medicine, doctor of osteopathy, or doctor of podiatric medicine licensed to practice medicine, osteopathy, or podiatry in the State in which the laboratory is located; and

(o)(1)(ii) Have training or experience that meets one of the following requirements:

(o)(1)(ii)(A) Have 4 years of laboratory training or experience, or both, within the specialty of histocompatibility;

or

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(o)(1)(ii)(B)(1) Have 2 years of laboratory training or experience, or both, in the specialty of general immunology;

and

(o)(1)(ii)(B)(2) Have 2 years of laboratory training or experience, or both, in the specialty of histocompatibility; or

(o)(2)(i) Have an earned doctoral degree in a biological or clinical laboratory science from an accredited institution; and

(o)(2)(ii) Have training or experience that meets one of the following requirements:

(o)(2)(ii)(A) Have 4 years of laboratory training or experience, or both, within the specialty of histocompatibility;

or

(o)(2)(ii)(B)(1) Have 2 years of laboratory training or experience, or both, in the specialty of general immunology;

and

(o)(2)(ii)(B)(2) Have 2 years of laboratory training or experience, or both, in the specialty of histocompatibility.

(p) If the laboratory performs tests in the specialty of clinical cytogenetics, the individual functioning as the technical supervisor must--

(p)(1)(i) Be a doctor of medicine, doctor of osteopathy, or doctor of podiatric medicine licensed to practice medicine, osteopathy, or podiatry in the State in which the laboratory is located; and

(p)(1)(ii) Have 4 years of training or experience, or both, in genetics, 2 of which have been in clinical cytogenetics; or

(p)(2)(i) Hold an earned doctoral degree in a biological science, including biochemistry, or clinical laboratory science from an accredited institution; and

(p)(2)(ii) Have 4 years of training or experience, or both, in genetics, 2 of which have been in clinical cytogenetics.

(q) If the requirements of paragraph (b) of this section are not met and the laboratory performs tests in the specialty of immunohematology, the individual functioning as the technical supervisor must--

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- (q)(1)(i) Be a doctor of medicine or a doctor of osteopathy licensed to practice medicine or osteopathy in the State in which the laboratory is located; and
- (q)(1)(ii) Be certified in clinical pathology by the American Board of Pathology or the American Osteopathic Board of Pathology or possess qualifications that are equivalent to those required for such certification; or

Note: The technical supervisor requirements for "laboratory training or experience, or both" in each specialty or subspecialty may be acquired concurrently in more than one of the specialties or subspecialties of service. For example, an individual, who has a doctoral degree in chemistry and additionally has documentation of 1 year of laboratory experience working concurrently in high complexity testing in the specialties of microbiology and chemistry and 6 months of that work experience included high complexity testing in bacteriology, mycology, and mycobacteriology, would qualify as the technical supervisor for the specialty of chemistry and the subspecialties of bacteriology, mycology, and mycobacteriology.

**FED - D6112 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451

**Type** Standard

**Regulation Definition**

The technical supervisor is responsible for the technical and scientific oversight of the laboratory. The technical supervisor is not required to be on site at all times testing is performed; however, he or she must be available to the laboratory on an as needed basis to provide supervision as specified in (a) of this section.

**Interpretive Guideline**

Interpretative Guidelines §493.1451

In a specialty in which neither the director nor the general supervisor can qualify to provide technical supervision, the laboratory may engage the services of a qualified person either on a part-time or full-time basis for this service. The technical supervisor is not required to be on the premises full-time or at all times tests are being performed in his/her specialty(ies). However, the technical supervisor must be available to provide consultation and is required to spend an amount of time in the laboratory sufficient to supervise the technical performance of the staff in his/her specialty(ies). There should be documentation, such as a log book or notes from training which indicate the technical

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supervisor performs his/her assigned duties. The technical supervisor is responsible for evaluating the capabilities of the testing personnel and the general supervisor's testing performance.

**FED - D6113 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(a)

**Type** Standard

**Regulation Definition**

The technical supervisor must be accessible to the laboratory to provide on-site, telephone, or electronic consultation.

**Interpretive Guideline**

**FED - D6114 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(b)(1)

**Type** Standard

**Regulation Definition**

The technical supervisor is responsible for selection of the test methodology that is appropriate for the clinical use of the test results.

**Interpretive Guideline**

**FED - D6115 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(b)(2)

**Type** Standard

**Regulation Definition**

The technical supervisor is responsible for verification of the

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test procedures performed and establishment of the laboratory's test performance characteristics, including the precision and accuracy of each test and test system.

**FED - D6116 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(b)(3)

**Type** Standard

**Regulation Definition**

The technical supervisor is responsible for enrollment and participation in an HHS approved proficiency testing program commensurate with the services offered.

**Interpretive Guideline**

Interpretative Guidelines §493.1451(b)(3)

Any laboratory testing patient specimens for the Human Papillomavirus (HPV) must enroll and successfully participate in a CMS-approved proficiency testing program for HPV beginning in 2008. Laboratories should refer to Subpart H for further information. The laboratory's CLIA certificate must include the subspecialty of Virology. The laboratory must also be in compliance with all the CLIA regulations governing the preanalytic, analytic, and postanalytic phases of testing including proficiency testing and personnel requirements.

**FED - D6117 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(b)(4)

**Type** Standard

**Regulation Definition**

The technical supervisor is responsible for establishing a quality control program appropriate for the testing performed and establishing the parameters for acceptable levels of analytic performance and ensuring that these levels are maintained throughout the entire testing process from the initial receipt of the specimen, through sample analysis and reporting of test results.

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**FED - D6118 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(b)(5)

**Type** Standard

**Regulation Definition**

The technical supervisor is responsible for resolving technical problems and ensuring that remedial actions are taken whenever test systems deviate from the laboratory's established performance specifications.

**Interpretive Guideline**

**FED - D6119 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(b)(6)

**Type** Standard

**Regulation Definition**

The technical supervisor is responsible for ensuring that patient test results are not reported until all corrective actions have been taken and the test system is functioning properly.

**Interpretive Guideline**

**FED - D6120 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(b)(7)(8)

**Type** Standard

**Regulation Definition**

(7) The technical supervisor is responsible for identifying

Interpretative Guidelines §493.1451(b)(7)

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training needs and assuring that each individual performing tests receives regular in-service training and education appropriate for the type and complexity of the laboratory services performed;

(8) Evaluating the competency of all testing personnel and assuring that the staff maintain their competency to perform test procedures and report test results promptly, accurately and proficiently.

In some instances, in-service training may be specifically related to an instrument or test, or may be very general in nature. The laboratory may establish its own format, content, and schedule or provide training on an as-needed basis. This is acceptable provided the laboratory does not have deficiencies related to test performance.

Interpretative Guidelines §493.1451(b)(8)

All testing personnel must be listed on the CMS Form 209 and must undergo documented competency assessment. The technical consultant/supervisor is responsible for assessing the competency of the testing personnel, and the 6 competency assessment criteria are found under the technical consultant/supervisor responsibilities. Depending on the situation, non-compliance can be cited at General Laboratory Systems (D5209/§493.1235), laboratory director (D6030/§493.1407 or D6103/§493.1445, or technical consultant/supervisor D6046-D6055/§493.1413(b)(8)-§493.1413(b)(9)).

Probes §493.1451(b)(8)

What mechanism is used to ensure that testing personnel are following the laboratory's policies and procedures? When approved by the director, these policies and procedures may include manufacturer's instructions.

**FED - D6121 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(b)(8)(i)

**Type** Standard

**Regulation Definition**

The procedures for evaluation of the competency of the staff must include, but are not limited to direct observations of routine patient test performance, including patient preparation, if applicable, specimen handling, processing and testing.

**Interpretive Guideline**

**FED - D6122 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(b)(8)(ii)

**Type** Standard

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**Regulation Definition**

The procedures for evaluation of the competency of the staff must include, but are not limited to monitoring the recording and reporting of test results.

**Interpretive Guideline**

**FED - D6123 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(b)(8)(iii)

**Type** Standard

**Regulation Definition**

The procedures for evaluation of the competency of the staff must include, but are not limited to review of intermediate test results or worksheets, quality control records, proficiency testing results, and preventive maintenance records.

**Interpretive Guideline**

**FED - D6124 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(b)(8)(iv)

**Type** Standard

**Regulation Definition**

The procedures for evaluation of the competency of the staff must include, but are not limited to direct observation of performance of instrument maintenance and function checks.

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**FED - D6125 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(b)(8)(v)

**Type** Standard

**Regulation Definition**

The procedures for evaluation of the competency of the staff must include, but are not limited to assessment of test performance through testing previously analyzed specimens, internal blind testing samples or external proficiency testing samples.

**Interpretive Guideline**

**FED - D6126 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(b)(8)(vi)

**Type** Standard

**Regulation Definition**

The procedures for evaluation of the competency of the staff must include, but are not limited to assessment of problem solving skills.

**Interpretive Guideline**

**FED - D6127 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(b)(9)

**Type** Standard

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**Regulation Definition**

The technical supervisor is responsible for evaluating and documenting the performance of individuals responsible for high complexity testing at least semiannually during the first year the individual tests patient specimens.

**Interpretive Guideline**

**FED - D6128 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(b)(9)

**Type** Standard

**Regulation Definition**

The technical supervisor is responsible for evaluating and documenting the performance of individuals responsible for high complexity testing at least annually after the first year, unless test methodology or instrumentation changes, in which case, prior to reporting patient test results, the individual's performance must be reevaluated to include the use of the new test methodology or instrumentation.

**Interpretive Guideline**

**FED - D6129 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(c)(1)

**Type** Standard

**Regulation Definition**

(c) In cytology, the technical supervisor or the individual qualified under §493.1449(k)(2)--  
(c)(1) May perform the duties of the cytology general supervisor and the cytotechnologist, as specified in §§493.1471 and 493.1485, respectively;

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**FED - D6130 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(c)(2)(3)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

(c) In cytology, the technical supervisor or the individual qualified under 493.1449(k)(2)--  
(c)(2) Must establish the workload limit for each individual examining slides and  
(c)(3) Must reassess the workload limit for each individual examining slides at least every 6 months and adjust as necessary.

**FED - D6131 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(c)(4)

**Type** Standard

**Regulation Definition**

**Interpretive Guideline**

In cytology, the technical supervisor or the individual qualified under 493.1449(k)(2) must perform the functions specified in 493.1257(d) and (e).

**FED - D6132 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(c)(5)

**Type** Standard

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**Regulation Definition**

In cytology, the technical supervisor or the individual qualified under 493.1449(k)(2) must ensure that each individual examining gynecologic preparations participates in an HHS approved cytology proficiency testing program, as specified in §493.945 and achieves a passing score, as specified in §493.855.

**Interpretive Guideline**

**FED - D6133 - TECHNICAL SUPERVISOR RESPONSIBILITIES**

**Title** TECHNICAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1451(c)(6)

**Type** Standard

**Regulation Definition**

In cytology, the technical supervisor or the individual qualified under 439.1449(k)(2), if responsible for screening cytology slide preparations, must document the number of cytology slides screened in 24 hours and the number of hours devoted during each 24-hour period to screening cytology slides.

**Interpretive Guideline**

**FED - D6134 - CLINICAL CONSULTANT**

**Title** CLINICAL CONSULTANT

**CFR** 493.1453

**Type** Condition

**Regulation Definition**

The laboratory must have a clinical consultant who meets the requirements of §493.1455 of this subpart and provides clinical consultation in accordance with §493.1457 of this subpart.

**Interpretive Guideline**

Interpretative Guidelines §493.1453

The Condition of clinical consultant is not met when the clinical consultant:

- o Position is not filled;
- o Is not qualified; or

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- o Does not fulfill the clinical consultant responsibilities.

**FED - D6135 - CLINICAL CONSULTANT QUALIFICATIONS**

**Title** CLINICAL CONSULTANT QUALIFICATIONS

**CFR** 493.1455

**Type** Standard

**Regulation Definition**

The clinical consultant must be qualified to consult with and render opinions to the laboratory's clients concerning the diagnosis, treatment and management of patient care. The clinical consultant must--

(a) Be qualified as a laboratory director under §493.1443(b) (1), (2), or (3)(i) or, for the subspecialty of oral pathology, §493.1443(b)(6); or

(b) Be a doctor of medicine, doctor of osteopathy, doctor of podiatric medicine licensed to practice medicine, osteopathy, or podiatry in the State in which the laboratory is located.

**Interpretive Guideline**

**FED - D6136 - CLINICAL CONSULTANT RESPONSIBILITIES**

**Title** CLINICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1457

**Type** Standard

**Regulation Definition**

The clinical consultant provides consultation regarding the appropriateness of the testing ordered and interpretation of test results.

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**FED - D6137 - CLINICAL CONSULTANT RESPONSIBILITIES**

**Title** CLINICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1457(a)

**Type** Standard

**Regulation Definition**

The clinical consultant must be available to provide consultation to the laboratory's clients.

**Interpretive Guideline**

**FED - D6138 - CLINICAL CONSULTANT RESPONSIBILITIES**

**Title** CLINICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1457(b)

**Type** Standard

**Regulation Definition**

The clinical consultant must be available to assist the laboratory's clients in ensuring that appropriate tests are ordered to meet the clinical expectations.

**Interpretive Guideline**

**FED - D6139 - CLINICAL CONSULTANT RESPONSIBILITIES**

**Title** CLINICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1457(c)

**Type** Standard

**Regulation Definition**

The clinical consultant must ensure that reports of test results include pertinent information required for specific patient interpretation.

**Interpretive Guideline**

Probe §493.1457(c)

Has the clinical consultant reviewed the reports to ensure that test results include patient information required for specific patient interpretations?

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**FED - D6140 - CLINICAL CONSULTANT RESPONSIBILITIES**

**Title** CLINICAL CONSULTANT RESPONSIBILITIES

**CFR** 493.1457(d)

**Type** Standard

**Regulation Definition**

The clinical consultant must ensure that consultation is available and communicated to the laboratory's clients on matters related to the quality of the test results reported and their interpretation concerning specific patient conditions.

**Interpretive Guideline**

**FED - D6141 - GENERAL SUPERVISOR**

**Title** GENERAL SUPERVISOR

**CFR** 493.1459

**Type** Condition

**Regulation Definition**

The laboratory must have one or more general supervisors who are qualified under §493.1461 of this subpart to provide general supervision in accordance with §493.1463 of this subpart.

**Interpretive Guideline**

Interpretative Guidelines §493.1459

The Condition of general supervisor is not met when the general supervisor:

- o Position is not filled;
- o Is not qualified; or
- o Does not fulfill the general supervisor responsibilities.

**FED - D6142 - GENERAL SUPERVISOR QUALIFICATIONS**

**Title** GENERAL SUPERVISOR QUALIFICATIONS

**CFR** 493.1461

**Type** Standard

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**Regulation Definition**

The laboratory must have one or more general supervisors who, under the direction of the laboratory director and supervision of the technical supervisor, provides day-to-day supervision of testing personnel and reporting of test results. In the absence of the director and technical supervisor, the general supervisor must be responsible for the proper performance of all laboratory procedures and reporting of test results.

**Interpretive Guideline**

Interpretative Guidelines: §493.1461

The type of experience required under this regulation is clinical in nature. This means examination and test performance on human specimens for purposes of obtaining information for the diagnosis, treatment, and monitoring of patients, or for providing information to others who will do the diagnosing and treating of the patient's condition.

Teaching experience directly related to a medical technology program, clinical laboratory sciences program, or a clinical laboratory section of a residency program is considered acceptable experience. Research experience is also acceptable experience if it is obtained while performing tests on human specimens. A year of laboratory training and experience is equivalent to 2080 hours and could extend over more than one 12 calendar-month period.

If all testing personnel have associate degrees, but none meet the training or experience requirement for general supervisor, the duties of the general supervisor must be fulfilled by an appropriately qualified individual. This individual need not be on-site at all times.

**FED - D6143 - GENERAL SUPERVISOR QUALIFICATIONS**

**Title** GENERAL SUPERVISOR QUALIFICATIONS

**CFR** 493.1461

**Type** Standard

**Regulation Definition**

- (a) The general supervisor must possess a current license issued by the State in which the laboratory is located, if such licensing is required; and
- (b) The general supervisor must be qualified as a--
  - (b)(1) Laboratory director under §493.1443; or
  - (b)(2) Technical supervisor under §493.1449.
- (c) If the requirements of paragraph (b)(1) or paragraph (b)(2) of this section are not met, the individual functioning as the general supervisor must--
  - (c)(1)(i) Be a doctor of medicine, doctor of osteopathy, or doctor of podiatric medicine licensed to practice medicine, osteopathy, or podiatry in the State in which the laboratory is

**Interpretive Guideline**

Interpretative Guidelines §493.1461(c)(1)(i)

See §493.2 for the definition of and guidance for an accredited institution.

Interpretative Guidelines §493.1461(d)(3)(i)

NOTE: Many blood gas systems are categorized as moderate complexity tests; therefore, only moderate complexity personnel requirements are applicable. To determine which tests are categorized as waived or nonwaived (i.e., moderate or high complexity tests), refer to the "Specific List For Categorization of Laboratory Test Systems, Assays, and Examinations by Complexity" (<<http://www.gpo.gov/fdsys/pkg/FR-1995-05-15/pdf/95-11653.pdf>>). Test systems, assays, and examinations not yet classified are considered high complexity.

Interpretative Guidelines §493.1461(e)

In the case of gross examinations, the technical supervisor may delegate to individuals qualified under §493.1489 the

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located or have earned a doctoral, master's, or bachelor's degree in a chemical, physical, biological or clinical laboratory science, or medical technology from an accredited institution; and

(c)(1)(ii) Have at least 1 year of laboratory training or experience, or both, in high complexity testing; or

(c)(2)(i) Qualify as testing personnel under §493.1489(b)(2); and

(c)(2)(ii) Have at least 2 years of laboratory training or experience, or both, in high complexity testing; or

(c)(3)(i) Except as specified in paragraph (3)(ii) of this section, have previously qualified as a general supervisor under §493.1462 on or before February 28, 1992.

(c)(3)(ii) Exception. An individual who achieved a satisfactory grade in a proficiency examination for technologist given by HHS between March 1, 1986 and December 31, 1987, qualifies as a general supervisor if he or she meets the requirements of §493.1462 on or before January 1, 1994.

(c)(4) On or before September 1, 1992, have served as a general supervisor of high complexity testing and as of April 24, 1995--

(c)(4)(i) Meet one of the following requirements:

(c)(4)(i)(A) Have graduated from a medical laboratory or clinical laboratory training program approved or accredited by the Accrediting Bureau of Health Education Schools (ABHES), the Commission on Allied Health Education Accreditation (CAHEA), or other organization approved by HHS.

(c)(4)(i)(B) Be a high school graduate or equivalent and have successfully completed an official U.S. military medical laboratory procedures course of at least 50 weeks duration and have held the military enlisted occupational specialty of Medical Laboratory Specialist (Laboratory Technician).

(c)(4)(ii) Have at least 2 years of clinical laboratory training, or experience, or both, in high complexity testing; or

responsibility for the physical examination/description, including color, weight, measurement and other characteristics of the tissue; or other mechanical procedures for which a specific written protocol has been developed.

The technical supervisor is ultimately responsible for the diagnosis related to the gross examination and must sign the examination report. The technical supervisor is not required to provide direct on-site supervision but is responsible for the accuracy of all test results reported. All physical examinations/descriptions of tissue including color, weight, measurement and other characteristics of the tissue; or other mechanical procedures including dissection, inking, marking, and specific orientation for diagnostic interpretation performed in the absence of the technical supervisor by individuals qualified under §493.1489 should be reviewed within 24 hours by the technical supervisor. All microscopic tissue examinations must be performed by individuals qualified under §493.1449(b), (l) or (m), as appropriate.

There should be documentation of the identity of the personnel performing the grossing portion of the test. The name does not necessarily need to be included in the final report because the final report is under the responsibility of the technical supervisor. The decision to include the name/initials of the person grossing in the final report is a laboratory decision, and does not fall under the CLIA requirements. The grossing information should be recorded and maintained to show who performed the test, somewhere in the test record. During a Mohs procedure the surgical test requisition may be the surgical report. D5787 §493.1283(a)(4)

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- (c)(5) On or before September 1, 1992, have served as a general supervisor of high complexity testing and--
- (c)(5)(i) Be a high school graduate or equivalent; and
- (c)(5)(ii) Have had at least 10 years of laboratory training or experience, or both, in high complexity testing, including at least 6 years of supervisory experience between September 1, 1982 and September 1, 1992.
- (d) For blood gas analysis, the individual providing general supervision must--
- (d)(1) Be qualified under §§493.1461(b)(1) or (2), or 493.1461(c); or
- (d)(2)(i) Have earned a bachelor's degree in respiratory therapy or cardiovascular technology from an accredited institution; and
- (d)(2)(ii) Have at least one year of laboratory training or experience, or both, in blood gas analysis; or
- (d)(3)(i) Have earned an associate degree related to pulmonary function from an accredited institution; and
- (d)(3)(ii) Have at least two years of training or experience, or both in blood gas analysis.
- (e) The general supervisor requirement is met in histopathology, oral pathology, dermatopathology, and ophthalmic pathology because all tests and examinations, must be performed:
- (e)(1) In histopathology, by an individual who is qualified as a technical supervisor under §§493.1449(b) or 493.1449(l)(1);
- (e)(2) In dermatopathology, by an individual who is qualified as a technical supervisor under §§493.1449(b) or 493.1449(l) or (2);
- (e)(3) In ophthalmic pathology, by an individual who is qualified as a technical supervisor under §§493.1449(b) or 493.1449(l)(3); and
- (e)(4) In oral pathology, by an individual who is qualified as a technical supervisor under §§493.1449(b) or 493.1449(m).

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**FED - D6144 - GENERAL SUPERVISOR RESPONSIBILITIES**

**Title** GENERAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1463

**Type** Standard

**Regulation Definition**

The general supervisor is responsible for day-to-day supervision or oversight of the laboratory operation and personnel performing testing and reporting test results.

**Interpretive Guideline**

Interpretative Guidelines §493.1463

Interview several testing personnel to elicit information about the duties they perform and the degree of supervision they receive.

**FED - D6145 - GENERAL SUPERVISOR RESPONSIBILITIES**

**Title** GENERAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1463(a)(1)

**Type** Standard

**Regulation Definition**

The general supervisor must be accessible to testing personnel at all times testing is performed to provide on-site, telephone or electronic consultation to resolve technical problems in accordance with policies and procedures established either by the laboratory director or technical supervisor.

**Interpretive Guideline**

**FED - D6146 - GENERAL SUPERVISOR RESPONSIBILITIES**

**Title** GENERAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1463(a)(2)

**Type** Standard

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**Regulation Definition**

The general supervisor is responsible for providing day-to-day supervision of high complexity test performance by a testing personnel qualified under §493.1489.

**Interpretive Guideline**

**FED - D6147 - GENERAL SUPERVISOR RESPONSIBILITIES**

**Title** GENERAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1463(a)(3)

**Type** Standard

**Regulation Definition**

Except as specified in paragraph (c) of this section, the general supervisor must be onsite to provide direct supervision when high complexity testing is performed by any individuals qualified under §493.1489(b)(5).

**Interpretive Guideline**

**FED - D6148 - GENERAL SUPERVISOR RESPONSIBILITIES**

**Title** GENERAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1463(a)(4)

**Type** Standard

**Regulation Definition**

The general supervisor is responsible for monitoring test analyses and specimen examinations to ensure that acceptable levels of analytic performance are maintained.

**Interpretive Guideline**

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**FED - D6149 - GENERAL SUPERVISOR RESPONSIBILITIES**

**Title** GENERAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1463(b)(1)

**Type** Standard

**Regulation Definition**

The director or technical supervisor may delegate to the general supervisor the responsibility for assuring that all remedial actions are taken whenever test systems deviate from the laboratory's established performance specifications.

**Interpretive Guideline**

**FED - D6150 - GENERAL SUPERVISOR RESPONSIBILITIES**

**Title** GENERAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1463(b)(2)

**Type** Standard

**Regulation Definition**

The director or technical supervisor may delegate to the general supervisor the responsibility for ensuring that patient test results are not reported until all corrective actions have been taken and the test system is properly functioning.

**Interpretive Guideline**

**FED - D6151 - GENERAL SUPERVISOR RESPONSIBILITIES**

**Title** GENERAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1463(b)(3)(4)

**Type** Standard

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**Regulation Definition**

- (3) The director or technical supervisor may delegate to the general supervisor the responsibility for providing orientation to all testing personnel; and
- (4) Annually evaluating and documenting the performance of all testing personnel.

**Interpretive Guideline**

**FED - D6152 - GENERAL SUPERVISOR RESPONSIBILITIES**

**Title** GENERAL SUPERVISOR RESPONSIBILITIES

**CFR** 493.1463(c)

**Type** Standard

**Regulation Definition**

Exception. For individuals qualified under 493.1489(b)(5), who were performing high complexity testing on or before January 19, 1993, the requirements of paragraph (a)(3) of this section are not effective, provided that all high complexity testing performed by the individual in the absence of an general supervisor is reviewed within 24 hours by a general supervisor qualified under 493.1461.

**Interpretive Guideline**

**FED - D6153 - CYTOLOGY GENERAL SUPERVISOR**

**Title** CYTOLOGY GENERAL SUPERVISOR

**CFR** 493.1467

**Type** Condition

**Regulation Definition**

For the subspecialty of cytology, the laboratory must have a general supervisor who meets the qualification requirements of §493.1469 of this subpart, and provides supervision in accordance with

**Interpretive Guideline**

Interpretative Guideline §493.1467

The Condition of cytology general supervisor is not met when the cytology general supervisor:

- o Position is not filled;
- o Is not qualified; or

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§493.1471 of this subpart.

- o Does not fulfill the cytology general supervisor responsibilities.

**FED - D6155 - CYTOLOGY SUPERVISOR QUALIFICATIONS**

**Title** CYTOLOGY SUPERVISOR QUALIFICATIONS

**CFR** 493.1469

**Type** Standard

**Regulation Definition**

The cytology general supervisor must be qualified to supervise cytology services. The general supervisor in cytology must possess a current license issued by the State in which the laboratory is located, if such licensing is required, and must--

(a) Be qualified as a technical supervisor under §493.1449 (b) or (k); or

(b)(1) Be qualified as a cytotechnologist under §493.1483; and

(b)(2) Have at least 3 years of full-time (2,080 hours per year) experience as a cytotechnologist within the preceding 10 years.

**Interpretive Guideline**

Interpretative Guidelines §493.1469(b)(2)

In addition to screening slides in a laboratory, the 3 years of full-time experience as a cytotechnologist can be fulfilled if the individual has been:

- o Teaching in schools of cytotechnology;
- o Teaching cytotechnology for residency programs in academic institutions; or
- o Participating in research directly related to cytotechnology, which includes screening slides, library research, and documentation.

**FED - D6156 - CYTOLOGY SUPERVISOR RESPONSIBILITIES**

**Title** CYTOLOGY SUPERVISOR RESPONSIBILITIES

**CFR** 493.1471

**Type** Standard

**Regulation Definition**

The technical supervisor of cytology may perform the duties of the cytology general supervisor or delegate the responsibilities to an individual qualified under §493.1469.

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**FED - D6157 - CYTOLOGY SUPERVISOR RESPONSIBILITIES**

**Title** CYTOLOGY SUPERVISOR RESPONSIBILITIES

**CFR** 493.1471(a)

**Type** Standard

**Regulation Definition**

The cytology general supervisor is responsible for the day-to-day supervision or oversight of the laboratory operation and personnel performing testing and reporting test results.

**Interpretive Guideline**

**FED - D6158 - CYTOLOGY SUPERVISOR RESPONSIBILITIES**

**Title** CYTOLOGY SUPERVISOR RESPONSIBILITIES

**CFR** 493.1471(b)(1)

**Type** Standard

**Regulation Definition**

The cytology general supervisor must be accessible to provide on-site, telephone, or electronic consultation to resolve technical problems in accordance with policies and procedures established by the technical supervisor of cytology.

**Interpretive Guideline**

**FED - D6159 - CYTOLOGY SUPERVISOR RESPONSIBILITIES**

**Title** CYTOLOGY SUPERVISOR RESPONSIBILITIES

**CFR** 493.1471(b)(2)

**Type** Standard

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**Regulation Definition**

The cytology general supervisor must document the slide interpretation results of each gynecologic and nongynecologic cytology case he or she examined or reviewed (as specified under §493.1274(c)).

**Interpretive Guideline**

**FED - D6160 - CYTOLOGY SUPERVISOR RESPONSIBILITIES**

**Title** CYTOLOGY SUPERVISOR RESPONSIBILITIES

**CFR** 493.1471(b)(3)

**Type** Standard

**Regulation Definition**

The cytology general supervisor must for each 24-hour period, document the total number of slides he or she examined or reviewed in the laboratory as well as the total number of slides examined or reviewed in any other laboratory or for any other employer.

**Interpretive Guideline**

**FED - D6161 - CYTOLOGY SUPERVISOR RESPONSIBILITIES**

**Title** CYTOLOGY SUPERVISOR RESPONSIBILITIES

**CFR** 493.1471(b)(4)

**Type** Standard

**Regulation Definition**

The cytology general supervisor must document the number of hours spent examining slides in each 24-hour period.

**Interpretive Guideline**

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**FED - D6162 - CYTOTECHNOLOGIST**

**Title** CYTOTECHNOLOGIST

**CFR** 493.1483

**Type** Condition

**Regulation Definition**

For the subspecialty of cytology, the laboratory must have a sufficient number of cytotechnologists who meet the qualifications specified in §493.1483 to perform the functions specified in §493.1485.

**Interpretive Guideline**

**FED - D6163 - CYTOTECHNOLOGIST QUALIFICATIONS**

**Title** CYTOTECHNOLOGIST QUALIFICATIONS

**CFR** 493.1483

**Type** Standard

**Regulation Definition**

Each person examining cytology slide preparations must meet the qualifications of §493.1449 (b) or (k).

**Interpretive Guideline**

**FED - D6164 - CYTOTECHNOLOGIST QUALIFICATIONS**

**Title** CYTOTECHNOLOGIST QUALIFICATIONS

**CFR** 493.1483

**Type** Standard

**Regulation Definition**

(a) Possess a current license as a cytotechnologist issued by the State in which the laboratory is located, if such licensing is

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required; and

(b) Meet one of the following requirements:

(b)(1) Have graduated from a school of cytotechnology accredited by the Committee on Allied Health Education and Accreditation or other organization approved by HHS; or

(b)(2) Be certified in cytotechnology by a certifying agency approved by HHS; or

(b)(3) Before September 1, 1992--

(b)(3)(i) Have successfully completed 2 years in an accredited institution with at least 12 semester hours in science, 8 hours of which are in biology; and

(b)(3)(i)(A) Have had 12 months of training in a school of cytotechnology accredited by an accrediting agency approved by HHS; or

(b)(3)(i)(B) Have received 6 months of formal training in a school of cytotechnology accredited by an accrediting agency approved by HHS and 6 months of full-time experience in cytotechnology in a laboratory acceptable to the pathologist who directed the formal 6 months of training; or

(b)(3)(ii) Have achieved a satisfactory grade to qualify as a cytotechnologist in a proficiency examination approved by HHS and designed to qualify persons as cytotechnologists; or

(b)(4) Before September 1, 1994, have full-time experience of at least 2 years or equivalent within the preceding 5 years examining slide preparations under the supervision of a physician qualified under §493.1449(b) or (k)(1), and before January 1, 1969, must have--

(b)(4)(i) Graduated from high school;

(b)(4)(ii) Completed 6 months of training in cytotechnology in a laboratory directed by a pathologist or other physician providing cytology services; and

(b)(4)(iii) Completed 2 years of full-time supervised experience in cytotechnology; or

(b)(5)(i) On or before September 1, 1994, have full-time experience of at least 2 years or equivalent examining

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cytology slide preparations within the preceding 5 years in the United States under the supervision of a physician qualified under §493.1449(b) or (k)(1); and  
(b)(5)(ii) On or before September 1, 1995, have met the requirements in either paragraph (b)(1) or (2) of this section.

**FED - D6165 - CYTOTECHNOLOGIST RESPONSIBILITIES**

**Title** CYTOTECHNOLOGIST RESPONSIBILITIES

**CFR** 493.1485(a)

**Type** Standard

**Regulation Definition**

The cytotechnologist is responsible for documenting the slide interpretation results of each gynecologic and nongynecologic cytology case he or she examined or reviewed (as specified in §493.1274(c)).

**Interpretive Guideline**

**FED - D6166 - CYTOTECHNOLOGIST RESPONSIBILITIES**

**Title** CYTOTECHNOLOGIST RESPONSIBILITIES

**CFR** 493.1485(b)

**Type** Standard

**Regulation Definition**

The cytotechnologist is responsible for documenting, for each 24-hour period, the total number of slides examined or reviewed in the laboratory as well as the total number of slides examined or reviewed in any other laboratory or for any other employer.

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**FED - D6167 - CYTOTECHNOLOGIST RESPONSIBILITIES**

**Title** CYTOTECHNOLOGIST RESPONSIBILITIES

**CFR** 493.1485(c)

**Type** Standard

**Regulation Definition**

The cytotechnologist is responsible for documenting the number of hours spent examining slides in each 24-hour period.

**Interpretive Guideline**

**FED - D6168 - TESTING PERSONNEL**

**Title** TESTING PERSONNEL

**CFR** 493.1487

**Type** Condition

**Regulation Definition**

The laboratory has a sufficient number of individuals who meet the qualification requirements of §493.1489 of this subpart to perform the functions specified in §493.1495 of this subpart for the volume and complexity of testing performed.

**Interpretive Guideline**

Interpretative Guidelines §493.1487

The Condition of Testing Personnel is not met when the testing personnel:

- o Are not qualified; or
- o Do not fulfill the testing personnel responsibilities.

The criteria used to determine the adequacy of the testing personnel involves evaluating testing personnel responsibilities, ensuring that these responsibilities are specified by the director in writing and are appropriate to ensure compliance with the reporting and recordkeeping requirements, quality control monitoring, quality assessment activities, and proficiency testing participation. Cite this deficiency only when problems are found in areas that can be directly related to insufficient numbers of testing personnel. (Use D6101 to relate the finding regarding insufficient personnel to director responsibilities.)

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**FED - D6170 - TESTING PERSONNEL QUALIFICATIONS**

**Title** TESTING PERSONNEL QUALIFICATIONS

**CFR** 493.1489(a)

**Type** Standard

**Regulation Definition**

Each individual performing high complexity testing must possess a current license issued by the State in which the laboratory is located, if such licensing is required.

**Interpretive Guideline**

**FED - D6171 - TESTING PERSONNEL QUALIFICATIONS**

**Title** TESTING PERSONNEL QUALIFICATIONS

**CFR** 493.1489(b)

**Type** Standard

**Regulation Definition**

- (b) Meet one of the following requirements:
- (b)(1) Be a doctor of medicine, doctor of osteopathy, or doctor of podiatric medicine licensed to practice medicine, osteopathy, or podiatry in the State in which the laboratory is located or have earned a doctoral, master's or bachelor's degree in a chemical, physical, biological or clinical laboratory science, or medical technology from an accredited institution;
- (b)(2)(i) Have earned an associate degree in a laboratory science, or medical laboratory technology from an accredited institution or--
- (b)(2)(ii) Have education and training equivalent to that specified in paragraph (b)(2)(i) of this section that includes--
- (b)(2)(ii)(A) At least 60 semester hours, or equivalent, from an accredited institution that, at a minimum, include either--

**Interpretive Guideline**

Interpretative Guidelines §493.1489(b)(1)

See §493.2 for the definition of an accredited institution.

Interpretative Guidelines §493.1489(b)(2)

"An associate degree in a laboratory science" is interpreted to mean an associate degree in a chemical or biological science.

Interpretative Guidelines §493.1489(b)(4)(ii)

Equate similar military courses with different titles. Evaluate the course length and content to ensure that it provides effective training for testing personnel. Refer to "A Guide to the Evaluation of Educational Experience in the Armed Services," American Council on Education, Washington, D.C.

Interpretative Guidelines §493.1489(b)(5)(ii)

The laboratory director is responsible for ensuring that testing personnel have the appropriate education and experience, and receive the appropriate training for the type and complexity of testing performed. The experience

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(b)(2)(ii)(A)(1) 24 semester hours of medical laboratory technology courses; or

(b)(2)(ii)(A)(2) 24 semester hours of science courses that include--

(b)(2)(ii)(A)(2)(i) Six semester hours of chemistry;

(b)(2)(ii)(A)(2)(ii) Six semester hours of biology; and

(b)(2)(ii)(A)(2)(iii) Twelve semester hours of chemistry, biology, or medical laboratory technology in any combination; and

(b)(2)(ii)(B) Have laboratory training that includes either of the following:

(b)(2)(ii)(B)(1) Completion of a clinical laboratory training program approved or accredited by the ABHES, the CAHEA, or other organization approved by HHS. (This training may be included in the 60 semester hours listed in paragraph (b)(2)(ii)(A) of this section.)

(b)(2)(ii)(B)(2) At least 3 months documented laboratory training in each specialty in which the individual performs high complexity testing.

(b)(3) Have previously qualified or could have qualified as a technologist under §493.1491 on or before February 28, 1992;

(b)(4) On or before April 24, 1995 be a high school graduate or equivalent and have either--

(b)(4)(i) Graduated from a medical laboratory or clinical laboratory training program approved or accredited by ABHES, CAHEA, or other organization approved by HHS; or

(b)(4)(ii) Successfully completed an official U.S. military medical laboratory procedures training course of at least 50 weeks duration and have held the military enlisted occupational specialty of Medical Laboratory Specialist (Laboratory Technician);

(b)(5)(i) Until September 1, 1997--

(b)(5)(i)(A) Have earned a high school diploma or

required is clinical in nature. This means examination of and test performance on human specimens for purposes of obtaining information for the diagnosis, treatment, and monitoring of patients, or for providing information to others who will do the diagnosing and treating of the patient's condition. (Use D6102.)

Each individual must have documentation of training applicable to the types and complexity of testing performed. This training should be such that the individual can demonstrate that he/she has the skills required for proper performance of preanalytic, analytic, and postanalytic phases of testing. For example, if the individual performs a manual differential, he/she should be able to demonstrate the skills for:

- o Proper specimen handling prior to testing, e.g., ensuring the specimen is properly drawn, if appropriate, properly labeled, the blood film is made within appropriate timeframes and is one-cell layer thick and without cell distortion;

- o Proper test performance according to the laboratory's policies and manufacturer's instructions, e.g., using stains that are not outdated, that lack contamination and precipitation, following staining procedures, including staining order and timing and allowing slide to air dry, identification of cells and interpretation of smear to be consistent with blood count, diagnosis, treatment; and

- o Proper reporting of patient test results in accordance with the laboratory's policies, e.g., notifying the person authorized to receive test results of a panic value, not reporting the test result if inconsistent with blood count and noting an explanation, such as "platelet clumping."

Training may include, but is not limited to, attendance at:

- o Seminars given by experts in the field, e.g., a lecture about antibiotic resistance given by the infection control officer of a local hospital;

- o On-site or off-site instrument trainings given by a manufacturer, e.g., a week-long training course given at the manufacturer's headquarters, or training by a manufacturer's technical representative on an instrument purchased by a laboratory;

- o Technical training sessions, workshops, or conferences given by a professional laboratory organization, e.g., CAP, ASMT, AACC, and ASCT;

- o Technical education classes or specialty courses that include hands-on test performance, e.g., parasitology, bacteriology, cytology, given by CDC, a State Health Department, or professional laboratory organizations;

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equivalent; and

(b)(5)(i)(B) Have documentation of training appropriate for the testing performed before analyzing patient specimens.

Such training must ensure that the individual has--

(b)(5)(i)(B)(1) The skills required for proper specimen collection, including patient preparation, if applicable, labeling, handling, preservation or fixation, processing or preparation, transportation and storage of specimens;

(b)(5)(i)(B)(2) The skills required for implementing all standard laboratory procedures;

(b)(5)(i)(B)(3) The skills required for performing each test method and for proper instrument use;

(b)(5)(i)(B)(4) The skills required for performing preventive maintenance, troubleshooting, and calibration procedures related to each test performed;

(b)(5)(i)(B)(5) A working knowledge of reagent stability and storage;

(b)(5)(i)(B)(6) The skills required to implement the quality control policies and procedures of the laboratory;

(b)(5)(i)(B)(7) An awareness of the factors that influence test results; and

(b)(5)(i)(B)(8) The skills required to assess and verify the validity of patient test results through the evaluation of quality control values before reporting patient test results; and

(b)(5)(i)(B)(8)(ii) As of September 1, 1997, be qualified under §493.1489(b)(1), (b)(2), or (b)(4), except for those individuals qualified under paragraph (b)(5)(i) of this section who were performing high complexity testing on or before April 24, 1995;

(b)(6) For blood gas analysis--

(b)(6)(i) Be qualified under §493.1489(b)(1), (b)(2), (b)(3), (b)(4), or (b)(5);

(b)(6)(ii) Have earned a bachelor's degree in respiratory therapy or cardiovascular technology from an accredited institution; or

o A formal laboratory training program; or

o In-services offered by a local hospital laboratory staff, pathologist, or medical technologist to a physician's office personnel.

Documentation may consist of, but is not limited to, letters from training programs or employers, attestation statements by the laboratory director, a log sheet initialed by the attendees indicating attendance at a training session/in-service, certificates from organizations providing the training session, workshop, conference, or specialty course.

Interpretative Guidelines §493.1489(b)(6)

This requirement applies only to performance of blood gas analysis procedures which are categorized as high complexity.

NOTE: Some blood gas systems are categorized as moderate complexity tests. Therefore, only moderate complexity personnel requirements are applicable to them. To determine which tests are categorized as waived or nonwaived (i.e., moderate or high complexity tests), refer to the "Specific List For Categorization of Laboratory Test Systems, Assays, and Examinations by Complexity" (<<http://www.gpo.gov/fdsys/pkg/FR-1995-05-15/pdf/95-11653.pdf>>). Test systems, assays, and examinations not yet classified are considered high complexity.

Interpretative Guidelines §493.1489(b)(7)

The tests in histopathology include both gross examination (macroscopic), and microscopic examination of the slide(s) with evaluation and diagnostic interpretation, and diagnostic findings reported.

In the case of gross examinations, the technical supervisor may delegate to individuals qualified under §493.1489 the responsibility for the physical examination/description, including color, weight, measurement and other characteristics of the tissue; or other mechanical procedures for which a specific written protocol has been developed. The technical supervisor is ultimately responsible for the diagnosis related to the gross examination and must sign the examination report. The technical supervisor is not required to provide direct on-site supervision but is responsible for the accuracy of all test results reported. All physical examinations/descriptions of tissue including color, weight, measurement and other characteristics of the tissue; or other mechanical procedures performed in the absence of the technical supervisor by individuals qualified under §493.1489 should be reviewed within 24 hours by the technical supervisor. All microscopic tissue examinations must be performed by individuals qualified under §493.1449(b), (l) or (m), as appropriate.

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(b)(6)(iii) Have earned an associate degree related to pulmonary function from an accredited institution; or  
(b)(7) For histopathology, meet the qualifications of §493.1449 (b) or (l) to perform tissue examinations.

**FED - D6173 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1495

**Type** Standard

**Regulation Definition**

The testing personnel are responsible for specimen processing, test performance and for reporting test results.

**Interpretive Guideline**

Interpretative Guidelines §493.1495

The tests in histopathology include gross examination (macro), microscopic slide evaluation and interpretation with diagnostic reporting.

**FED - D6174 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1495(a)

**Type** Standard

**Regulation Definition**

Each individual performs only those high complexity tests that are authorized by the laboratory director and require a degree of skill commensurate with the individual's education, training or experience, and technical abilities.

**Interpretive Guideline**

**FED - D6175 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1495(b)(1)

**Type** Standard

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**Regulation Definition**

Each individual performing high complexity testing must follow the laboratory's procedures for specimen handling and processing, test analyses, reporting and maintaining records of patient test results.

**Interpretive Guideline**

**FED - D6176 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1495(b)(2)

**Type** Standard

**Regulation Definition**

Each individual performing high complexity testing must maintain records that demonstrate that proficiency testing samples are tested in the same manner as patient specimens.

**Interpretive Guideline**

**FED - D6177 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1495(b)(3)

**Type** Standard

**Regulation Definition**

Each individual performing high complexity testing must adhere to the laboratory's quality control policies, document all quality control activities, instrument and procedural calibrations and maintenance performed.

**Interpretive Guideline**

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**FED - D6178 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1495(b)(4)

**Type** Standard

**Regulation Definition**

Each individual performing high complexity testing must follow the laboratory's established policies and procedures whenever test systems are not within the laboratory's established acceptable levels of performance.

**Interpretive Guideline**

**FED - D6179 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1495(b)(5)

**Type** Standard

**Regulation Definition**

Each individual performing high complexity testing must be capable of identifying problems that may adversely affect test performance or reporting of test results and either must correct the problems or immediately notify the general supervisor, clinical consultant, or director.

**Interpretive Guideline**

Interpretative Guidelines §493.1495(b)(5) Guidelines:

If, during the survey, testing personnel demonstrate an inability to identify a problem that adversely affects a patient test result, cite 493.1445(e)(12) under the director responsibilities.

Some examples of problems that may adversely affect patient test results may include:

- o A pleural fluid that is mislabeled as a urine specimen and, therefore, is cultured as a urine culture;
- o Performing a potassium on a hemolyzed sample; or
- o Tests are incubated at 37°C when the manufacturer's instructions require 25°C incubation.

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**FED - D6181 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1495(b)(6)

**Type** Standard

**Regulation Definition**

Each individual performing high complexity testing must document all corrective actions taken when test systems deviate from the laboratory's established performance specifications.

**Interpretive Guideline**

**FED - D6182 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1495(b)(7)

**Type** Standard

**Regulation Definition**

Each individual performing high complexity testing must, except as specified in paragraph (c) of this section, if qualified under §493.1489(b)(5), perform high complexity testing only under the onsite, direct supervision of a general supervisor qualified under §493.1461.

**Interpretive Guideline**

**FED - D6183 - TESTING PERSONNEL RESPONSIBILITIES**

**Title** TESTING PERSONNEL RESPONSIBILITIES

**CFR** 493.1495(c)

**Type** Standard

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**Regulation Definition**

Exception. For individuals qualified under 493.1489(b)(4) who were performing high complexity testing on or before January 19, 1993, the requirements of paragraph (b)(7) of this section are not effective, provided that all high complexity testing performed by the individual in the absence of a general supervisor is reviewed within 24 hours by a general supervisor qualified under 493.1461.

**Interpretive Guideline**

**FED - D8100 - INSPECTION REQUIREMENTS**

**Title** INSPECTION REQUIREMENTS

**CFR** 493.1771

**Type** Condition

**Regulation Definition**

Each laboratory issued a CLIA certificate must meet the requirements in §493.1773 and the specific requirements for its certificate type, as specified in §§493.1775 through 493.1780. All CLIA-exempt laboratories must comply with the inspection requirements in §§493.1773 and 493.1780, when applicable.

**Interpretive Guideline**

**FED - D8101 - BASIC INSPECTION REQUIREMENTS**

**Title** BASIC INSPECTION REQUIREMENTS

**CFR** 493.1773(a)

**Type** Standard

**Regulation Definition**

A laboratory issued a certificate must permit CMS or a CMS agent to conduct an inspection to assess the laboratory's compliance with the requirements of this part. A CLIA-exempt

**Interpretive Guideline**

Interpretative Guidelines §493.1773(a)

If for any reason a facility denies entry to or does not permit you to conduct a survey, the following steps should be taken:

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laboratory and a laboratory that requests, or is issued a certificate of accreditation, must permit CMS or a CMS agent to conduct validation and complaint inspections.

- o Explain your authority to conduct the survey and the consequences of failure to permit a survey;
- o If necessary, consult with your supervisor or the RO; and
- o For failure to permit entry into or inspection of the laboratory, use D8101.

If the laboratory continues to refuse a survey, refer to Subpart R - Enforcement Procedures beginning at §493.1800 and the Adverse Action section of the SOM at 6250.

Conduct complaint surveys on an unannounced basis.

The CLIA application will solicit the laboratory's hours of operation. For complaint or revisit surveys, you may phone the laboratory to confirm the hours of testing prior to a survey without revealing your identity or the scheduled date.

Make every effort to minimize the impact of the survey on the laboratory operations and patient care activities. Be flexible; accommodate staffing schedules and workloads as much as possible. In facilities providing direct patient care, e.g., physician's offices, clinics, residential care facilities, hospitals, respect patient privacy and do not interrupt or interfere with patient care. Be well prepared, courteous and make requests, not demands.

Maintain documentation for all on-site follow-up surveys in the laboratory's official file.

**FED - D8103 - BASIC INSPECTION REQUIREMENTS**

**Title** BASIC INSPECTION REQUIREMENTS

**CFR** 493.1773(b)(c)(d)

**Type** Standard

**Regulation Definition**

(b) General Requirements. As part of the inspection process, CMS or a CMS agent may require the laboratory to do the following:

- (b)(1) Test samples, including proficiency testing samples, or perform procedures.
- (b)(2) Permit interviews of all personnel concerning the

**Interpretive Guideline**

Interpretative Guidelines §493.1773(b)-(c)

The regulations do not require a laboratory to maintain records on-site. During the survey, the laboratory must be able to retrieve copies of all records and necessary information upon request. Determine what constitutes a reasonable timeframe based on the information requested.

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laboratory's compliance with the applicable requirements of this part.

(b)(3) Permit laboratory personnel to be observed performing all phases of the total testing process preanalytic, analytic, and postanalytic).

(b)(4) Permit CMS or a CMS agent access to all areas encompassed under the certificate including, but not limited to, the following:

(b)(4)(i) Specimen procurement and processing areas.

(b)(4)(ii) Storage facilities for specimens, reagents, supplies, records, and reports.

(b)(4)(iii) Testing and reporting areas.

(b)(5) Provide CMS or a CMS agent with copies or exact duplicates of all records and data it requires.

(c) Accessible records and data. A laboratory must have all records and data accessible and retrievable within a reasonable time frame during the course of the inspection.

(d) Requirement to provide information and data. A laboratory must provide, upon request, all information and data needed by CMS or a CMS agent to make a determination of the laboratory's compliance with the applicable requirements of this part.

**FED - D8105 - BASIC INSPECTION REQUIREMENTS**

**Title** BASIC INSPECTION REQUIREMENTS

**CFR** 493.1773(e)(f)(g)

**Type** Standard

**Regulation Definition**

(e) Reinspection. CMS or a CMS agent may reinspect a laboratory at any time to evaluate the ability of the laboratory to provide accurate and reliable test results.

(f) Complaint inspection. CMS or a CMS agent may conduct an inspection when there are complaints alleging noncompliance with any of the requirements of this part.

**Interpretive Guideline**

Interpretative Guidelines §493.1773(e-g)

If for any reason a facility denies entry to or does not permit you to conduct a survey, the following steps should be taken:

- o Explain your authority to conduct the survey and the consequences of failure to permit a survey;

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(g) Failure to permit CMS or a CMS agent to conduct an inspection or reinspection results in the suspension or cancellation of the laboratory's participation in Medicare and Medicaid for payment, and suspension or limitation of, or action to revoke the laboratory's CLIA certificate, in accordance with subpart R of this part.

- o If necessary, consult with your supervisor or the RO; and
- o For failure to permit entry into or inspection of the laboratory, use D8101.

If the laboratory continues to refuse a survey, refer to Subpart R - Enforcement Procedures beginning at §493.1800 and the Adverse Action section of the SOM at 6250.

Conduct complaint surveys on an unannounced basis.

The CLIA application will solicit the laboratory's hours of operation. For complaint or revisit surveys, you may phone the laboratory to confirm the hours of testing prior to a survey without revealing your identity or the scheduled date.

Make every effort to minimize the impact of the survey on the laboratory operations and patient care activities. Be flexible, accommodate staffing schedules and workloads as much as possible. In facilities providing direct patient care, e.g., physician's offices, clinics, residential care facilities, hospitals, respect patient privacy and do not interrupt or interfere with patient care. Be well prepared, courteous and make requests, not demands.

Maintain documentation for all on-site follow-up surveys in the laboratory's official file.

**FED - D8201 - INSPECTION OF COW OR PPMP LABS**

**Title** INSPECTION OF COW OR PPMP LABS

**CFR** 493.1775(b)

**Type** Standard

**Regulation Definition**

(b) If necessary, CMS or a CMS agent may conduct an inspection of a laboratory issued a certificate of waiver or a certificate for provider-performed microscopy procedures at anytime during the laboratory's hours of operation to do the following:

(b)(1) Determine if the laboratory is operated and testing is performed in a manner that does not constitute an imminent and serious risk to public health.

(b)(2) Evaluate a complaint from the public.

**Interpretive Guideline**

Interpretative Guidelines §493.1775(b)

In any laboratory holding a CLIA certificate, tests listed on the waived list are not subject to routine surveys. A survey for waived tests may be conducted only when authorized by the RO in one of the following instances:

- o To collect information on waived tests;
- o To determine whether the laboratory is testing beyond its certificate;
- o If a complaint is alleged; or

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(b)(3) Determine whether the laboratory is performing tests beyond the scope of the certificate held by the laboratory.

(b)(4) Collect information regarding the appropriateness of tests specified as waived tests or provider-performed microscopy procedures.

o You have information that the performance of such tests poses an imminent and serious risk that adversely affects patient test results.

When authorized to perform a survey of waived tests, in addition to the requirements in this subpart, refer to the requirements at §493.15, subpart A, and §§493.35, 493.37 and 493.39, subpart B, of these guidelines.

Section 493.35(d) requires that laboratories performing only waived tests and no other tests must agree to permit inspections by HHS in order to receive a certificate of waiver.

Make every effort to minimize the impact of the survey on the laboratory operations and patient care activities. Be flexible, accommodate staffing schedules and workloads as much as possible. In facilities providing direct patient care, (i.e., physician's offices, clinics, residential care facilities, hospitals, etc.), respect patient privacy and do not interrupt or interfere with patient care. Be well prepared, courteous and make requests, not demands.

Interpretative Guidelines §493.1775(b)(3)

When a laboratory has failed to obtain a registration certificate before performing and reporting patient results for nonwaived testing, notify the RO of a possible action by the Office of the Inspector General (OIG) if the laboratory does not obtain the appropriate certificate or cease the nonwaived testing.

**FED - D8203 - INSPECTION OF COW OR PPMP LABS**

**Title** INSPECTION OF COW OR PPMP LABS

**CFR** 493.1775(c)

**Type** Standard

**Regulation Definition**

The laboratory must comply with the basic inspection requirements of §493.1773.

**Interpretive Guideline**

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**FED - D8301 - INITIAL INSPECTION OF CERT OF COMPLIANCE LABS**

**Title** INITIAL INSPECTION OF CERT OF COMPLIANCE LABS

**CFR** 493.1777(a)

**Type** Standard

**Regulation Definition**

(a) Initial inspection.

(a)(1) A laboratory issued a registration certificate must permit an initial inspection to assess the laboratory's compliance with the requirements of this part before CMS issues a certificate of compliance.

(a)(2) The inspection may occur at any time during the laboratory's hours of operation.

**Interpretive Guideline**

Interpretative Guidelines §493.1777(a)

If for any reason a facility denies entry to or does not permit you to conduct a survey, take the following steps:

- o Explain your authority to conduct the survey and the consequences of failure to permit a survey;
- o If necessary, consult with your supervisor or the RO; and
- o For failure to permit entry into or an inspection of the laboratory, use D8101.

If the laboratory continues to refuse a survey, refer to Subpart R - Enforcement Procedures beginning at §493.1800 and the Adverse Action section of the SOM at 6250.

**FED - D8303 - SUBSEQUENT INSPECTIONS OF CERT OF COMPL LABS**

**Title** SUBSEQUENT INSPECTIONS OF CERT OF COMPL LABS

**CFR** 493.1777(b)

**Type** Standard

**Regulation Definition**

(b) Subsequent inspections.

(b)(1) CMS or a CMS agent may conduct subsequent inspections on a biennial basis or with such other frequency as CMS determines to be necessary to ensure compliance with the requirements of this part.

(b)(2) CMS bases the nature of subsequent inspections on the laboratory's compliance history.

**Interpretive Guideline**

Interpretative Guidelines §493.1777(b)

In any laboratory holding a CLIA certificate, tests listed on the waived list are not subject to routine surveys. A survey for waived tests may be conducted only when authorized by the RO in one of the following instances:

- o To collect information on waived tests;
- o To determine whether the laboratory is testing beyond its certificate;

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- o If a complaint is alleged; or
- o You have information that the performance of such tests poses an imminent and serious risk that adversely affects patient test results.

When authorized to perform a survey of waived tests, in addition to the requirements in this subpart, refer to the requirements at §493.15, subpart A, and §§493.35, 493.37 and 493.39, subpart B, of these guidelines. Section 493.35(d) requires that laboratories performing only waived tests and no other tests must agree to permit inspections by HHS in order to receive a certificate of waiver.

Make every effort to minimize the impact of the survey on the laboratory operations and patient care activities. Be flexible; accommodate staffing schedules and workloads as much as possible. In facilities providing direct patient care, (i.e., physician's offices, clinics, residential care facilities, hospitals, etc.), respect patient privacy and do not interrupt or interfere with patient care. Be well prepared, courteous and make requests, not demands.

**FED - D8305 - PROVIDER-PERFORMED MICROSCOPY PROCEDURES**

**Title** PROVIDER-PERFORMED MICROSCOPY PROCEDURES

**CFR** 493.1777(c)

**Type** Standard

**Regulation Definition**

The inspection sample for review may include testing in the subcategory of provider-performed microscopy procedures.

**Interpretive Guideline**

**FED - D8307 - COMPLIANCE WITH BASIC INSPECTION REQUIREMENTS**

**Title** COMPLIANCE WITH BASIC INSPECTION REQUIREMENTS

**CFR** 493.1777(d)

**Type** Standard

**Regulation Definition**

The laboratory must comply with the basic inspection

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requirements of Sec. 493.1773.

**FED - D8401 - INSPECTION OF CLIA-EXEMPT AND ACCREDITED LABS**

**Title** INSPECTION OF CLIA-EXEMPT AND ACCREDITED LABS

**CFR** 493.1780

**Type** Standard

**Regulation Definition**

- (a) Validation inspection. CMS or a CMS agent may conduct a validation inspection of any accredited or CLIA-exempt laboratory at any time during its hours of operation.
- (b) Complaint inspection. CMS or a CMS agent may conduct a complaint inspection of a CLIA-exempt laboratory or a laboratory requesting or issued a certificate of accreditation at any time during its hours of operation upon receiving a complaint applicable to the requirements of this part.
- (c) Noncompliance determination. If a validation or complaint inspection results in a finding that the laboratory is not in compliance with one or more condition-level requirements, the following actions occur:
- (c)(1) A laboratory issued a certificate of accreditation is subject to a full review by CMS, in accordance with subpart E of this part and §488.11 of this chapter.
- (c)(2) A CLIA-exempt laboratory is subject to appropriate enforcement actions under the approved State licensure program.
- (d) Compliance with basic inspection requirements. CLIA-exempt laboratories and laboratories requesting or issued a certificate of accreditation must comply with the basic inspection requirements in §493.1773.

**Interpretive Guideline**

Interpretative Guidelines §493.1780

Validation surveys of accredited laboratories will be conducted by the State survey agencies. Refer to special procedures for accredited laboratories in the SOM. The RO is responsible for conducting validations of CLIA-exempt laboratories.

Interpretative Guidelines §493.1780(b)

In any laboratory holding a CLIA certificate, tests listed on the waived list are not subject to routine surveys. A survey for waived tests may be conducted only when authorized by the RO in one of the following instances:

- o To collect information on waived tests;
- o To determine whether the laboratory is testing beyond its certificate;
- o If a complaint is alleged; or
- o You have information that the performance of such tests poses an imminent and serious risk that adversely affects patient test results.

When authorized to perform a survey of waived tests, in addition to the requirements in this subpart, refer to the requirements at §493.15, subpart A, and §§493.35, 493.37 and 493.39, subpart B, of these guidelines.

Section 493.35(d) requires that laboratories performing only waived tests and no other tests must agree to permit inspections by HHS in order to receive a certificate of waiver.

Make every effort to minimize the impact of the survey on the laboratory operations and patient care activities. Be flexible, accommodate staffing schedules and workloads as much as possible. In facilities providing direct patient care, (i.e., physician's offices, clinics, residential care facilities, hospitals, etc.), respect patient privacy and do not interrupt or interfere with patient care. Be well prepared, courteous and make requests, not demands.

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**FED - D9999 - CLOSING COMMENTS**

**Title** CLOSING COMMENTS

**CFR**

**Type** Memo Tag

**Regulation Definition**

**Interpretive Guideline**